

<b>Virus Name: Patois</b>		<b>Abbreviation: PATV</b>
Status <b>Arbovirus</b>	Select Agent <b>No</b>	SALS Level <b>2</b>
SALS Basis <b>Results of SALS surveys and information from the Catalogue.</b>		
Other Information		
Antigenic Group <b>Patois</b>		

**SECTION I - Full Virus Name and Prototype Number**

Prototype Strain Number / Designation <b>BT 4971</b>	Accession Number	Original Date Submitted <b>12/13/1984</b>
Family <b>Bunyavirus</b>	Genus	
Information From <b>Sunthorn Srihongse</b>	Address <b>Gorgas Memorial Laboratory, P.O. Box 2016, Balboa Heights, Canal Zone</b>	
Information Footnote <b>Reviewed by editor</b>		

**Section II - Original Source**

Isolated By (name) <b>Margaret A. Grayson (1)</b>	Isolated at Institute <b>Gorgas Memorial Laboratory, Panama</b>	
Host Genus <b>Sigmodon hispidus</b>	Species	Host Age/Stage <b>adult</b>
Sex <b>Male</b>		
<u>Isolated From</u> <b>Serum/Plasma</b>	<u>Isolation Details</u>	
Signs and Symptoms of Illness <b>presumably health wild animal</b>	Arthropod	
Time Held Alive before Inoculation		
Collection Method <b>baited live trap, heart blood taken by syringe</b>	Collection Date <b>6/16/1961</b>	
Place Collected (Minimum of City, State, Country) <b>two miles north of Almirante</b>		
Latitude <b>10° N</b>	Longitude <b>80° W</b>	
Macrohabitat <b>tropical rain forest (edge)</b>	Microhabitat <b>edge of fresh-water marsh</b>	Method of Storage until Inoculated <b>thermos (wet ice) for transportation. Revco at -65dC later</b>
Footnotes		

**Section III - Method of Isolation**

Inoculation Date  
**9/26/1961**

Animal (Details will be in Section 6)  
**nb mice**

Route Inoculated <b>ic and ip</b>	Reisolation <b>Not tried</b>
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Other Reasons  
**no other isolations of this virus have been made before**

Homologous Antibody Formation by Source Animal  
**Not tested**

Test(s) Used

Footnotes

**Section IV - Virus Properties**

**Physicochemical**

Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

**Stability of Infectivity (effects)**

pH (infective range)

Lipid Solvent (ether - % used to test) <b>1:2</b>	After Treatment Titer <b>&lt;2.0 dex</b>	Control Titer <b>4.6 dex</b>
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate)	After Treatment Titer	Control Titer

Other (formalin, radiation)

**Virion Morphology**

Shape	Dimensions <b>&lt;=50 nm</b>	
Mean nm	Range nm	
Measurement Method <b>filtration (Mexican strain) (6)</b>	Surface Projections/Envelope	Nucleocapsid Dimensions, Symmetry

**Morphogenesis**

Site of Constituent Formation in Cell

Site of Virion Assembly

Site of Virion Accumulation

Inclusion Bodies

Other

**Hemagglutination**

Hemagglutination

Antigen Source

Erythrocytes (species used)

**Yes****SM mouse serum, acetone ext.****goose**

pH Range

pH Optimum

**5.75-6.2****5.75**

Temperature Range

Temperature Optimum

**4dC-37dC****22dC**

Remarks

Serologic Methods Recommended

Footnotes

Immune Sera	Antigen of Registered Virus					Antigens	Immune Serum of Registered Virus				
	HI		CF		NT		HI		CF		NT
	Ht/Ho	Ratio	Ht/Ho	Ratio	Ht/Ho		Ht/Ho	Ratio	Ht/Ho	Ratio	Ht/Ho
Murutucu	20/640	1/32	0/16	0		Oriboca	40/2560	1/64	0/64	0	
Madrid	0/1280	0	0/64	0		Marituba	80/2560	1/32	0/64	0	
Zegla	20/640	1/32	32/128	1/4	2.2/>3.5	Mardri	40/2560	1/64	0/64	0	
Gr. C hyperimmune ascitic fluid * (does not include Patois and Zegla)	40					Zegla	80/2560	1/32	128/64	1/1	2.3/>3.6

\* Kindly supplied by Dr. Robert E. Shope

NT: LNI in dex

CF: Crude brain or liver antigens did not fix complement in the presence of mouse hyperimmune sera prepared with the following agents: VEE, EEE, WEE, Pixuna, Mayaro, Aura, Una, Guama, Guaroa, Wyeomyia, Kairi, Cache Valley and California.

HI: Acetone extracted serum antigen was not inhibited by hyperimmune sera or ascitic fluid prepared with the following agents: VEE, EEE, Pixuna, Mayaro, Aura, Ilheus, SLE, YF, Bussuquara, DEN-2, Cache Valley, Guaroa, Wyeomyia, Maguari, Kairi, California, Capim, Guajara, Oropouche, Turlock, Bwamba, Anopheles A and B, Tacaribe, Changuinola, Cocal, VS-Indiana, Sicilian sandfly fever, Icoaraci and Chagres viruses.

Section VI - Biologic Characteristics

Virus Source (all VERTEBRATE isolates)  
blood (LV)

Lab Methods of Virus Recovery (ALL ISOLATIONS)  
newborn mice

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
hamster kidney (PC)	SMB 4	6	CPE	4.0*				
Vero (CL)		6	CPE	5.0				
HeLa (CL)			CPE (6)					
Vero (CL)	SMB 6					No Plaques (7)		
LLC-MK2 (CL)					3	1 mm	8.3* (7)	

\* Expressed in dex

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Man	0/398	6/256 HI	Almirante, Bocas del Toro, Province, Panama
Sigmodon hispidus (cotton rat)	3/488	40/270 HI	
Proechimys semispinosus (spiny rat)	0/138	16/76 HI	
Birds	0/1,244		
Reptile	0/311		
Other wild vertebrates	0/52		
Sentinel mouse litters	1/647		
Arthropods	0/377,492		
Sentinel mice	7		Mexico (4)
Sentinel hamsters	9		Mexico,, Belize,, Guatamala (4)
Sentinel hamster	1		Almirante, Panama (8)
Culex sp.	2		Mexico (4)
Culex iolambdis	1		
Cx thriambus	2		
Cx opisthopus	1		

**Section VIII - Susceptibility to Experimental Infection (include viremia)**

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log <sub>10</sub> /ml	
mice (nb)	SMB 4	ic 0.02	illness and death	2.5	7.7	
mice (nb)		ip 0.02	illness and death	2.5	6.3	
mice (nb)		sc				
mice (wn)		ic 0.02	irregular deaths			
mice (wn)		ip 0.1	antibody formation			
guinea pigs				antibodies produced (6)		
rabbits				antibodies produced (6)		

**Section IX - Experimental Arthropod Infection and Transmission**

Arthropod species & virus source(a)	Method of Infection log <sub>10</sub> /ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log <sub>10</sub> /ml (e)			
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System	
	Hamsters infected in laboratory by bite of wild-caught <i>Culex vomerifer</i> (5).									

**Section X - Histopathology**

Character of lesions (specify host)	
<u>Inclusion Bodies</u>	<u>Intranuclear</u>
Organs/Tissues Affected	
Category of tropism	

**Section XI - Human Disease**

In Nature	Residual	Death
Subclinical	Overt Disease	
Clinical Manifestations		
Number of Cases	Category (i.e. febrile illness, etc.)	

**Section XII - Geographic Distribution**

Known (Virus detected)
Suspected (Antibody only detected)

**Section XIII - References**

1. Srihongse, S., et al. 1966. Am. J. Trop. Med. Hyg. 15:379-384.
2. Galindo, P., et al. 1966. Am. J. Trop. Med. Hyg. 15:385-400.
3. Srihongse, S. and Shope, R.E. 1968. Acta Virol. 12:453-456.
4. Scherer, W.F., et al. 1972. Am. J. Trop. Med. Hyg. 21:194-200.
5. Galindo, P. and Srihongse, S. 1967. Am. J. Trop. Med. Hyg. 16:525-530.
6. Zarate, M.L., et al. 1968. Am. J. Epidem. 88:273-286.
7. Stim, T.B. 1969. J. Gen. Virol. 5:329-338.
8. Srihongse, S., et al. 1967. Am. J. Trop. Med. Hyg. 16:519-524.

**Remarks**

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