

<b>Virus Name: Llano Seco</b>		<b>Abbreviation: LLSV</b>
Status <b>Probable Arbovirus</b>	Select Agent <b>No</b>	SALS Level <b>3</b>
SALS Basis <b>Insufficient experience with virus; i.e., experience factor from SALS surveys was less than 500 in laboratory facilities with low biocontainment.</b>		
Other Information		
Antigenic Group <b>*</b>		

**SECTION I - Full Virus Name and Prototype Number**

Prototype Strain Number / Designation <b>BFN 3112</b>	Accession Number	Original Date Submitted <b>10/11/1984</b>
Family <b>Reoviridae</b>	Genus <b>Orbivirus</b>	
Information From <b>Robert P. Scrivani</b>	Address <b>Arbovirus Res. Lab., Schl. of Public Health, Univ. of California, Berkeley, CA</b>	
Information Footnote <b>Reviewed by editor</b>		

**Section II - Original Source**

Isolated By (name) <b>Dr. W.C. Reeves</b>	Isolated at Institute <b>Arbovirus Lab, Berkeley, CA</b>	
Host Genus <b>Culex tarsalis, pool of 50</b>	Species	Host Age/Stage <b>Adult</b>
Sex <b>Female</b>		
<u>Isolated From</u>	<u>Isolation Details</u>	
Signs and Symptoms of Illness	Arthropod <b>Gravid</b>	
Time Held Alive before Inoculation		
Collection Method <b>Aspiration from shelter</b>	Collection Date <b>7/23/1971</b>	
Place Collected (Minimum of City, State, Country) <b>Llano Seco Ranchero, Butte Co., CA</b>		
Latitude <b>39° N</b>	Longitude <b>122° W</b>	
Macrohabitat <b>Riparian area with livestock pens</b>	Microhabitat <b>Tule swamp</b>	Method of Storage until Inoculated <b>Revco storage at -70dC</b>
Footnotes		



### **Morphogenesis**

Site of Constituent Formation in Cell	Site of Virion Assembly	Site of Virion Accumulation
Inclusion Bodies	Other	

### **Hemagglutination**

Hemagglutination <b>No</b>	Antigen Source <b>Duck embryo cell culture; SMB ext. by sucrose- acetone</b>	Erythrocytes (species used) <b>Goose</b>
pH Range <b>5.8-7.0</b>	pH Optimum	
Temperature Range	Temperature Optimum	

#### Remarks

**\* Although it has been demonstrated that Llano Seco virus is antigenically related to Umatilla virus, its antigenic relationship to other established orbivirus serogroups is uncertain. Further serological studies are needed to clarify i**

Serologic Methods Recommended  
**CF, NT, IFA (broadly reacting)**

#### Footnotes

**\* Although it has been demonstrated that Llano Seco virus is antigenically related to Umatilla virus, its antigenic relationship to other established orbivirus serogroups is uncertain. Further serological studies are needed to clarify i**

Hemagglutination by WEE, SLE, POW, MOD, RB, CE, LOK, BUT, MD, TUR, and VEE antigens was not inhibited by BFN 3112 hyperimmune mouse serum (HIMS). There was no neutralization of BFN 3112 virus by plaque-reduction neutralization test using 1:5 dilutions of polyvalent grouping ascitic fluids for Group C, Group Guama, Group Simbu, Group VSV, Group Bunyamwera, Group California, Group Tacaribe, Group Phlebotomus, Group A, Polyvalent Quarantil, Poly. Anopheles A, Poly. Bwamba, Poly. Patois, Group Capim, Group B, Poly. Palyam, Group Kemerovo, Poly. Congo, Polyvalents 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 12, and rabies. The homologous PRNT titer with BFN 3112 virus and HIMS was >80. BFN 3112 virus is related to Umatilla virus by cross-indirect fluorescent antibody test (IFA). By CF test, BFN 3112 antigen (homologous serum titer 32) reacted with Umatilla HIMS at 1:4 and 1:8 serum dilutions, but Umatilla antigen was nonreactive even homologously. BFN 3112 antigen did not react by CF with grouping ascitic fluids for Group Kemerovo and Polyvalent 8, but showed a questionable reaction with a 1:4 dilution of Polyvalent Palyam ascitic fluid. Also, see editor's remarks in footnote, page 1. Llano Seco antigenically related to Umatilla and a new unregistered orbivirus, Netivot [2]. They now comprise the Umatilla serogroup. Complement-fixation and plaque-reduction neutralization test results with Llano Seco virus and antibody to some orbiviruses.

Antibody <sup>a</sup>	Complement- fixation	Plaque-reduction neutralization test
Llano Seco	32 <sup>b</sup>	> 160 <sup>b</sup>
Umatilla	8	<5
Bluetongue 8	<4	<5
CV 73 <sup>c</sup>	32	<5
V5-4387 <sup>d</sup>	<4	<5
Polyvalent 8 <sup>e</sup>	<4	<5
Polyvalent Palyam <sup>e</sup>	4	<5
Kemerovo Group <sup>e</sup>	<4	<5

<sup>a</sup> Hyperimmune mouse sera or ascitic fluid except Bluetongue 8 which is guinea pig serum obtained from Lederle Laboratories.

<sup>b</sup> Reciprocal of antibody dilution giving 3+ to 4+ fixation or 80% reduction of a viral challenge of 60-70 PFU.

<sup>c</sup> Immune mouse ascitic fluid obtained from Dr. H.M.S. Watkins, Naval Biosciences Laboratory, Oakland, CA prepared against an unidentified agent isolated by Dr. Watkins.

<sup>d</sup> Hyperimmune mouse serum obtained from Dr. R. Emmons, Calif. State Department of Public Health, prepared against unidentified agent recovered in his laboratory.

<sup>e</sup> Reference grouping orbiviral reagents obtained from N.I.A.I.D.

**Section VI - Biologic Characteristics**

Virus Source (all VERTEBRATE isolates)

Lab Methods of Virus Recovery (ALL ISOLATIONS)  
Newborn mice

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
Duck embryo (CL)	SM6	3	4+	7.5 (f)	3	1-2 mm	6.6 (f)	
Vero (CL)						No plaques		
PS-Y-15 (CL)					3	1-2 mm	5.6	
Culex tarsalis (CL)			No CPE					+

(f) Expressed in dex

**Section VII - Natural Host Range (Additional text can be added below table)**

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Culex tarsalis	12/thousands		Sacramento Valley, CA, USA
None from thousands of samples of other mosquito species, wild avian and mammalian bloods collected in the Central Valley of California.			

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log <sub>10</sub> /ml
Mice (nb)	SM 6	ic 0.01	Sickness and death	3	
Mice (nb)		ip			
Mice (nb)		sc			
Mice (wn)		ic 0.03	None*		
Mice (wn)		ip 0.10	None*		
		4.0 dex PFU			
rabbits		sc 0.10	None**		
hamsters		sc 0.10	None**		
guinea pig		sc 0.10	None**		
1-week old chick		sc 0.10	None**		

\* Antibody present 7 days after the second ic injection and 7 days after the fourth ip injection given at 7 day intervals.

\*\* Tested for viremia at 1-7 days after inoculation and for PRNT antibody at 21 of 35 days after inoculation.

**Section IX - Experimental Arthropod Infection and Transmission**

Arthropod species & virus source(a)	Method of Infection log10/ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log10/ml (e)			
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System	
Culex tarsalis, SM6	6.6	6.2	10	26C	Drop of blood(1)	10/14	7.8		Duck embryo	
			7							
		6.0	14	26C						13/22
			21							14/25
Aedes dorsalis, SM6		5.0	12	26C			3.3		Duck embryo	
							>3.0		Duck embryo	

**Section X - Histopathology**

Character of lesions (specify host)

Inclusion Bodies

Intranuclear

Organs/Tissues Affected

Category of tropism

**Section XI - Human Disease**

In Nature

Residual

Death

Subclinical

Overt Disease

Clinical Manifestations

Number of Cases

Category (i.e. febrile illness, etc.)

**Section XII - Geographic Distribution**

Known (Virus detected)  
**Sacramento Valley, CA, USA**

Suspected (Antibody only detected)

**Section XIII - References**

1. Gubler, D.J. and Rosen, L. 1976. Am. J. Trop. Med. Hyg. 25:146-150.
2. Tesh, R.B., et al. 1986. Am. J. Trop. Med. Hyg. 35:418-428.

**Remarks**