

<b>Virus Name: Kowanyama</b>		<b>Abbreviation: KOWV</b>
Status <b>Possible Arbovirus</b>	Select Agent <b>No</b>	SALS Level <b>2</b>
SALS Basis <b>Results of SALS surveys and information from the Catalogue.</b>		
Other Information		
Antigenic Group <b>Ungrouped</b>		

**SECTION I - Full Virus Name and Prototype Number**

Prototype Strain Number / Designation <b>MRM1178</b>	Accession Number	Original Date Submitted <b>12/5/1984</b>
Family <b>Bunyaviridae</b>	Genus <b>Bunyavirus-like</b>	
Information From <b>R.L. Doherty</b>	Address <b>Queensland Institute of Medical Research, Brisbane, Australia</b>	
Information Footnote <b>Reviewed by editor</b>		

**Section II - Original Source**

Isolated By (name) <b>Doherty, et al. (1)</b>	Isolated at Institute <b>Brisbane</b>	
Host Genus <b>Anopheles annulipes Walker</b>	Species	Host Age/Stage <b>Adult</b>
Sex <b>Female</b>		
<u>Isolated From</u>	<u>Isolation Details</u>	
Signs and Symptoms of Illness	Arthropod	
Time Held Alive before Inoculation		
Collection Method <b>Light trap</b>	Collection Date <b>11/13/1963</b>	
Place Collected (Minimum of City, State, Country) <b>Mitchell River Mission, Australia</b>		
Latitude <b>15° 30' S</b>	Longitude <b>141° 40' E</b>	
Macrohabitat <b>Low-lying plain bordering Gulf of Carpentaria</b>	Microhabitat	Method of Storage until Inoculated <b>Transported to Brisbane on dry ice, then in -60dC Revco</b>
Footnotes		



**Morphogenesis**

Site of Constituent Formation in Cell	Site of Virion Assembly	Site of Virion Accumulation
Inclusion Bodies	Other	

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**Hemagglutination**

Hemagglutination <b>Yes</b>	Antigen Source <b>SMB, blood, liver, carcass ext. by sucrose-acetone + prot. tr.</b>	Erythrocytes (species used) <b>Goose</b>
pH Range <b>6.0-7.6</b>	pH Optimum <b>5.75, 6.0</b>	
Temperature Range	Temperature Optimum	

Remarks  
**Shows HA when VAD contains 0.4 M NaCl (8)**

Serologic Methods Recommended  
CF

Footnotes  
**Shows HA when VAD contains 0.4 M NaCl (8)**

Studies at QIMR, Brisbane: No relationship demonstrated by CF and neutralization test to MVE, Kunjin, Kokobera, Edge Hill, Stratford, Sindbis (MRM39), Ross River, Getah, Bebaru, Koongol, Wongal, Mapputta, Corriparta and Eubenangee (MRM1178 homologous CF = 64/32).

Studies at Yale University Department of Epidemiology and Public Health (Dr. J. Casals).

MRM1178 antigen (homologous titre = 128/64) tested by CF against antisera to the following viruses with no reactions found.

Group A polyvalent	Ilesha
Group B polyvalent	Uukuniemi
California encephalitis	Kairi
Colorado tick fever	Kemerovo
Hughes	Manzanilla
Wad Medani	Oropouche
Quaranfil	Silverwater
Chenuda	Simbu
Nyamanini	Thogoto
Eretmapodites 147	Tacaiuma
Mouse encephalomyelitis (GDI)	Tacaribe
Guaroa	Calovo
Mouse hepatoencephalitis	Turlock
Ganjam	Congo
IG673	Wyeomyia
Bhanja	California group polyvalent

Section VI - Biologic Characteristics

Virus Source (all VERTEBRATE isolates)

Lab Methods of Virus Recovery (ALL ISOLATIONS)  
Newborn mice

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
PS (CL)	P-4				4	1 mm	6.0* (1)	
Vero (CL)	P-Unk.				6	5 mm	6.0 (6)	
LLC-MK2 (CL)					3	1 mm	7.2 (6)	
PS (CL)			CPE (7)					
BHK-21 (CL)			CPE			Plaques (7)		
VSW (CL)			CPE			Plaques (7)		

\* Expressed in dex

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Anopheles annulipes	2/171		Queensland, Australia
Anopheles amictus amictus	1/69		Queensland, AS, Oct.-Nov. 1963
Man		0/69 NT	Queensland, AS
Domestic fowl		1/47 NT	
Horse		6/31 NT	
Kangaroos		5/11 NT	

Subsequent tests have shown that neutralizing antibody in man was only in adult Aborigenes from north Queensland (4,5), and more extensively in cattle, horses, sheep, pigs, kangaroos, wallabies, bandicoots, rats and wild birds (5).

**Section VIII - Susceptibility to Experimental Infection (include viremia)**

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log <sub>10</sub> /ml
Mice (nb)	P-4	ic 0.015	Death	5	6.8
Mice (nb)		ip 0.03	Death	9	3.5
Mice (nb)		sc			
Mice (wn)		ic 0.03	None		<3.5
Mice (wn)		ip	Production of antibody		
(nb)	P-12	ip 0.015	Death	4-5	7.5
(nb)		ip 0.03	Death	5-7	6.7

**Section IX - Experimental Arthropod Infection and Transmission**

Arthropod species & virus source(a)	Method of Infection log <sub>10</sub> /ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log <sub>10</sub> /ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System
Culex quinquefasciatus	Adult female intrathoracically inoculated with 0.0006 ml; virus multiplication demonstrated by titration in mice; 5.3/mosquito after inoculation; <1.3/mosq. at 12 hours; 5.0/mosq. at 10 days; 4.0/mosq. at 20 days. Serial passage of salivary gland by intrathoracic inoculation, first passage successful, second and third negative.								

**Section X - Histopathology**

Character of lesions (specify host)		
<u>Inclusion Bodies</u>	<u>Intranuclear</u>	
Organs/Tissues Affected		
Category of tropism		

**Section XI - Human Disease**

In Nature	Residual	Death
Subclinical	Overt Disease	
Clinical Manifestations		
Number of Cases	Category (i.e. febrile illness, etc.)	

**Section XII - Geographic Distribution**

Known (Virus detected) <b>Australia</b>
Suspected (Antibody only detected)

**Section XIII - References**

<ol style="list-style-type: none"><li>1. Doherty, R.L., et al. 1968. Trans. R. Soc. Trop. Med. Hyg. 62:430-438.</li><li>2. Standfast, H.A. and Carley, J.G. Personal communication.</li><li>3. Holmes, I.H. 1971. Personal communication.</li><li>4. Doherty, R.L. 1967. Aust. Paediat. J. 3:213-218.</li><li>5. Doherty, R.L., et al. 1970. Trans. R. Soc. Trop. Med. Hyg. 64:748-753.</li><li>6. Stim, T.B. 1969. J. Gen. Virol. 5:329-338.</li><li>7. Director, Queensland Inst. Med. Res. 1971. Unpublished observations.</li><li>8. Shope, R.E. Personal communication. 1977.</li></ol>
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**Remarks**

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