

<b>Virus Name: Ippy</b>		<b>Abbreviation: IPPYV</b>
Status <b>Possible Arbovirus</b>	Select Agent <b>No</b>	SALS Level <b>2</b>
SALS Basis <b>Results of SALS surveys and information from the Catalogue.</b>		
Other Information		
Antigenic Group <b>Tacaribe</b>		

**SECTION I - Full Virus Name and Prototype Number**

Prototype Strain Number / Designation <b>DakAnB 188d</b>	Accession Number	Original Date Submitted <b>8/28/1984</b>
Family <b>Arenaviridae</b>	Genus <b>Arenavirus</b>	
Information From <b>J.P. Digoutte</b>	Address <b>Institut Pasteur B.P. 304 Cayenne Guyane Francaise</b>	
Information Footnote <b>Reviewed by editor</b>		

**Section II - Original Source**

Isolated By (name) <b>J.P. Digoutte and F.X. Pajot</b>	Isolated at Institute <b>Institut Pasteur, Bangui</b>	
Host Genus <b>Arvicanthis</b>	Species	Host Age/Stage <b>Adult</b>
Sex <b>Male</b>		
<u>Isolated From</u>	<u>Isolation Details</u>	
<b>Organs/Tissues</b>	<b>Brain, liver, spleen pool</b>	
Signs and Symptoms of Illness	<b>Arthropod</b>	
Time Held Alive before Inoculation		
Collection Method <b>Caught by trap</b>	Collection Date <b>6/26/1970</b>	
Place Collected (Minimum of City, State, Country) <b>Ippy, Central African Republic</b>		
Latitude <b>6° 15' N</b>	Longitude <b>21° 12' E</b>	
Macrohabitat <b>Savannah with forest gallery</b>	Microhabitat	Method of Storage until Inoculated <b>Liquid nitrogen, then Revco at -75dC</b>
Footnotes		

**Section III - Method of Isolation**

Inoculation Date  
**8/10/1970**

Animal (Details will be in Section 6)  
**nb mice**

Route Inoculated  
**ic, ip and sc**

Reisolation  
**Yes**

Other Reasons

Homologous Antibody Formation by Source Animal

Test(s) Used

Footnotes

**Section IV - Virus Properties**

Physicochemical

Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

**Stability of Infectivity (effects)**

pH (infective range)

Lipid Solvent (ether - % used to test)	After Treatment Titer	Control Titer
Lipid Solvent (chloroform)	After Treatment Titer <b>&lt;2.0 dex</b>	Control Titer <b>5.0 dex</b>
Lipid Solvent (deoxycholate)	After Treatment Titer	Control Titer

Other (formalin, radiation)

**Virion Morphology**

Shape	Dimensions	
Mean nm	Range nm	
Measurement Method	Surface Projections/Envelope	Nucleocapsid Dimensions, Symmetry

### Morphogenesis

Site of Constituent Formation in Cell                      Site of Virion Assembly                      Site of Virion Accumulation

Inclusion Bodies                      Other

### Hemagglutination

Hemagglutination                      Antigen Source                      Erythrocytes (species used)

Not tried

pH Range                      pH Optimum

Temperature Range                      Temperature Optimum

Remarks

Serologic Methods Recommended

CF

Footnotes

### **Section V - Antigenic Relationship and Lack of Relationship to Other Viruses**

CF titer of the immune ascitic fluid = 512 with homologous antigen diluted to 1/6 Institut Pasteur (Dakar) [4] . DakAnB 188 gives negative results with the following viruses:

Group A;	Semliki Forest virus, chikungunya, o'nyong-nyong, Igbo-Ora (B 543), Sindbis, ArY 251, Middelburg, Ndumu.
Group B;	Ntaya, Bagaza (ArB 209), Wesselsbron, ArY 310, Usutu, ArY 276, West Nile, Koutango (AnD 5443), Dakar bat, Uganda S, Saboya (AnD 4600), Banzi, Bouboui (ArB 490), yellow fever, Zika, Spondweni, Bukalasa bat (AnD 595), Royal Farm (T 285), Kadam (UgAr 6640), Cambodia (AnC 38), AnY 589.
Bunyamwera;	Bunyamwera, Germiston, Ilhesha, Shokwe, Birao (ArB 2198).
Simbu;	Buttonwillow, Ingwavuma, Simbu, Nola (ArB 2882).
Bwamba;	Bwamba, Pongola.
California;	Lumbo.
Olifantsvlei;	Olifantsvlei, Bobia (ArB 1569), Botambi (ArB 937).
Tete;	Tete (SAAn 3518), Bahig (EgB 90), Matruh (EgAn 1047), DakAnB 1422, AnB 1292, AnB 1544, AnB 1564.
Phlebotomus;	Arumowot (SudAr 1284), Gabek Forest (AnD 3150), Gordil (AnB 496), Saint-Floris (AnB 512-Re), Zinga (ArB 1976).
Boteke;	Boteke (ArB 1077), Zingilamo (AnB 1245).
Corriparta;	Acado (EhAr 1846), Corriparta, ArB 3689.
Eubenangee;	Eubenangee (In 1074), Pata (ArB 1327).

Matariya;	Burg el Arab (EgAn 3782), Matariya (EgAn 1477), AnB 423.
Mossuril;	Bangoran (ArB 2053), Mossuril, Kamese.
Nyando;	Eret 147 (AnY 176), Nyando.
Turlock;	Yaba 1 (ArB 365).
Congo;	Congo (IbAr 10200).
NSD;	Dugbe (IbAn 1792).
Kemerovo;	Chenuda, Wad Medani.
Qalyub;	Bandia (ArD 611), Qalyub (EgAr 370).
Quaranfil;	Quaranfil.
Thogoto;	Thogoto.
Keuraliba;	Keuraliba (AnD 5314), Le Dantec (HD 763).
Nyamanini;	Nyamanini.
Ungrouped;	Lebombo, Nkolbisson, Okola, Tanga, Tataguine, Witwatersrand, Orungo (ArB 2078), SudAr 1275, Bhanja (ArD 9540), Jos (ArD 15131), Somone (ArD 4499), IbAn 39048 (ArYT 34), Gossas (AnD 401), Toure (AnD 4611), Yogue (AnD 5634), AnD 11411, Yata (ArB 2181), Bangui (HB 754), Landjia (AnB 769), Gomoka (AnB 787), Kolongo (AnB 1094), Bimbo (AnB 1054), Ouango (AnB 1582), Bobaya (AnB 2208), Oubangui (ArB 3816), Sandjimba (AnB 373), AnB 277, AnB 1227, AnB 1398, AnB 904, AnYV 6, AnYV 177.
Nonarbovirus;	Herpesvirus (HB 3667).

In addition, DakAnB 188 antigen failed to react with the following immune fluids:

Group B;	Entebbe bat, Montana Myotis leukoencephalitis, dengue 1, 2, 3, 4, Potiskum (IbAn 10069).
Simbu;	Sango, Shamonda, Sabo, Shuni, Sathuperi, Yaba 7.
California;	Group serum.
EHD;	IbAr 22619.
VSV;	Chandipura (IbAn 9978).
Phlebotomus;	Group serum.
Kaisodi-Qalyub-Quaranfil;	Serum polyvalent (Johnston Atoll, Kaisodi, Bandia, Silverwater, Quaranfil, Lanjan, Qalyub).
Uukuniemi;	Grand Arbaud, Ponteves, EgAn 1825.
Bluetongue;	type 10 (IbAr 22618).
AHS;	IbAn 53177

Rabies serogroup;	Lagos bat.
Ungrouped;	IbAr 23380, IbAn 2898 (Oyo), IbAn 17143, Mount Elgon bat, IbAn 33709, IbAn 28946, EgAn 1398/61 (IbAn 39652).
Nonarbovirus;	LCM, rabies, IbAn 27377 and IbH 29777 (related to rabies), IbAn 20433 (NDV).

In addition DakAnB 188 antigen was screened against NIH grouping fluids with negative results [4] .

Recent cross-immunofluorescence studies have shown that Ippy virus is related to Lassa virus. Antibody to Ippy virus reacted to high titer with Lassa virus and Lassa-related viruses, while monoclonal antibody to Lassa virus reacted with Ippy virus. The exact relationship of Ippy virus to other arenaviruses of the Tacaribe serogroup remains to be determined [5] , [6] .

### Section VI - Biologic Characteristics

Virus Source (all VERTEBRATE isolates)      Lab Methods of Virus Recovery (ALL ISOLATIONS)  
**Blood (LV), CNS (LV), liver (LV), spleen (LV) (pooled tissues)**      **Newborn mice**

Cell system (a)	Virus passage history (b)	Evidence of Infection							Growth Without CPE +/- (g)
		CPE			PLAQUES				
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)		

Section VII - Natural Host Range (Additional text can be added below table)

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Lemniscomys striatus	1/103		Centr.Afr.Republic (1)
Mastomys	1/448		
Arvicanthus	3/81		Centr.Afr.Republic (1, 2)
Praomys (blood) *	10/225		Centr.Afr.Republic (2, 3)

\* Two isolates were obtained from blood and organs of the same Praomys.

Section VIII - Susceptibility to Experimental Infection (include viremia)

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log <sub>10</sub> /ml
Mice (nb)	SM 13	ic 0.02	Death	9-14	5
Mice (nb)		ip 0.03	No illness		
Mice (nb)		sc			
Mice (wn)		ic 0.03	No illness		
Mice (wn)		ip 0.01	No illness		



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