

Virus Name: Eubenangee		Abbreviation: EUBV
Status Possible Arbovirus	Select Agent No	SALS Level 2
SALS Basis Results of SALS surveys and information from the Catalogue.		
Other Information		
Antigenic Group Eubenangee		

SECTION I - Full Virus Name and Prototype Number

Prototype Strain Number / Designation IN1074 (4)	Accession Number	Original Date Submitted 10/25/1984
Family Reoviridae	Genus Orbivirus	
Information From R.L. Doherty	Address Queensland Institute of Medical Research, Brisbane	
Information Footnote Reviewed by editor		

Section II - Original Source

Isolated By (name) Doherty and colleagues	Isolated at Institute Queensland Inst of Medical Research, Brisbane	
Host Genus 31 mosquitoes of 11 species	Species	Host Age/Stage Adult
Sex Female		
<u>Isolated From</u>	<u>Isolation Details</u>	
Signs and Symptoms of Illness	Arthropod	
Time Held Alive before Inoculation		
Collection Method Light and chicken-baited traps	Collection Date 10/16/1963	
Place Collected (Minimum of City, State, Country) Dinner Creek, near Innisfail, N. Queensland, Australia		
Latitude 17° 30' S	Longitude 146° 0' E	
Macrohabitat In mixed mesophyll vine forest between Graham Range and Eubenangee swamp	Microhabitat Light traps 1.4 meters above ground, chicken traps 0.4-1 meters above ground	Method of Storage until Inoculated On solid CO2 or in Revco at -57dC
Footnotes		

Section III - Method of Isolation

Inoculation Date
10/25/1963

Animal (Details will be in Section 6)
nb mice

Route Inoculated Intracerebral	Reisolation Not tried
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Other Reasons

Homologous Antibody Formation by Source Animal

Test(s) Used

Footnotes

Section IV - Virus Properties

Physicochemical

Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

Stability of Infectivity (effects)

pH (infective range)

Lipid Solvent (ether - % used to test) 50% final	After Treatment Titer (a) 3.5;(b) 4.7 dex	Control Titer (a) 4.2;(b) 4.8 dex
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate) 1:400	After Treatment Titer (a) 3.5;(b) 4.6 dex	Control Titer (a) 4.6;(b) 5.3 dex
Other (formalin, radiation)		

Virion Morphology

Shape Orbivirus-like taxon	Dimensions 58-63 nm	
Mean nm	Range nm	
Measurement Method Electron microscopy (1)	Surface Projections/Envelope	Nucleocapsid Dimensions, Symmetry

Morphogenesis

Site of Constituent Formation in Cell	Site of Virion Assembly	Site of Virion Accumulation
Inclusion Bodies	Other	

Hemagglutination

Hemagglutination No	Antigen Source SMB, blood, carcass, liver ext. by sucrose-acetone	Erythrocytes (species used) Gander
pH Range 6.0-7.6	pH Optimum	
Temperature Range RT	Temperature Optimum	
Remarks		
Serologic Methods Recommended CF		
Footnotes		

Section V - Antigenic Relationship and Lack of Relationship to Other Viruses

No hemagglutin could be prepared but antisera to IN1074 (prepared in mice or rabbits by three or six weekly intraperitoneal inoculations) did not inhibit hemagglutination by antigens of Groups A (Sindbis, Getah, Ross River), B (Murray Valley encephalitis) or Koongol (Koongol, Wongal).

Complement-fixation tests (homologous titres - antigen = 32-64, mouse serum after inoculations = 64) and neutralization tests (homologous neutralization indices of mouse or rabbit antisera 1.6-2.1 dex) failed to show any relationship between IN1074, its antisera, and antigens or antisera to arboviruses previously isolated in Australia-Murray Valley encephalitis, Kunjin, Kokobera, Edge Hill, Stratford, Ross River, Sindbis, Getah, Koongol, Wongal, Corriparta, Mapputta, Kowanyama, Trubanaman, or to the exotic viruses Japanese B encephalitis, Bebaru, and dengue types 1-4 [2].

Serologically related to Pata virus, DakAr 1327, isolated from a pool of 35 *Aedes palpalis* captured in Pata, Central African Republic in November 1968 ([4], [5]).

The Eubenangee serogroup presently consists of three registered viruses including Eubenangee, Pata and Tilligerry viruses.

In addition, viruses of the bluetongue, Eubenangee and EHD serogroups share antigens which can be detected in complement-fixation tests and in agar-gel precipitin tests [8] - [10].

Section VI - Biologic Characteristics

Virus Source (all VERTEBRATE isolates)

Lab Methods of Virus Recovery (ALL ISOLATIONS)
Newborn mice

Cell system (a)	Virus passage history (b)	Evidence of Infection							Growth Without CPE +/- (g)		
		CPE			PLAQUES						
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)				
PS (CL)			No CPE								
Vero (CL)			No CPE								
BHK-21 (CL)			No CPE					+			

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Mixed mosquito pool *	1/29,395		Innisfail, N. Queensland, AS, 1963-64
Man		0/311 NT	Coastal areas of Queensland, AS
Cattle		3/36 NT	Mitchell River, AS
Kangaroo		5/50 NT	W. Queensland, AS
Wallaby		2/20 NT	Mitchell River, AS
Wallaby		10/49 NT	SE Queensland, AS
culicoides marksi	1		Beatrice Hill, No. Terr., Australia(7)
Cx. annulirostris	1		

Negative NT results were obtained with serum from the following species collected between 1958 and 1969 in various areas of Queensland: cattle, 60; horse, 50; sheep, 38; pig, 32; goat, 8; kangaroo, 4; bandicoot, 51; rat species, 85; Pteropus sp., 6; bat, 8; domestic fowl, 223; and wild birds, 36. (3)

* Mixed pool included 7 Culex (Lop) sp., 1 Cx sitiens, 3 Cx pullus, 6 Coquillettidia crassipes, 3 Hodgesia quasisanguinea, 1 Mansonia septempunctata, 3 Aedes notsocriptus, 1 Ae kochi, 3 Ae carmentis, 2 Anopheles bancroftii, and 1 An farauti.

Section VIII - Susceptibility to Experimental Infection (include viremia)

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log10/ml
Mice (nb)	SMB 4	ic 0.15	Death	6-9	7.0
Mice (nb)		ip 0.3	None		<3.5
Mice (nb)		sc			
Mice (wn)		ic 0.3	None	<3.5	
Mice (wn)		ip			
Mice (nb)	SMB 16	ic 0.015	Death	4-6	7.5

Section IX - Experimental Arthropod Infection and Transmission

Arthropod species & virus source(a)	Method of Infection log10/ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log10/ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System
Culex quinquefasciatus (SMB 16)			Adults intrathoracically inoculated: multiplication demonstrated by titration of whole mosquitoes, content per mosquito 1.9 dex immediately after inoculation, 1.3 dex at 12 and 24 hours, 5.0 dex at 7 days, 3.5 dex at 15 days. Serial weekly passage of salivary gland inoculated into thorax maintained virus for 2 passages but not for 3. (4)						
Culicoides variipennis C. nubeculosis			Intrathoracically inoculated; multiplication (6).						
C. variipennis			Oral ingestion; multiplication and transmission (6).						

Section X - Histopathology

Character of lesions (specify host)	
<u>Inclusion Bodies</u>	<u>Intranuclear</u>
Organs/Tissues Affected	
Category of tropism	

Section XI - Human Disease

In Nature	Residual	Death
Subclinical	Overt Disease	
Clinical Manifestations		
Number of Cases	Category (i.e. febrile illness, etc.)	

Section XII - Geographic Distribution

Known (Virus detected) Innisfail, North Queensland, Australia
Suspected (Antibody only detected)

Section XIII - References

1. Schnagl, et al. 1969. *Virology*. 38:347-349.
2. Doherty, R.L., et al. 1968. *Trans. R. Soc. Trop. Med. Hyg.* 62:862-867.
3. Doherty, R.L., et al. 1970. *Trans. R. Soc. Trop. Med. Hyg.* 64:748-753.
4. Director, Yale Arbovirus Research Unit. Personal communication. 1972.
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6. Mellor, P.S. and Jennings, M. 1980. *Arch. Virol.* 63:203-208.
7. Mahoney, D.F., Chief, CSIRO Div. Animal Hlth. Personal communication. 1983.
8. Gorman, B.M. 1979. *J. Gen. Virol.* 44:1-15.
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10. Voerwoerd, D.W., et al. 1979. *Orboviruses*. In: *Comprehensive Virology* (H. Fraenkel-Conrat and R.R. Wagner, eds.), Vol. 14, pp. 285-345, Plenum Press, New York.

Remarks
