

<b>Virus Name: Fort Morgan</b>		<b>Abbreviation: FMV</b>
Status <b>Arbovirus</b>	Select Agent <b>No</b>	SALS Level <b>2</b>
SALS Basis <b>Results of SALS surveys and information from the Catalogue.</b>		
Other Information		
Antigenic Group <b>A</b>		

**SECTION I - Full Virus Name and Prototype Number**

Prototype Strain Number / Designation <b>73V4009</b>	Accession Number	Original Date Submitted <b>10/1/1984</b>
Family <b>Togaviridae</b>	Genus <b>Alphavirus</b>	
Information From <b>D.B. Francy</b>	Address <b>Vector-Borne Diseases Division, Center for Disease Control, Fort Collins, Colorado</b>	
Information Footnote <b>Reviewed by editor</b>		

**Section II - Original Source**

Isolated By (name) <b>Vector-Borne Diseases Division (1)</b>	Isolated at Institute <b>CDC, Fort Collins, Colorado</b>	
Host Genus <b>Oeciacus vicarius Horvath (Hemiptera: Cimicidae), pool of 25</b>	Species	Host Age/Stage <b>Adult</b>
Sex <b>Not Answered</b>		
<u>Isolated From</u>	<u>Isolation Details</u>	
Signs and Symptoms of Illness	Arthropod <b>Depleted</b>	
Time Held Alive before Inoculation <b>2 weeks at 22 +/- 2dC</b>		
Collection Method <b>Hand collection from cliff swallow nests</b>	Collection Date <b>9/26/1973</b>	
Place Collected (Minimum of City, State, Country) <b>Bijou Bridge, Morgan County, Colorado</b>		
Latitude <b>40° 16' N</b>	Longitude <b>103° 45' W</b>	
Macrohabitat <b>Concrete bridge over cottonwood-lined stream in irrigated farmland, S. Platte River Basin</b>	Microhabitat <b>Nest of cliff swallow occupied by house sparrows</b>	Method of Storage until Inoculated <b>Held with nest material in plastic sack at room temperature.</b>
Footnotes		

**Section III - Method of Isolation**

Inoculation Date  
**9/1/1973**

Animal (Details will be in Section 6)  
**(Tissue Culture)**

Route Inoculated

Reisolation

Other Reasons

**Different from all other alphaviruses in laboratory by serologic and physicochemical tests**

Homologous Antibody Formation by Source Animal

Test(s) Used

Footnotes

**Section IV - Virus Properties**

Physicochemical  
**RNA, Single Strand**

Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number <b>3 (2)</b>	Details <b>E1 glycoprotein, MW 58,000, pI 7.8; E2 glycoprotein, MW 47,000, pI 9.2; C (nucleocapsid), MW 30,000</b>	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

**Stability of Infectivity (effects)**

pH (infective range)  
**2.0-4.0 inactivates 5.0 dex Vero PFU infectivity**

Lipid Solvent (ether - % used to test)	After Treatment Titer	Control Titer
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate) <b>1:1000</b>	After Treatment Titer <b>&lt;2.0 dex</b>	Control Titer <b>5.4 dex</b>
Other (formalin, radiation)		

**Virion Morphology**

Shape <b>Spherical</b>	Dimensions	
Mean nm	Range <b>55-60 nm</b>	
Measurement Method <b>Thin section electron microscopy (3)</b>	Surface Projections/Envelope <b>Spikes</b>	Nucleocapsid Dimensions, Symmetry

**Morphogenesis**

Site of Constituent Formation in Cell  
**Cytoplasm (nucleocapsids accumulate in nucleus late in infection)**

Site of Virion Assembly  
**Cytoplasmic and plasma membranes**

Site of Virion Accumulation  
**Extracellular**

Inclusion Bodies

Other

**Hemagglutination**

Hemagglutination  
**Yes**

Antigen Source  
**Primary duck embryo, Vero cells, SMB**

Erythrocytes (species used)  
**Goose**

pH Range  
**5.9-6.4**

pH Optimum  
**6.0**

Temperature Range

Temperature Optimum  
**37dC**

Remarks

Serologic Methods Recommended  
**HI, CF, NT**

Footnotes

**Section V - Antigenic Relationship and Lack of Relationship to Other Viruses**

Antigen or Antibody of Indicated Virus	Fort Morgan Antigen*			Fort Morgan Antibody*		
	HI	CF	NT	HI	CF	NT
Fort Morgan	40	256	320	40	256	320
WEE (Fleming)	320/320	128/>1024	-/640	10/40	128/256	-/320
Y62-33	320/640	128/512	80/640	-/40	128/256	-/320
HJ	20/80	8/64	-/320	10/40	32/256	-/320
Sindbis	320/640	128/512	-/160	-/40	64/256	-/320
Whataroa	20/	-/32	-/160	ND	-/256	-/320
Aura	20/80	-/64	20/160	-/40	-/256	-/320
Chikungunya	40/160	-/256	-/>640	-/40	-/256	-/320
Mayaro	-/40	-/32	-/160	-/40	-/256	-/320
Semliki Forest	-/640	-/64	-/>640	-/40	-/256	-/320

Una	-/	-/16	-/40	ND	-/256	-/320
O'nyong nyong	-/40	-/32	-/160	-/40	-/256	-/320
Middelburg	-/	-/16	-/320	ND	-/256	-/320
EEE	-/80	-/128	-/>640	-/40	-/256	-/320
VEE IA	-/160	8/128	-/>640	-/40	-/256	-/320
IB	-/80	-/64	-/320	-/40	-/256	-/320
IC	-/640	-/64	-/>640	-/40	-/256	-/320
ID	40/80	16/128	-/160	-/40	-/256	-/320
IE	-/80	-/64	-/160	-/40	-/256	-/320
II	-/640	-/256	-/>640	-/40	64/256	-/320
III	-/160	-/32	-/160	-/40	-/256	-/320
IV	-/>640	-/256	20/160	-/40	-/256	-/320
Bijou Bridge (CM4-146)	10/640	-/64	-/640	-/40	-/256	-/320

\* Ht/Ho; - = <10 HI, NT; <8 CF

Section VI - Biologic Characteristics

Virus Source (all VERTEBRATE isolates)  
Serum (LV), brain (LV)

Lab Methods of Virus Recovery (ALL ISOLATIONS)  
Plaque formation in primary duck embryo or Vero cell cultures; ic inoculation of newborn mice unsatisfactory for primary isolation

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
Duck embryo (PC)	unpass.	2-3	4+		2-3	2-4 mm		
	DE 1 Vero 1, Vero 5						5.7**	
Vero (CL)		2-3	4+		2-3	2-4 mm	7.8	
Aedes albopictus (CL)								+ -
L929 (CL)						No plaques	<2.0	

\*\* Expressed in dex

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Oeciacus vicarius	154/28,453 (1,201 pools)		NE Colorado, USA
Oeciacus vicarius	6/880 (18 pools)		South Dakota (Bon Home Co.), USA
Oeciacus vicarius	1/8,068 (100 pools)		West Texas, USA
Oeciacus vicarius	2/590 (14 pools)		Washington (Bent Co.), USA
P. pyrrhonota nestling (serum/brain)	14/291		NE Colorado, USA
P. pyrrhonota adult		43/296 NT	
P. domesticus nestling (serum/brain)	127/1,171		
P. domesticus adult		64/312 NT	
Dipodomys ordii		3/64 NT	
Peromyscus maniculatus		1/41 NT	
Perognathus flavus		0/7 NT	
Mus musculus		0/4 NT	
Sylvilagus sp.		0/7 NT	

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log <sub>10</sub> /ml
Mice (nb)	Unpass.; Vero 1, or Vero 5	ic	Viral growth in CNS, other tissues *		
Mice (nb)		ip	Viral growth in CNS, other tissues *		
Mice (nb)		sc			
Mice (wn)		ic	Antibody		
Mice (wn)		ip	Antibody		
hamsters (nb)		ic	Death	4.0	7.1
hamsters (nb)		ip	Death	4.0	6.4
hamsters (wn)		ic	Paralysis, scattered deaths		
hamsters (wn)		ip	Antibody, paralysis, occasional death		
Passer domesticus					
nestling		sc	Viremia, occasional death (5)		
adult		sc	Viremia		
Petrochelidon					
pyrrhonota: nestling		sc	Viremia		
pyrrhonota: adult		sc	Viremia		

\* and variable mortality: 5-10% for unpassaged, 20-80% for Vero 5 passaged virus at all dilutions 5-7 days after inoculation.

**Section IX - Experimental Arthropod Infection and Transmission**

Arthropod species & virus source(a)	Method of Infection log10/ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log10/ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System

*O. vicarius* bugs fed on suspensions of Fort Morgan virus containing 4.5-6.9 logs were shown to contain virus after up to 312 days. Infected bugs transmitted the virus after 33 and 47 days incubation to nestling sparrows. Infection and transmission was demonstrated after 234 and 312 days incubation in bugs held at temperatures simulating hibernation.

*Culex tarsalis* and *Cx pipiens pipiens* fed on virus (suspensions and viremic hosts) and held for 9-23 days at 22-24C contained no detectable virus. Minimal levels (1-1.3 dex PFU/ml) were detected in a small proportion of the mosquitoes after intrathoracic inoculation of virus.

*Culiseta melanura* and *Cs inornata* did not become infected after oral feeding (pledget suspensions, 6.3 dex/ml) but following intrathoracic inoculation apparent replication to higher levels than in *Culex* mosquitoes was observed.

**Section X - Histopathology**

Character of lesions (specify host)

**Inflammation, necrosis in brain, spinal cord, skeletal muscle of infected infant mice.**

Inclusion Bodies

Intranuclear

Organs/Tissues Affected

**Brain and cord: lymphocytic perivascular cuffing, endothelial cell swelling, diffuse gliosis, focal necrosis, neuronal degeneration Striated muscle: degeneration and necrosis of myofibers, calcification.**

Category of tropism

**Neuromuscular**

**Section XI - Human Disease**

In Nature	Residual	Death
Subclinical	Overt Disease	
Clinical Manifestations		
Number of Cases	Category (i.e. febrile illness, etc.)	

**Section XII - Geographic Distribution**

Known (Virus detected)

**Colorado; South Dakota; Texas; Washington, U.S.A.**

Suspected (Antibody only detected)

**Section XIII - References**

1. Hayes, R.O. et al. 1977.. J. Med. Ent. 14:257-262.
2. Trent, D.W. et al. 1980. J. Gen. Virol. 47:261-282.
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5. Monath, T.P. et al. 1980. Am. J. Trop. Med. Hyg. 29:969-983.
6. Rush, W.A. et al. 1980. Ann. Ent. Soc. Am. 73:315-318.

**Remarks**