

Virus Name: Lipovnik		Abbreviation: LIPV
Status Possible Arbovirus	Select Agent No	SALS Level 2
SALS Basis Results of SALS surveys and information from the Catalogue.		
Other Information		
Antigenic Group Kemerovo		

SECTION I - Full Virus Name and Prototype Number

Prototype Strain Number / Designation Lip 91	Accession Number	Original Date Submitted 7/25/1984
Family Reoviridae	Genus Orbivirus	
Information From H. Libikova, M.D.	Address Institute of Virology, Slovak Ac. Sci., 80939 Bratislava, Czechoslovakia	
Information Footnote Revised		

Section II - Original Source

Isolated By (name) Libikova, et al. (1,2,4)	Isolated at Institute Lipovnik, Czechoslovakia	
Host Genus Ixodes ricinus	Species	Host Age/Stage Imago
Sex Female		
<u>Isolated From</u>	<u>Isolation Details</u>	
Signs and Symptoms of Illness	Arthropod Depleted	
Time Held Alive before Inoculation 7 days		
Collection Method Woolen flag	Collection Date 5/25/1963	
Place Collected (Minimum of City, State, Country) Lipovnik village, East Slovakia, Czechoslovakia		
Latitude 48° 35' N	Longitude 20° 30' E	
Macrohabitat Mixed forests	Microhabitat	Method of Storage until Inoculated Ticks stored at +4dC
Footnotes		

Section III - Method of Isolation

Inoculation Date
6/7/1963

Animal (Details will be in Section 6)
(Tissue Culture)

Route Inoculated _____ Reisolation
Yes

Other Reasons
No similar viruses were present in the laboratory

Homologous Antibody Formation by Source Animal

Test(s) Used _____

Footnotes _____

Section IV - Virus Properties

Physicochemical
RNA, Single Strand

Pieces (number of genome segments) 10	Infectivity No	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

Stability of Infectivity (effects)

pH (infective range)
Stable at pH 6-9, inactivated at pH 3-5.

Lipid Solvent (ether - % used to test) 20%	After Treatment Titer 3.0 dex	Control Titer 5.5 dex
Lipid Solvent (chloroform) 5%	After Treatment Titer loss of 5 dex	Control Titer
Lipid Solvent (deoxycholate) 0.1%	After Treatment Titer 0.5 dex	Control Titer 5.0 dex

Other (formalin, radiation)
Sensitive to interferon (9); resistant to 6-azauridine (9,11)

Virion Morphology

Shape Spherical	Dimensions 70 nm	
Mean nm	Range nm	
Measurement Method Electron microscopy	Surface Projections/Envelope None	Nucleocapsid Dimensions, Symmetry

Morphogenesis

Site of Constituent Formation in Cell
Cytoplasm

Site of Virion Assembly
Area of endoplasmic reticulum (9)

Site of Virion Accumulation

Inclusion Bodies

Other

Hemagglutination

Hemagglutination
No

Antigen Source
SMB ext. by sucrose-acetone

Erythrocytes (species used)
Goose

pH Range
5.75-7.2

pH Optimum

Temperature Range
4dC - 20dC

Temperature Optimum

Remarks

Other sources for antigen prod.: infant rat brain, CE cell cult. fluid; all were neg.

Serologic Methods Recommended
CF, NT (12,13)

Footnotes

Other sources for antigen prod.: infant rat brain, CE cell cult. fluid; all were neg.

No relationship: [1] TBE, JBE, SLE, WEE, EEE, Sindbis, Colorado tick fever, pseudorabies virus, Newcastle disease virus.

Dr. J. Casals (personal communication) tested the Lipovnik antigen in dilutions 1:4, 1:16 and 1:64 in complement-fixation against antisera for the following viruses: polyvalent groups A and B, California encephalitis, Colorado tick fever, Hughes, Quarantil, Nyamanini, Eretrmapodites 147, Guaroa, Ganjam, Bhanja, Ilesha, Uukuniemi, Kairi, Manzanilla, Oropouche, Silverwater, Simbu, Thogoto, Tacaiuma, Tacaribe, Calovo, Turlock, Wyeomyia, Congo, lymphocytic choriomeningitis, mouse encephalomyelitis (GD 1) and mouse hepatoencephalitis. The sera were used in dilution 1:4 through 1:128. None of the above sera reacted positively with Lipovnik antigen.

NT among Kemerovo group viruses by ic route in mice (data of J. Casals). Protection expressed in dex as LNI.

Serum Virus	Species	Virus			
		KEM	LIP	TRB	CNU
Kemerovo	Mouse	5.5	1.7	1.7	0.6
	Rat	3.6	0.6	0.5	-0.1
Lipovnik	Mouse	2.2	3.6	2.5	0.5
	Rat	1.9	3.8	2.6	0.0
	Rabbit	0.8	2.4	1.3	-0.1
Tribec	Mouse	0.7	1.6	2.5	
	Rat	0.7	2.4	3.3	0.1
	Rabbit	0.7	2.0	4.4	0.1
Chenuda	Rat	-0.8	0.5	0.3	4.0
	Rabbit	-0.1	0.5	0.7	3.7

Closest relationship shown to Koliba (Western Slovakia, Lat. 48/10'N., Long. 17/10'E) and Tribec viruses and also to Kemerovo virus. Related to other members of the Kemerovo group ([1], [2], [8], [10], [11]). Also see Table 2 from References [12] and [13]. On other related Eurasian isolates, see Reference [19]. Strain Cvilin [18] isolated in North Moravia near Ostrava is closely related or identical with CIP virus.

Section VI - Biologic Characteristics

Virus Source (all VERTEBRATE isolates)
CNS (LV), heart (LV), spleen (LV) (pooled tissues)

Lab Methods of Virus Recovery (ALL ISOLATIONS)
Newborn mice

Cell system (a)	Virus passage history (b)	Evidence of Infection						Growth Without CPE +/- (g)
		CPE			PLAQUES			
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
Chick embryo (PC)	CEC 3, emb. egg Pass 6 SMB 1-3	1-3	4+	7-8*	3-4	1-5 mm	7-8* (1,9)	
BHK-21 (CL)		1-3	4+	8.5 (13)				
Vero (CL)		2-3	4+	7.5-8.5	4-5	1-4 mm	7-8.5 (13)	
L cells (CL)		2-3	4+	5.0 (11)				
Hyalomma dromedarii (PC)			No CPE			No plaques		+ (16) 10-14 days multiplied 10,000-fold

* Expressed in dex

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Ixodes ricinus Man	8/1,060 (106 pools)	18% pos. NT	Lipovnik village, East Slovakia, CZ (1, 5)
Ixodes ricinus	3/1,540 (154 pools)		Koliba Hill, West Slovakia, CZ (1, 5)
Ixodes ricinus Man	1/680 (68 pools)	8.5%/711 NT 9.4%/32 NT	Civilin forest, North Moravia, CZ (18) Kronv, Czechoslovakia (19) Albrechtice, Czechoslovakia (19)

Section VIII - Susceptibility to Experimental Infection (include viremia)

Experimental host and age	Passage history and strain	Inoculation Route-Dose		Evidence of infection	AST (days)	Titer log ₁₀ /ml
Mice (nb)	CEC 3	ic	0.01	Fatal encephalitis	2-3	8.0
Mice (nb)	emb. eggs 1-2,	ip	0.05	Viremia		
Mice (nb)	SMB 1-3	sc	0.05	Viremia (7)		
Mice (wn)		ic	0.03	Discrete encephalitis		
Mice (wn)		ip	0.1	Formation of antibodies		
rats, hamsters (nb)		ic	0.03	Fatal encephalitis	2-3	8.0
rats, hamsters (ad)		ic	0.05	No deaths		
guinea pigs, rabbits (subadult)		ic	0.05	No deaths (3)		
embryonated eggs (7-8 day)	Lip 91, Lip 25, Koliba 154, 156, CEC 3 emb. egg 6	ys or al.c.		Death in 2-3 days (9)		5-7

Section IX - Experimental Arthropod Infection and Transmission

Arthropod species & virus source(a)	Method of Infection log ₁₀ /ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log ₁₀ /ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System

Section X - Histopathology

Character of lesions (specify host)

Nb mice, ic: acute necrotizing encephalitis. Wn mice: degeneration of the ventricular ependymal lining, inflammatory infiltration in the subependymal areas and around vessels in the white and gray matter (3).

Inclusion Bodies

Intranuclear

Lower Vertebrates

Organs/Tissues Affected

Brain (LV); skin, intestines, lungs, metanephron (LV) (3,9)

Category of tropism

Neurotropic

Section XI - Human Disease

In Nature
Reported

Residual

Death

Subclinical
Reported

Overt Disease

Clinical Manifestations

Meningitis; Involved in the pathogenesis of overt tick-borne encephalitis as concurrent infection (18,19,20). CSF antibodies detected in 50% of multiple sclerosis cases (19).

Number of Cases

8

Category (i.e. febrile illness, etc.)

Febrile illness, meningitis

Section XII - Geographic Distribution

Known (Virus detected)

Czechoslovakia

Suspected (Antibody only detected)

Section XIII - References

1. Libikova, H., et al. 1965. Acta Virol. 9:76-82.
2. Libikova, H., et al. 1964. Acta Virol. 8:96.
3. Libikova, H., et al. 1965. Acta Virol. 9:423-430.
4. Libikova, H., et al. In: Proceedings of a Symposium held in 1963. Ed. B. Rosicky, K. Heyberger. Cz. Ac. Sci. 1965. pp. 429-100%.
5. Libikova, H. 1964. Lek Obzor 13:607-614.
6. Libikova, H. and Sokol, F. 1966. Acta Virol. 10:551-553.
7. Libikova, H. 1966. Acta Virol. 10:554-556.
8. Gresikova, M., et al. 1965. Acta Virol. 9:83-88.
9. Libkova, H. 1970. Acta Virol. 14:217-228.
10. Casals, J. In: International Symposium on Tick-borne Arboviruses excluding Group B. Gresikova, M., ed. Publ. House Slovak Ac. Sci. 1971. pp. 13-20.
11. Libikova, H. In: International Symposium on Tick-borne Arboviruses excluding Group B. Gresikova, M., ed. Publ. House Slovak Ac. Sci. 1971. pp. 53-58.
12. Libikova, H. and Casals, J. 1971. Acta Virol. 15:65-78.
13. Libikova, H. and Buckley, S.M. 1971. Acta Virol. 15:79-86.
14. Libikova, H., et al. 1970. Acta Virol. 14:64-69.
15. Zavadova, Z. and Libikova, H. 1975. Acta Virol. 19:88-90.
16. Libikova, H., et al. 1974. Cs. Epidem. Mikrobiol. Immunol. 23:332-340.
17. Libikova, H. and Rajcani, J. 1975. Acta Virol. 19:1-9.
18. Libikova, H., et al. 1977. Cs. Epidem. Mikrobiol. Immunol. 26:135-138.
19. Libikova, H., et al. 1978. Med. Microbiol. Immunol. 116:255-263.
20. Libikova, H. and Zavada, J. In: Biologicke Prace 22/2. Publ. House Slovak Ac. Sci. 1976. pp. 84-95.

--