

<b>Virus Name: Cocal</b>		<b>Abbreviation: COCV</b>
Status <b>Arbovirus</b>	Select Agent <b>No</b>	SALS Level <b>3</b>
SALS Basis <b>Disease is sheep, cattle, or horses.</b>		
Other Information		
Antigenic Group <b>Vesicular Stomatitis</b>		

**SECTION I - Full Virus Name and Prototype Number**

Prototype Strain Number / Designation <b>TRVL 40233</b>	Accession Number	Original Date Submitted <b>2/24/1985</b>
Family <b>Rhabdovirus</b>	Genus <b>Vesiculovirus</b>	
Information From <b>Trinidad Regional Virus Laboratory</b>	Address <b>P.O. Box 164, Port of Spain, Trinidad</b>	
Information Footnote <b>Reviewed by editor</b>		

**Section II - Original Source**

Isolated By (name) <b>TRVL (1)</b>	Isolated at Institute <b>Port of Spain, Trinidad</b>	
Host Genus <b>Gigantolaelaps sp. (249 mites)</b>	Species	Host Age/Stage <b>Ad + immature</b>
Sex <b>Not Answered</b>		
<u>Isolated From</u>	<u>Isolation Details</u>	
Signs and Symptoms of Illness	Arthropod	
Time Held Alive before Inoculation		
Collection Method <b>Off of 11 trapped Oryzomys laticeps velutinus</b>	Collection Date <b>9/25/1961</b>	
Place Collected (Minimum of City, State, Country) <b>Nariva County, Trinidad</b>		
Latitude <b>10° 24' N</b>	Longitude <b>61° 3' W</b>	
Macrohabitat <b>Bush Bush Forest, Nariva Swamp, Eastern Trinidad</b>	Microhabitat <b>Semi-evergreen seasonal forest</b>	Method of Storage until Inoculated <b>Stored while alive in Revco at -55dC until ground and inoculated</b>
Footnotes		

**Section III - Method of Isolation**

Inoculation Date  
**10/30/1961**

Animal (Details will be in Section 6)  
**nb mice**

Route Inoculated <b>Intracerebral</b>	Reisolation <b>Yes</b>
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Other Reasons  
**No virus of this type previously encountered**

Homologous Antibody Formation by Source Animal

Test(s) Used

Footnotes

**Section IV - Virus Properties**

Physicochemical

Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

**Stability of Infectivity (effects)**

pH (infective range)

Lipid Solvent (ether - % used to test)	After Treatment Titer <b>2.6 dex</b>	Control Titer <b>&gt;7.0 dex</b>
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate)	After Treatment Titer	Control Titer
Other (formalin, radiation)		

**Virion Morphology**

Shape <b>Identical to VSI (9)</b>	Dimensions <b>60 x 200 nm</b>	
Mean nm	Range nm	
Measurement Method <b>Electron microscopy (2)</b>	Surface Projections/Envelope	Nucleocapsid Dimensions, Symmetry

**Morphogenesis**

Site of Constituent Formation in Cell                      Site of Virion Assembly                      Site of Virion Accumulation

Inclusion Bodies                      Other

**Hemagglutination**

Hemagglutination                      Antigen Source                      Erythrocytes (species used)

**No**

pH Range                      pH Optimum

Temperature Range                      Temperature Optimum

Remarks

**Morphologically indistinguishable from VSV (9)**

Serologic Methods Recommended

**CF and NT**

Footnotes

**Morphologically indistinguishable from VSV (9)**

**Section V - Antigenic Relationship and Lack of Relationship to Other Viruses**

Cocal virus has been shown by both CF and NT to be related to but not identical with the prototype strain of VS-Indiana [1] . The same tests failed to demonstrate relationship to VS-New Jersey. It appears to be identical with BeAr 39377 isolated in Belem, Brazil. For relationship to other strains of VS-Indiana see registration card.

Because of its antigenic relationship and similar size and shape to VS- Indiana, it has been placed in the VSV group [3] . As far as it is known, it is not related to any viruses not included in this group. For a list of those with which it has been compared, see Reference [1] .

**Section VI - Biologic Characteristics**

Virus Source (all VERTEBRATE isolates)  
**CNS, heart, liver, spleen, and kidney pool (LV)**

Lab Methods of Virus Recovery (ALL ISOLATIONS)  
**Newborn mice**

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TC50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
HeLa (CL)	MB 5		CPE	7.5** (8)				
KB cells(CL)	SMB 6	24 hours	CPE	5.5				
Chick embryo(PC)		24 hours	CPE	6.5				
BHK-21 (CL)						Plaques (7)		
Monkey kidney(PC)						Plaques (7)		

\*\* Expressed in dex

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Man		0/75 NT	Trinidad
Equines	1		Argentina (11)
Equines		8/114 NT	Trinidad
Pigs		0/30 NT	
Sentinel mice	1/1700 litters		Bush Bush Forest Nariwa Swamp Trinidad
Heteromys anomalus	1/400	35/273 NT	
Oryzomys spp.		72/256 NT	
Zygodontomys brevicauda		58/313 NT	
Culex (Mel) portesi	1/23000		Bush Bush Forest Nariwa Swamp Trinidad (6)
Gigantolaelaps sp.	1/4000		
Gigantolaelaps sp.	1		Belem Brazil (1 4)
Ameiva ameiva atrigularis		0/30 NT	Bush Bush Forest

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log <sub>10</sub> /ml
Mice (nb)		ic 0.02	Death	2-3	7.0
Mice (nb)		ip 0.03	Death	3-5	6.2
Mice (nb)		sc			
Mice (wn)		ic 0.03	Death	3-5	7.0
Mice (wn)		ip 0.2	Death	4-10	3.2
embryonated eggs		al.c. 0.1	Death		7.5
		am.s. 0.1	Death		8.2
		ys 0.5	Death		8.2
guinea pigs			Vesicular lesions on all		
		7 dex LD50	four foot pads; antibodies		
Zygodontomys	MB 8	sc,ip,in	Paralysis, virus recovery		
Myotis l.lucifugus		sc	Long-term viremia (10)		

**Section IX - Experimental Arthropod Infection and Transmission**

Arthropod species & virus source(a)	Method of Infection log10/ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log10/ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System
Culex quinquefasciatus			Infected by inoc., 5 serial passages of sg virus and transmission by bite (1).						
Trichoprosopon digitatum			Infected by inoc., 2 serial passages of sg virus and transmission by bite (1).						
Anopheles quadrimaculatus, Culex quinquefasciatus and Aedes aegypti: virus multiplication and serially maintained by inoculation. (3)									
Aedes aegypti which were allowed to feed on infected bats in turn transmitted virus to suckling mice (10).									

**Section X - Histopathology**

Character of lesions (specify host)  
**Mice: distinctive, with destruction of cells of ependymal canal of spinal medulla, and pronounced karyorrhexis (L.B.Dias)**

Inclusion Bodies Intranuclear

Organs/Tissues Affected

Category of tropism

**Section XI - Human Disease**

In Nature Residual Death

Subclinical Overt Disease

Clinical Manifestations

Number of Cases Category (i.e. febrile illness, etc.)

**Section XII - Geographic Distribution**

Known (Virus detected)  
**Trinidad, Brazil, Argentina**

Suspected (Antibody only detected)

### Section XIII - References

1. Jonkers, A.H., et al. 1964. *Am. J. Vet. Res.* 25:236-242.
2. Bergold, G. Personal communication.
3. Rockefeller Foundation Virus Laboratory, N.Y. 1962. Unpublished observations.
4. Woodall, J.P. 1967. *Atas Simpos. Biota Amazon* 6:31-63.
5. Jonkers, A.H., et al. 1964. *Am. J. Trop. Med. Hyg.* 13:613-619.
6. Jonkers, A.H., et al. 1965. *Am. J. Vet. Research* 26:758-763.
7. Pinheiro, F.p. Personal communication.
8. Buckley, S.M. 1964. *Proc. Soc. Exp. Biol. Med.* 116:354-358.
9. Ditchfield, J. and Almeida, J.D. 1964. *Virology* 24:232-235.
10. Donaldson, A.I. 1970. *Am. J. Epidem.* 92:132-136.
11. Federer, K.E., et al. 1967. *Res. Vet. Sci.* 8:103-117.

### Remarks