

<b>Virus Name: Jurona</b>		<b>Abbreviation: JURV</b>
Status <b>Possible Arbovirus</b>	Select Agent <b>No</b>	SALS Level <b>2</b>
SALS Basis <b>Results of SALS surveys and information from the Catalogue.</b>		
Other Information		
Antigenic Group <b>Vesicular Stomatitis</b>		

**SECTION I - Full Virus Name and Prototype Number**

Prototype Strain Number / Designation <b>BeAr 40578</b>	Accession Number	Original Date Submitted <b>1/27/1985</b>
Family <b>Rhabdoviridae</b>	Genus <b>Vesiculovirus</b>	
Information From <b>Belem Virus Lab.</b>	Address <b>Belem Virus Laboratory, Instituto Evandro Chagas, Belem, Para Brazil</b>	
Information Footnote <b>Reviewed by editor</b>		

**Section II - Original Source**

Isolated By (name) <b>Belem Virus Laboratory</b>	Isolated at Institute <b>Belem, Para, Brazil</b>	
Host Genus <b>Haemagogus sp.</b>	Species	Host Age/Stage <b>Adult</b>
Sex <b>Female</b>		
<u>Isolated From</u>	<u>Isolation Details</u>	
Signs and Symptoms of Illness	Arthropod	
Time Held Alive before Inoculation		
Collection Method <b>By hand off human bait</b>	Collection Date <b>3/3/1962</b>	
Place Collected (Minimum of City, State, Country) <b>Belem-Brasilia Highway, km 87, Brazil</b>		
Latitude <b>3° S</b>	Longitude <b>48° W</b>	
Macrohabitat <b>Virgin forest</b>	Microhabitat <b>Canopy</b>	Method of Storage until Inoculated <b>-60dC</b>
Footnotes		

**Section III - Method of Isolation**

Inoculation Date  
**3/12/1962**

Animal (Details will be in Section 6)  
**nb mice**

Route Inoculated  
**Intracerebral**

Reisolation  
**Yes**

Other Reasons

Homologous Antibody Formation by Source Animal

Test(s) Used

Footnotes

**Section IV - Virus Properties**

Physicochemical

Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

**Stability of Infectivity (effects)**

pH (infective range)

Lipid Solvent (ether - % used to test)	After Treatment Titer	Control Titer
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate) <b>1:1000</b>	After Treatment Titer <b>&lt;3.5 dex</b>	Control Titer <b>6.3 dex</b>
Other (formalin, radiation)		

**Virion Morphology**

Shape <b>Bullet-shaped (5)</b>	Dimensions	
Mean nm	Range nm	
Measurement Method	Surface Projections/Envelope	Nucleocapsid Dimensions, Symmetry

### Morphogenesis

Site of Constituent Formation in Cell	Site of Virion Assembly	Site of Virion Accumulation
Inclusion Bodies	Other	

### Hemagglutination

Hemagglutination <b>Yes</b>	Antigen Source <b>SMB ext. by sucrose-acetone + protamine</b>	Erythrocytes (species used) <b>Goose</b>
pH Range <b>6.1-6.2</b>	pH Optimum <b>6.2</b>	
Temperature Range	Temperature Optimum <b>37dC</b>	
Remarks		
Serologic Methods Recommended <b>CF, HI, NT</b>		
Footnotes		

### **Section V - Antigenic Relationship and Lack of Relationship to Other Viruses**

Jurona HA antigen inhibited by Oropouche 1:20 (8 units) Simbu Grouping serum 1:10, Bunyamwera Grouping serum 1:40.

Jurona serum did not react with EEE, Aura, Una, Mayaro, YF, Ilheus, Bussuquara, SLE, Oriboca, Murutucu, Caraparu, Guama, Maguari, Guaroa, Icoaraci, Tacaiuma, Turlock, Candiru, Anhangá. in the HI test.

In CF Jurona antigen did not react with grouping sera of Groups A, B, C, Guama, Capim, Bunyamwera, Phlebotomus fever, California, Simbu, Bakau; nor with sera of epizootic hemorrhagic disease of deer-NJ, Navarro, Trinité, Ieri, Aruac, Tacaribe, Hart Park, CTF, Anopheles A and B, Panama J55, Tete, Cas Cas, TRVL 42336, Nyamanini, Witwatersrand, Quarantil, Lebombo, SF Sicilian, Bwamba, Wad Medani, Mossuril, Wongal, Mapputta, Wanowrie, Ganjam (IG 3159), Nodamura, Tsuruse, K-622, Chenuda, VSNJ, Mirim, Oropouche, Icoaraci, Palyam, Tacaiuma, Turlock, Cocal, Lukuni, Candiru, Piry, Pacui, Acara, Irituia, Marco, Timbo, Chaco, or Anhangá.

Following the observation that Jurona virus possessed rhabdovirus morphology [5], it was serologically placed in the VSV serogroup [6].

**Section VI - Biologic Characteristics**

Virus Source (all VERTEBRATE isolates)

Lab Methods of Virus Recovery (ALL ISOLATIONS)  
Newborn mice

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
BHK-21 (CL)	P-5				<4	Plaques	8.3* (4)	
Chick embryo (PC)					2-3	2 sizes	7.0 (4)	
Turkey embryo(PC)						Plaques	6.0 (4)	
GMK (CL)			CPE (4)					
Vero (CL)					2	8 mm	7.7 (3)	
LLC-MK2 (CL)					4	11 mm	7.2 (3)	

\* Expressed in dex

**Section VII - Natural Host Range (Additional text can be added below table)**

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Haemagogus spegazzinii	1		Para, Brazil (1)
Birds, forest		Of 174 sampled by HI, 10% had low-titered antibody not confirmable by NT (2).	



**Section XI - Human Disease**

In Nature	Residual	Death
Subclinical	Overt Disease	
Clinical Manifestations		
Number of Cases	Category (i.e. febrile illness, etc.)	

**Section XII - Geographic Distribution**

Known (Virus detected) <b>Brazil</b>
Suspected (Antibody only detected)

**Section XIII - References**

<ol style="list-style-type: none"><li>1. Woodall, J.P. 1967. Atas Simpos. Biota Amazon. 6:31-63.</li><li>2. Shope, R.E., et al. 1966. Am. J. Epidem. 84:467-477.</li><li>3. Stim, T.B. 1969. J. Gen. Virol. 5:329-338.</li><li>4. Pinheiro, F.P. Personal communication.</li><li>5. Araujo, R. Personal communication. 1982.</li><li>6. Tesh. R.B., et al. 1983. J. Gen. Virol. 64:169-176.</li></ol>
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**Remarks**

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