

<b>Virus Name: Anhanga</b>		<b>Abbreviation: ANHV</b>
Status <b>Possible Arbovirus</b>	Select Agent <b>No</b>	SALS Level <b>2</b>
SALS Basis <b>Results of SALS surveys and information from the Catalogue.</b>		
Other Information		
Antigenic Group <b>Phlebotomus Fever</b>		

**SECTION I - Full Virus Name and Prototype Number**

Prototype Strain Number / Designation <b>BeAn 46852</b>	Accession Number	Original Date Submitted <b>2/27/1985</b>
Family <b>Bunyavirus</b>	Genus <b>Phlebovirus</b>	
Information From <b>Belem Virus Lab.</b>	Address <b>Belem Virus Laboratory, Instituto Evandro Chagas, Belem, Para, Brazil</b>	
Information Footnote <b>Reviewed by editor</b>		

**Section II - Original Source**

Isolated By (name) <b>Belem Virus Laboratory</b>	Isolated at Institute <b>Belem, Para, Brazil</b>	
Host Genus <b>Choloepus brasiliensis</b>	Species	Host Age/Stage
Sex <b>Female</b>		
<u>Isolated From</u>	<u>Isolation Details</u>	
<b>Organs/Tissues</b>	<b>Heart, liver, spleen, kidney pool</b>	
Signs and Symptoms of Illness <b>None</b>	Arthropod	
Time Held Alive before Inoculation		
Collection Method <b>Captured by hand</b>	Collection Date <b>10/1/1962</b>	
Place Collected (Minimum of City, State, Country) <b>Castanhal Forest, Para, Brazil</b>		
Latitude <b>2° S</b>	Longitude <b>48° W</b>	
Macrohabitat <b>Virgin forest</b>	Microhabitat <b>Canopy</b>	Method of Storage until Inoculated <b>At -60dC</b>
Footnotes		

**Section III - Method of Isolation**

Inoculation Date <b>10/8/1962</b>	
Animal (Details will be in Section 6) <b>nb mice</b>	
Route Inoculated <b>Intracerebral</b>	Reisolation <b>Yes</b>
Other Reasons	
Homologous Antibody Formation by <u>Source Animal</u> <b>Not tested</b>	
Test(s) Used	
Footnotes	

**Section IV - Virus Properties**

<b>Physicochemical</b>		
Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	
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<b><u>Stability of Infectivity (effects)</u></b>		
pH (infective range)		
Lipid Solvent (ether - % used to test)	After Treatment Titer	Control Titer
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate) <b>1:1000</b>	After Treatment Titer <b>2.6 dex</b>	Control Titer <b>5.1 dex</b>
Other (formalin, radiation)		
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<b><u>Virion Morphology</u></b>		
Shape	Dimensions	
Mean nm	Range nm	
Measurement Method	Surface Projections/Envelope	Nucleocapsid Dimensions, Symmetry

### Morphogenesis

Site of Constituent Formation in Cell      Site of Virion Assembly      Site of Virion Accumulation

Inclusion Bodies      Other

### Hemagglutination

Hemagglutination      Antigen Source      Erythrocytes (species used)  
**Yes**      **SMB ext. by sucrose-acetone**      **Goose**

pH Range      pH Optimum  
**5.7-7.0**      **6.6**

Temperature Range      Temperature Optimum  
**37dC**

Remarks  
**HI, CF, NT**

Serologic Methods Recommended

Footnotes  
**HI, CF, NT**

### **Section V - Antigenic Relationship and Lack of Relationship to Other Viruses**

HI (4 units):

Sera	Antigens				
	Icoaraci	Candiru	Itaporanga	Anhanga	Bujaru
Icoaraci	> 1280	> 640	80	20	160
Candiru	40	> 640	0	0	0
Itaporanga	320	0	> 640	0	0
Anhanga	320	0	0	160	40
Bujaru	80	0	0	0	> 640
Chagres	640	0	0	20	20

0 = <10

All sera were hyperimmune mouse 10090 .

For additional information on antigenic relationships, see References [4] , [5] , [6] .

**Section VI - Biologic Characteristics**

Virus Source (all VERTEBRATE isolates)  
Pool of heart, liver, spleen, kidney (LV)

Lab Methods of Virus Recovery (ALL ISOLATIONS)  
Newborn mice

Cell system (a)	Virus passage history (b)	Evidence of Infection						Growth Without CPE +/- (g)
		CPE			PLAQUES			
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
Vero (CL)		P-2 prototype			5	1mm	5.6 <sup>*</sup> (3)	
LLC-MK2 (CL)					3	3mm	6.4 (3)	

<sup>\*</sup> Expressed in dex

**Section VII - Natural Host Range (Additional text can be added below table)**

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tsted Test used	Country and region
Choloepus brasiliensis (sloth)	1/35		Para, Brazil
Choloepus brasiliensis (sloth)		0/29 HI	Panama (2)
Bradypus		0/21 HI	



**Section XII - Geographic Distribution**

Known (Virus detected)

**Brazil**

Suspected (Antibody only detected)

**Section XIII - References**

1. Woodall, J.P. 1967. Atas. Simpos. Biota Amazon. 6:31-63.
2. Belem Virus Laboratory, Belem, Brazil. 1966. Unpublished results.
3. Stim, T.B. 1969. J. Gen. Virol. 5:329-338.
4. Tesh, R.B., et al. 1975. Am. J. Trop. Med. Hyg. 24:135-144.
5. Tesh, R.B., et al. 1982. Ibid. 31:149-155.
6. Travassos Da Rosa, A.P.A., et al. 1983. Ibid. 32:1164-1171.

**Remarks**