

Virus Name: Murray Valley encephalitis		Abbreviation: MVEV
Status Arbovirus	Select Agent No	SALS Level 3
SALS Basis Results of SALS surveys and information from the Catalogue.		
Other Information DOC Permit Required		
Antigenic Group B		

SECTION I - Full Virus Name and Prototype Number

Prototype Strain Number / Designation MVE/1/1951	Accession Number	Original Date Submitted 3/24/1985
Family Flaviviridae	Genus Flavivirus	
Information From Eric L. French, Ph.D.	Address CSIRO Animal Health Laboratory, Private Bag No. 1, Parkville, N.2, Victoria, Australia	
Information Footnote Reviewed by editor		

Section II - Original Source

Isolated By (name) Dr. E.L. French (1)	Isolated at Institute Walter and Eliza Hall Inst., Melbourne	
Host Genus Human	Species	Host Age/Stage 19 years
Sex Male		
<u>Isolated From</u>	<u>Isolation Details</u>	
Organs/Tissues	Pooled specimens of cerebellum, pons, medulla, and cerebrum	
Signs and Symptoms of Illness Acute encephalitis*	Arthropod	
Time Held Alive before Inoculation		
Collection Method At necropsy	Collection Date 2/9/1951	
Place Collected (Minimum of City, State, Country) District Base Hospital, Mooropna, Victoria, AS		
Latitude 36° 20' S	Longitude 145° 20' E	
Macrohabitat	Microhabitat	Method of Storage until Inoculated Sealed in tubes on solid carbon dioxide
Footnotes		

Section III - Method of Isolation

Inoculation Date
2/10/1951

Animal (Details will be in Section 6)
(Embryonated Egg)

Route Inoculated
CAM

Reisolation
Yes

Other Reasons

Four similar viruses isolated during epidemic of 1951; 36% NT antibody rate in area.

Homologous Antibody Formation by Source Animal
Yes

Test(s) Used
CF, NT

Footnotes

Section IV - Virus Properties

Physicochemical
RNA

Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
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Percentage wt, of Virion Protein	Lipid	Carbohydrate
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Virion Polypeptides: Number	Details 7.8% nucleic acid; 11% lipid (4); 10% MB virus: 7 dex to 2.5 dex, 20 min, 50C (5); in 10% rabbit serum inact. at 56C for 15 min.(3)
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Non-virion Polypeptides: Number	Details
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Virion Density	Sedimentation Coefficients(s) (S)
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Nucleocapsid Density	Sedimentation Coefficients(s) (S)
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Stability of Infectivity (effects)

pH (infective range)

Lipid Solvent (ether - % used to test) 2:1	After Treatment Titer 3.6 dex	Control Titer 7.0 dex
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Lipid Solvent (chloroform) 2:1	After Treatment Titer 1.8 dex	Control Titer 7.0 dex
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Lipid Solvent (deoxycholate) 0.1%	After Treatment Titer 2.8 dex	Control Titer 6.9 dex
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Other (formalin, radiation)

Butanol 2:1: 7.6 to 0 dex; Phospholipase A 10 mcg/ml: 7.0 to 2.3 dex (6)

Virion Morphology

Shape	Dimensions 20-50 nm
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Mean nm	Range nm
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Measurement Method
Filtration (3); electron microscopy
25-35 nm (4)

Surface Projections/Envelope

Nucleocapsid Dimensions,
Symmetry

Morphogenesis

Site of Constituent Formation in
Cell

Site of Virion Assembly

Site of Virion Accumulation

Inclusion Bodies

Other

Hemagglutination

Hemagglutination
Yes

Antigen Source
SMB susp. as crude hemagglutinin

Erythrocytes (species used)
Goose

pH Range

pH Optimum
6.5-6.75

Temperature Range

Temperature Optimum
4dC

Remarks

* Clinically indistinguishable from Australian X disease, JBE, SLE, or EEE and WEE in humans (2).

Serologic Methods Recommended
NT, CF, HI

Footnotes

* Clinically indistinguishable from Australian X disease, JBE, SLE, or EEE and WEE in humans (2).

The three viruses isolated in Melbourne in 1951 from MVE Cases were sent to the Army Medical Service Graduate School at Washington, D.C., because, due to quarantine regulations, the full range of type arboviruses were not available in Australia. The evidence, based on CF and NT tests, indicating that MVE virus is a member of Group B arboviruses, is from reference [9].

Cross Neutralization Tests [9] :					
Rabbit Immune Serum	Neutralizing indices (dex)				
	Test viruses				
	JE	MVE/1/51	WN	SLE	WEE
Japanese Enc.	4.7	1.4	0.5	0.9	0.9
MVE/1/51	1.9	3.6	1.0	1.6	0
WN	1.7	1.0	3.5	1.2	0
SLE	0.9	0.6	1.0	2.4	0
WEE	0.9	1.0	0.1	0.1	3.6

Cross-Complement Fixation (9):					
Hyperimmune GP Serum	Antigen 1:4				
	JE	MVE/1/51	WN	SLE	WEE
JE	128	32	0	0	0
MVE/1/51	8	64	0	0	0
WN	0	16	64	0	0
SLE	8	16	0	16	0
WEE	0	0	0	0	64

The identification of the New Guinea isolate MVE/1/56 is given in Reference [11]. As far as is known it is not related to any virus not in Group B.

In a cross-neutralization study involving 42 flaviviruses, MVE virus was antigenically placed in a subgroup or complex consisting of JBE, WN, SLE, KUN, USU, KOK, STR, and ALF viruses [25].

Section VI - Biologic Characteristics

Virus Source (all VERTEBRATE isolates)
Blood (LV), pool liver, kidney (LV)

Lab Methods of Virus Recovery (ALL ISOLATIONS)
Newborn mice

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
Chick embryo(PC)	MB 16				5	3 mm (16)		
PS(CL)	P-9	3	4+	7.0**	4	2.5- 3.5mm	8.2**(26)	
Vero(CL)	P-15				2	9 mm	6.7(27)	
LLC-MK2 (CL)					4	2 mm	8.2(27)	

In addition, may other animal cell lines and mosquito cell cultures are susceptible.

** Expressed in dex

Section VII - Natural Host Range (Additional text can be added below table)

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Man	3/16		Victoria and New South Wales, Australia (1)
Man		109/1,524 CF	Northern Victoria, Australia (12)
Man	1/1		South Australia (10)
Man		15/55 CF,NT	Australia (12)
Man	1		NW Australia (23)
Man	1/1	140/485 NT	New Guinea (22)
Domestic animals			

Horses		31/62 CF	Northern Victoria, Australia (12)
Dogs		5/10 CF	
Foxes		5/9 CF	Mildura, Victoria, Australia (12)
Opossum		2/3 CF	
Birds			
Water birds		40/99 NT	
Land birds		11/60 NT	
Domestic chicks		31/56 NT	
Arthropods			
Cx annulirostris	5/8,963		Mitchell River Mission Australia (13)
Ae normanensis	1/904		
Cx bitaeniorhynchus	1/682		
Sentinal chickens	2		Australia (28)
Whitefaced heron (blood)	1		Australia (29)

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log ₁₀ /ml
Mice (nb)	MB 5	ic 0.03	Death	7	8.5
Mice (nb)		ip 0.03	Death	7	8.5
Mice (nb)		sc			
Mice (wn)	MB 11	ic 0.03	Death	7	7.7
Mice (wn)		ip 0.03	Death	13	<1.0
monkey (ad)	MB 4	ic 0.5	Death (1)	13	5.0
sheep (1-6 wk)		ic 0.5	Death (1)	7	3.0
chickens (2 day)	MB 5	sc 0.1	Viremia after 48 hours		11.2
chickens (4 day)		oral 0.1	Same		3.0
chickens (28 day)		sc 0.1	Same		
chick embryo (10-12 day)	MB 4	CAM 0.05	Specific pocks (1)	1.75	6.0
chick embryo (10-12 day)		all 0.05	Death (1)	2	5.5
chick embryo (5-8 day)		ys 0.1	Death (1)	2.2	5.5
guinea pigs (1-12 wk)		ic 0.1	Temp rise (1)		
hamsters (6-10 wk)	MB 12	ic,ip,sc 0.1	Death (14)		9.1- 8.0

Section IX - Experimental Arthropod Infection and Transmission

Arthropod species & virus source(a)	Method of Infection log10/ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log10/ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System
Cx annulirostris Cx quinquefasciatus Ae occidentalis Ae vigilax	Adults fed on infected blood; transmitted infection by bite to chicks; virus content in mosquitoes titrated on CAM (15)								
Ae aegypti	Transovarial transmission following oral infection (24)								

Section X - Histopathology

Character of lesions (specify host)
Brains from fatal infections in man and monkey show perivascular cuffing, neuronal degeneration, neuronophagia and necrosis in brains and cords. Purkinje cells destroyed. Plaques of encephalomalacia in brains of patients dying late in disease. (17, 18)

Inclusion Bodies Intranuclear

Organs/Tissues Affected
Brain (M)(LV), spinal cord (M), skeletal muscles (LV)

Category of tropism
Neurotropic

Section XI - Human Disease

In Nature Significant	Residual Significant	Death Significant
Subclinical Reported	Overt Disease	
Clinical Manifestations Fever (S), headache (S), stiff neck (S), myalgia (R), CNS signs (including encephalitis)(S), respiratory involvement (R), leukopenia (R), CNS pleocytosis (R), rash (R), and vomiting (S)		
Number of Cases 44 described; 40 (20,2), 4 (10,11,18,23)	Category (i.e. febrile illness, etc.) Encephalitis	

Section XII - Geographic Distribution

Known (Virus detected)
Australia (1,10,13,18), New Guinea (11),

Suspected (Antibody only detected)

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12. ANDERSON, S.G. 1954. J. Hyg. 52:447.
13. DOHERTY, R.L., et al. 1963. Aust. J. Exp. Biol. Med. Sci. 41:17-40.
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16. AUSTIN, F.J. 1963. Aust. J. Exp. Biol. Med. Sci. 41:205-209.
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25. DE MADRID, A.T. and Porterfield, J.S. 1974. J. Gen. Virol. 23:91-96.
26. WESTAWAY, E.G. 1966. Am. J. Epidem. 84:439-456.
27. STIM, T.B. 1969. J. Gen. Virol. 5:329-338.
28. Doherty, R.L. 1977. Aust. J. Exp. Biol. Med. Sci. 55:103-130.
29. JCSMR, Canberra, Australia. 1975. Unpublished.
30. The Subcommittee on Arbovirus Laboratory Safety of The American Committee on Arthropod-Borne Viruses. 1980. Am. J. Trop. Med. Hyg. 29:1359-1381.

Remarks

MVE Virus, when suitably diluted, produces well defined pocks on the CAM of 10-12 day chick embryos. In this situation the virus is not neutralized by specific antibody and consequently inoculation of the CAM is the method of choice for the isolation of this virus from clinical material. Two of the 1951 isolations were made only on the CAM, in these instances mouse inoculations failed