

<b>Virus Name: Kumlinge</b>		<b>Abbreviation: KUMV</b>
Status <b>Arbovirus</b>	Select Agent <b>No</b>	SALS Level <b>4</b>
SALS Basis <b>Placed in Level 4 based on the close antigenic relationship with a known Level 4 agent, Russian spring-summer encephalitis, plus insufficient laboratory experience.</b>		
Other Information <b>Vaccination Recommended</b>		
Antigenic Group <b>B</b>		

**SECTION I - Full Virus Name and Prototype Number**

Prototype Strain Number / Designation <b>A 52 (Prototype of Finnish strains of TBE)</b>	Accession Number	Original Date Submitted <b>12/25/1984</b>
Family <b>Flaviviridae</b>	Genus <b>Flavivirus</b>	
Information From <b>N. Oker-Blom</b>	Address <b>Department of Virology, University of Helsinki, Finland SF 00290</b>	
Information Footnote <b>Reviewed by editor</b>		

**Section II - Original Source**

Isolated By (name) <b>N. Oker-Blom, et al. (1)</b>	Isolated at Institute <b>Helsinki, Finland</b>	
Host Genus <b>Ixodes ricinus</b>	Species	Host Age/Stage <b>See below*</b>
Sex <b>Not Answered</b>		
<u>Isolated From</u>	<u>Isolation Details</u>	
Signs and Symptoms of Illness	Arthropod	
Time Held Alive before Inoculation		
Collection Method <b>Blanketing</b>	Collection Date <b>6/30/1959</b>	
Place Collected (Minimum of City, State, Country) <b>Kumlinge, Aland, SW Finland</b>		
Latitude <b>60° 15' N</b>	Longitude <b>20° 48' E</b>	
Macrohabitat <b>Deciduous forest-pasture</b>	Microhabitat <b>Ground vegetation of Corylus avellana grove</b>	Method of Storage until Inoculated <b>Alive at +22dC</b>
Footnotes		

**Section III - Method of Isolation**

Inoculation Date  
**7/18/1959**

Animal (Details will be in Section 6)  
**nb mice**

Route Inoculated <b>Intracerebral</b>	Reisolation <b>Yes</b>
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Other Reasons  
**Repeated isolations from the same area**

Homologous Antibody Formation by Source Animal

Test(s) Used

Footnotes

**Section IV - Virus Properties**

**Physicochemical**

Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density <b>1.23</b>	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

**Stability of Infectivity (effects)**

pH (infective range)

Lipid Solvent (ether - % used to test) <b>1:5; 20 hr, 4dC</b>	After Treatment Titer <b>4.4 dex</b>	Control Titer <b>7.6 dex</b>
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate) <b>1:1000</b>	After Treatment Titer <b>2.0 dex</b>	Control Titer <b>7.6 dex</b>
Other (formalin, radiation)		

**Virion Morphology**

Shape <b>Spherical</b>	Dimensions <b>36 nm</b>	
Mean nm	Range nm	
Measurement Method <b>Electron microscopy (2,3)</b>	Surface Projections/Envelope <b>Envelope observed; diameter = 3-5 nm (3);</b>	Nucleocapsid Dimensions, Symmetry

### Morphogenesis

Site of Constituent Formation in Cell                      Site of Virion Assembly                      Site of Virion Accumulation

Inclusion Bodies                      Other

### Hemagglutination

Hemagglutination                      Antigen Source                      Erythrocytes (species used)  
**Yes**                      **SMB ext. by sucrose-acetone\*\***                      **Goose**

pH Range                      pH Optimum  
**6.0-6.6**                      **6.2-6.4**

Temperature Range                      Temperature Optimum  
**4-37dC**                      **21dC (room)**

#### Remarks

**In density gradient at least two separable hemagglutinins, CF antigens and precipitins (3). \* Pool of 14 females, 12 males, 182 nymphes and 93 larvae \*\* Also aqueous-prot. ext.; cell culture fluid; other methods, see Reference 4.**

#### Serologic Methods Recommended

**NT, HI, antibody absorption, immunodiffusion in ag**

#### Footnotes

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### **Section V - Antigenic Relationship and Lack of Relationship to Other Viruses**

In neutralization and complement-fixation tests, the virus was shown to belong to the TBE complex [1] . By using antibody absorption and diffusion-in-gel techniques, Clarke has shown the Kumlinge (A 52) virus to be a Central European subtype of the tick-borne encephalitis virus [5] , [6] .

**Section VI - Biologic Characteristics**

Virus Source (all VERTEBRATE isolates)  
**Blood (LV)**

Lab Methods of Virus Recovery (ALL ISOLATIONS)  
**Newborn mice**

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
HeLa (CL)	MB 13 1-20.TC		SI. CPE	7.5 <sup>a</sup> (12)				
Hela (CL)	MB 2		CPE	9.7 (13)				
U cells (CL)	MB 13 1-10.TC		SI. CPE	7.0 <sup>b</sup> (12)				

<sup>a</sup> Titer of 9.0 by titration in mice; titer expressed in dex.

<sup>b</sup> Titer of 7.0 by titration in HeLa; 9.7 by titration in mice.

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Man	5 *		SW Finland (7)
Man		27/8,010 MI, NT, HI	Finland (8)
Endemic areas:			
Man		108/286 NT,HI	Kumlinge(SW Finland)
Man		21/305 NT,HI	Uukuniemi(SE Finland)
Cow		201/211 NT	Kumlinge, Finland
Cow		86/172 NT	Uukuniemi, Finland
Cow		13/6,585 NT,HI	Finland (9)
<i>Sciurus vulgaris</i>	5/9		Kumlinge, Finland(7)
<i>Microtus agrestis</i>	1/14		
<i>Lepus timidus</i>	1/1		Uukuniemi, Finland (7)
<i>Turdus merula</i>	2/9		Kumlinge, Finland (7)
<i>Turdus philomelos</i>	1/8		Uukuniemi, Finland (7)
<i>Emberiza citrinella</i>	1/10		Kumlinge, Finland (7)
<i>Ixodes ricinus</i>	19/12,001		SW, S, SE Finland (7)

\* From acute phase blood

**Section VIII - Susceptibility to Experimental Infection (include viremia)**

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST	Titer	
				(days)	log <sub>10</sub> /ml	
Mice (nb)	MB 3	ic 0.01	Paralysis and death	4	9.8	
Mice (nb)		ip 0.03	Paralysis and death	5	9.0	
Mice (nb)		sc				
Mice (wn)		ic 0.03	Paralysis and death	5	9.1	
Mice (wn)		ip 0.1	Paralysis and death	6	8.0	
guinea pig (2 mo)				Antibody formation		
chickens (7 days)		sc		Viremia up to 13 days(11)		6.6
chickens (ad.)		sc		Viremia in 2/12, 1/12 isolations from brain and gizzard after 2 years		4.3
Turdus merula (10 days)		MB 2	sc 100LD50	Viremia in 1/4, antibody formation		2.7
Rana temporaria (ad)		MB 3	sc 0.1	No virus detected		
Bufo bufo (ad)	sc 0.1		No virus detected			

**Section IX - Experimental Arthropod Infection and Transmission**

Arthropod species & virus source(a)	Method of Infection log <sub>10</sub> /ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log <sub>10</sub> /ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System

**Section X - Histopathology**

Character of lesions (specify host)	
<u>Inclusion Bodies</u>	<u>Intranuclear</u>
Organs/Tissues Affected	
Category of tropism	

**Section XI - Human Disease**

In Nature	Residual	Death
<b>Significant</b>		
Subclinical	Overt Disease	
Clinical Manifestations		
<b>Fever (S), headache (S), prostration (S), conjunctival inflammation (R), stiff neck (S), myalgia (R), CNS signs (including encephalitis)(S), CNS Pleocytosis (S), rash (R), vomiting (S) and biphasic fever</b>		
Number of Cases	Category (i.e. febrile illness, etc.)	
<b>About 5-20 annually (14,15)</b>		

**Section XII - Geographic Distribution**

Known (Virus detected)
<b>Finland (1,7,16)</b>
Suspected (Antibody only detected)

**Section XIII - References**

1. Oker-Blom, N., et al. 1962. In "Biology of viruses of the tick-borne Encephalitis Complex". II. 1960. (ed. H. Libikova). Academic Press Inc. New York. p. 423-429.
2. Weckström, P. and Nyholm, M. 1965. Nature 205:211-212.
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11. Brummer-Korvenkontio, M., et al. 1970. Scand. J. Clin. Lab. Invest. 25 Suppl.105;113.
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**Remarks**

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