

Virus Name: Amapari		Abbreviation: AMAV
Status Not Arbovirus	Select Agent No	SALS Level 2
SALS Basis Level 2 arenaviruses are not known to cause serious acute disease in man and are not acutely pathogenic for laboratory animals, including primates. Survey experience is sufficient to conclude that laboratory aerosol infection does not occur in the course of routine work with cell cultures and animals not subject to chronic infection. In view of reported high frequency of laboratory aerosol infection that occurred in workers manipulating high concentrations of Pichinde virus, it is strongly recommended that work with high concentrations of Level 2 arenaviruses be done at Level 3.		
Other Information		
Antigenic Group Tacaribe		

SECTION I - Full Virus Name and Prototype Number

Prototype Strain Number / Designation BeAn 70563	Accession Number	Original Date Submitted 12/14/1984
Family Arenaviridae	Genus Areavirus	
Information From F.P. Pinheiro	Address Instituto Evandro Chagas, Caixa Postal 232, Belem, Para, Brazil	
Information Footnote Reviewed by editor		

Section II - Original Source

Isolated By (name) Pinheiro et al.	Isolated at Institute Belem Virus Laboratory	
Host Genus Neacomys guianae	Species	Host Age/Stage Young Adult
Sex Male		
<u>Isolated From</u>	<u>Isolation Details</u>	
Organs/Tissues	Pool of spleen, liver, heart, and kidney	
Signs and Symptoms of Illness	Arthropod	
Time Held Alive before Inoculation		
Collection Method Hardwood live trap	Collection Date 7/8/1964	
Place Collected (Minimum of City, State, Country) Serra do Navio, T.F. Amapa, Brazil		
Latitude 1° N	Longitude 52° W	
Macrohabitat Tropical forest, periodically inundated	Microhabitat Primary vegetation, ground level, shade	Method of Storage until Inoculated At -60° C
Footnotes		

Section III - Method of Isolation

Inoculation Date
7/20/1964

Animal (Details will be in Section 6)
nb mice

Route Inoculated Intracerebral	Reisolation Not tried
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Other Reasons
13 further strains isolated from Neacomys and Oryzomys

Homologous Antibody Formation by Source Animal
Not tested

Test(s) Used

Footnotes

Section IV - Virus Properties

Physicochemical

Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

Stability of Infectivity (effects)

pH (infective range)

Lipid Solvent (ether - % used to test)	After Treatment Titer	Control Titer
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate)	After Treatment Titer	Control Titer
Other (formalin, radiation)		

Virion Morphology

Shape	Dimensions 60 - 280 nm	
Mean nm	Range nm	
Measurement Method Electron microscopy	Surface Projections/Envelope	Nucleocapsid Dimensions, Symmetry

Morphogenesis

Site of Constituent Formation in Cell

Site of Virion Assembly

Site of Virion Accumulation

Inclusion Bodies

Other

Hemagglutination

Hemagglutination

No

Antigen Source

SMB ext. by sucrose-acetone

Erythrocytes (species used)

Goose

pH Range

6.0 - 7.0

pH Optimum

Temperature Range

Room and 37° C

Temperature Optimum

Remarks

Serologic Methods Recommended

CF and NT

Footnotes

Section V - Antigenic Relationship and Lack of Relationship to Other Viruses

Immune Sera or Antigens	Amapari Antigen			Amapari Immune Serum		
	CF Ht/Ho	NT Ht/Ho	PNT* Ht/Ho	CF Ht/Ho	NT Ht/Ho	PNT Ht/Ho
Tacaribe	32/256	0.2/2.0	<4/2048	32/128	0.6/1.6	<4/1024
Junin	32/128	0.3/3.0	<4/256	64/128	<0.5/1.6	<4/1024
Machupo			<4/128			<4/1024

NT: LNI in dex

PNT: Plaque neutralization test

Section VI - Biologic Characteristics

Virus Source (all VERTEBRATE isolates)
 Blood (LV), heart, liver, spleen, kidney poll (LV)

Lab Methods of Virus Recovery (ALL ISOLATIONS)
 newborn mice

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
GMK (CL)	SMB 6	CF antigen in fluid.	No CPE	8.5 *				
BHK-21 (CL)			No CPE					
HEp-2 (CL)			No CPE					
Chick embryo (PC)						No plaques		
Vero (CL)						Plaques	c.5.0 *(5)	
Vero (CL)	P-Unk.				2	2mm	7.7 (6)	
LLC-MK2 (CL)					6	2mm	6.3 (6)	
BHK-21 (CL)	SMB 11	4	2+-3+	7.5 (7)				

* Expressed in dex

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Oryzomys capito	124/918	2/18 CF	Serra do Navio, Brazil
Oryzomys goeldii young	5/41		Territorio, Federal do Amapa, Brazil
Neacomys guianae	97/1,095	1/4 CF	
Ectoparasites (Laelopidae)	4/341 pools		Brazil

No isolations made from 3,762 specimens obtained from bats, marsupials, birds, primates or other species of rodents; nor from 24,140 mosquitoes or 792 groups of sentinel mice.

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log ₁₀ /ml	
Mice (nb)	SMB 4	ic 0.02	Death or br.CF antigen	13.7	4.2	
Mice (nb)		ip 0.02	Death or br.CF antigen	18.0		
Mice (nb)		sc				
Mice (wn)		ic 0.02	Antibody formation			
Mice (wn)		ip 0.2	Antibody formation			
Mice (nb)		ic 0.02	Death		6.7	
guinea pigs	SMB 5	ip	Antibody			
ID50 as determined by presence of CF antigen in brain					8.4 ic	
					<4.2 ip	

Section IX - Experimental Arthropod Infection and Transmission

Section X - Histopathology

Character of lesions (specify host)

3/8 nat. infected forest rodents had focal degenerative inflammatory lesions of myocardium (3). SM, ic: choroiditis, glial hyperplasia, vasculitis, perivasculitis with infiltration of lymphocytes in CNS. Mortality and histopath. changes not influenced by thymectomy

Inclusion BodiesIntranuclear

Organs/Tissues Affected

Category of tropism

Section XI - Human Disease

In Nature	Residual	Death
Subclinical	Overt Disease	
Clinical Manifestations		
Number of Cases	Category (i.e. febrile illness, etc.)	

Section XII - Geographic Distribution

Known (Virus detected) Brazil
Suspected (Antibody only detected)

Section XIII - References

1. Pinheiro, F.P., et al. 1966. Proc. Soc. Exp. Biol. Med. 122:531-535. 2. Murphy, F.A., et al. 1970. J. Virol. 6:507-518. 3. Dias, L.B. Personal communication. 4. Besuchio, S.C., et al. 1973. Arch. ges. Virusforsch. 40:21-28. 5. Webb, P.A. Personal communication. 6. Stim, T.B. 1969. J. Gen. Virol. 5:329-338. 7. Karabatsos, N. and Buckley, S.M. 1967. Am. J. Trop. Med. Hyg. 16:99-105.
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Remarks

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