

Virus Name: Entebbe bat		Abbreviation: ENTV
Status Possible Arbovirus	Select Agent No	SALS Level 3
SALS Basis Insufficient experience with virus; i.e., experience factor from SALS surveys was less than 500 in laboratory facilities with low biocontainment.		
Other Information		
Antigenic Group Ungrouped		

SECTION I - Full Virus Name and Prototype Number

Prototype Strain Number / Designation IL-30	Accession Number	Original Date Submitted 2/3/1985
Family Flaviviridae	Genus Flavivirius	
Information From J.P. Woodall	Address YARU, Yale University School of Medicine, New Haven, Connecticut 06510, USA	
Information Footnote Reviewed by editor		

Section II - Original Source

Isolated By (name) Lumsden, et al. (1)	Isolated at Institute Entebbe, Uganda	
Host Genus Bat (Tadarida (Chaerephon) limbata); (pool of 40)	Species	Host Age/Stage Ad, juvenile
Sex Male		
<u>Isolated From</u>	<u>Isolation Details</u>	
Organs/Tissues	Pool of submaxillary and greater sublingual glands	
Signs and Symptoms of Illness None	Arthropod	
Time Held Alive before Inoculation		
Collection Method Hand captured	Collection Date 7/4/1957	
Place Collected (Minimum of City, State, Country) Entebbe, Uganda		
Latitude 0° 3' N	Longitude 32° 27' E	
Macrohabitat Lakeshore with small-holdings and swamp forest	Microhabitat Roof of East African Virus Research Institute	Method of Storage until Inoculated 4dC
Footnotes		

Section III - Method of Isolation

Inoculation Date	
Animal (Details will be in Section 6) nb mice	
Route Inoculated ic and ip	Reisolation No
Other Reasons Different from all other arboviruses and mouse viruses in laboratory	
Homologous Antibody Formation by <u>Source Animal</u> Not tested	
Test(s) Used	
Footnotes	

Section IV - Virus Properties

Physicochemical		
Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	
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<u>Stability of Infectivity (effects)</u>		
pH (infective range)		
Lipid Solvent (ether - % used to test) 1:1	After Treatment Titer 3:1 dex	Control Titer 7.0 dex
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate)	After Treatment Titer	Control Titer
Other (formalin, radiation)		
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<u>Virion Morphology</u>		
Shape	Dimensions	
Mean nm	Range nm	
Measurement Method	Surface Projections/Envelope	Nucleocapsid Dimensions, Symmetry

Morphogenesis

Site of Constituent Formation in Cell	Site of Virion Assembly	Site of Virion Accumulation
Inclusion Bodies	Other	

Hemagglutination

Hemagglutination Yes	Antigen Source SMB ext. by acetone-ether; sucrose-acetone + prot; fluorocarbon	Erythrocytes (species used) Goose
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pH Range 6.2-7.0	pH Optimum 6.6-6.8
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Temperature Range Room temperature	Temperature Optimum
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Remarks
Some preparations are unstable on overnight incubation in the cold

Serologic Methods Recommended
HI, CF, NT

Footnotes
Some preparations are unstable on overnight incubation in the cold

For list of antisera tested by NT, see Reference [1]; only Group B antisera gave any cross-reaction. Easily separable from Rio Bravo by NT [1].

Unpublished HI results are:

Antisera to	Antigen						
	ENT	Ntaya	WN	UGS	YF	Zika	Banzi
ENT	5*	5	4	0	0	0	1
Ntaya	10+	10	10	6	7	4	5
West Nile	7	6	8	3	2	1	3
Uganda S	5	5	5	6	1	0	4
Yellow fever	4	3	3	0	4	0	0
Zika	1	4	3	0	0	8	2

Antisera	Antigen		
	ENT	Dakar bat	BP 111
ENT	4	0	0
MML	7	4	2
Dakar bat	6	8	2
BP 111	3	3	9

* Number of dilutions with HI activity; initial dilution of 1:10 = 1, 1:20 = 2, etc.; 0 = <10.

References: BP 111 [2], Dakar bat [3], MML [4]. For further information on antigenic relationships, see References [8] and [9].

Section VI - Biologic Characteristics

Virus Source (all VERTEBRATE isolates)
Salivary gland (LV)

Lab Methods of Virus Recovery (ALL ISOLATIONS)
Newborn mice

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
BHK-21 (CL)	MB 2	5-7	2+ - 3+	7.5** (6)				
Vero (CL)	P-2				16	2 mm	6.8** (7)	
LLC-MK2 (CL)					4	2 mm	8.4 (7)	

** Expressed in dex

Section VII - Natural Host Range (Additional text can be added below table)

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Man		0/20 NT	Uganda
Tadarida (C.) limbata (syn. T. (C.) pumila)	1/13 pools (335 bats)	46/49 HI	Entebbe, Uganda (5)
T. (Mops) condylura		13/56 HI	Uganda (5)
Bats		1/20 NT	Uganda

Section VIII - Susceptibility to Experimental Infection (include viremia)

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log ₁₀ /ml
Mice (nb)		ic 0.02	Death	4	9.5
Mice (nb)		ip 0.02	Death	5	9.5
Mice (nb)		sc			
Mice (wn)		ic 0.03	Paralysis and death	9	8.5
Mice (wn)		ip 0.1	Antibody		

Resistance of mice to ip inoc. begins to appear between day 14 and 16 of life and is almost complete by day 20.

Section IX - Experimental Arthropod Infection and Transmission

Arthropod species & virus source(a)	Method of Infection log ₁₀ /ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log ₁₀ /ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System

Section X - Histopathology

Character of lesions (specify host)

Adult mice: gross disorganization lateral part of hippocampus; no marked cellular infiltration; no inclusion bodies.

Inclusion Bodies

Intranuclear

Organs/Tissues Affected

Brain (LV)

Category of tropism

Section XI - Human Disease

In Nature

Residual

Death

Subclinical

Overt Disease

Clinical Manifestations

Number of Cases

Category (i.e. febrile illness, etc.)

Section XII - Geographic Distribution

Known (Virus detected)

Uganda

Suspected (Antibody only detected)

Section XIII - References

1. Lumsden, W.H.R., et al. 1961. Ann. Trop. Med. Parasit. 55:389-97.
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3. Bres, P. and Chambon, L. 1963. Ann. Inst. Pasteur 104:705-711.
4. Bell, J.F. and Thomas, L.A. 1964. Am. J. Trop. Med. Hyg. 13:607-612.
5. Shepherd, C. and Williams, M.C. 1964. Zoonoses Res. 3:125-139.
6. Karabatsos, N. and Buckley, S.M. 1967. Am. J. Trop. Med. Hyg. 16:99-105.
7. Stim, T.B. 1969. J. Gen. Virol. 5:329-338.
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9. De Madrid, A.T. and Porterfield, J.S. 1974. J. Gen. Virol. 23:91-96.

Remarks