

Virus Name: Piry		Abbreviation: PIRYV
Status <b>Possible Arbovirus</b>	Select Agent <b>No</b>	SALS Level <b>3</b>
SALS Basis <b>Results of SALS surveys and information from the Catalogue.</b>		
Other Information <b>USDA Permit Required</b>		
Antigenic Group <b>Vesicular Stomatitis</b>		

#### SECTION I - Full Virus Name and Prototype Number

Prototype Strain Number / Designation <b>BeAn 24232</b>	Accession Number	Original Date Submitted <b>1/27/1985</b>
Family <b>Rhabdoviridae</b>	Genus <b>Vesiculovirus</b>	
Information From <b>Belem Virus Lab.</b>	Address <b>Belem Virus Laboratory, Instituto Evandro Chagas, Belem, Para, Brazil</b>	
Information Footnote <b>Reviewed by editor</b>		

#### Section II - Original Source

Isolated By (name) <b>Belem Virus Lab.</b>	Isolated at Institute <b>Belem, Para, Brazil</b>	
Host Genus <b>Philander opossum</b>	Species	Host Age/Stage <b>Adult</b>
Sex <b>Male</b>		
<u>Isolated From</u>	<u>Isolation Details</u>	
<b>Organs/Tissues</b>	<b>Spleen and liver</b>	
Signs and Symptoms of Illness <b>None</b>	Arthropod	
Time Held Alive before Inoculation		
Collection Method <b>Trapped</b>	Collection Date <b>10/12/1960</b>	
Place Collected (Minimum of City, State, Country) <b>Utinga forest, Brazil</b>		
Latitude <b>1° 28' S</b>	Longitude <b>48° 27' W</b>	
Macrohabitat <b>Watershed forest</b>	Microhabitat <b>Ground level</b>	Method of Storage until Inoculated <b>Not stored</b>
Footnotes		

### Section III - Method of Isolation

Inoculation Date

**10/12/1960**

Animal (Details will be in Section 6)

**nb mice**

Route Inoculated

**Intracerebral**

Reisolation

**Yes**

Other Reasons

Homologous Antibody Formation by Source Animal

**Not tested**

Test(s) Used

Footnotes

### Section IV - Virus Properties

Physicochemical

Pieces (number of genome segments)

Infectivity

Sedimentation  
Coefficients(s)  
(S)

Percentage wt, of Virion Protein

Lipid

Carbohydrate

Virion Polypeptides: Number

Details

Non-virion Polypeptides: Number

Details

Virion Density

Sedimentation Coefficients(s)  
(S)

Nucleocapsid Density

Sedimentation Coefficients(s)  
(S)

#### Stability of Infectivity (effects)

pH (infective range)

Lipid Solvent (ether - % used to test)

After Treatment Titer

Control Titer

Lipid Solvent (chloroform)

After Treatment Titer

Control Titer

Lipid Solvent (deoxycholate)  
**1:1000**

After Treatment Titer  
**<4.5 dex**

Control Titer  
**>9.3 dex**

Other (formalin, radiation)

#### Virion Morphology

Shape

**Bullet-shaped**

Dimensions

**155 x 62 nm**

Mean  
nm

Range  
nm

Measurement Method

**Electron microscopy (1)**

Surface Projections/Envelope

Nucleocapsid  
Dimensions,

**Morphogenesis**

Site of Constituent Formation in Cell

Site of Virion Assembly

Site of Virion  
Accumulation

Inclusion Bodies

Other

**Hemagglutination**Hemagglutination  
**Yes**

Antigen Source

**Vero roller cultures (2), supernatant fluid Attempts with  
SMB, liver, serum tr. by acetone; sucrose-acetone; prot.,  
freon; and freon-n-heptane have failed.**Erythrocytes (species  
used)  
**Goose**

pH Range

pH Optimum  
**6.2**

Temperature Range

Temperature Optimum  
**4dC**

Remarks

Serologic Methods Recommended  
**HI, CF, NT**

Footnotes

Not related by CF to viruses isolated in Belem using sera for groups A, B, C, Guama; for complexes Capim, California, Wyeomyia; and specific sera for Cache Valley, Kairi, Guaroa, Oropouche, Tacaiuma, Mirim, Icoaraci, Candiru, Acara, Pacui, Irituia, Jurona.

CF tests with Piry antigen against antisera for 48 arboviruses and LCM showed no reactions. CF tests with Piry hyperimmune mouse serum against 62 arboviruses and Herpes simplex antigens were negative. In HI tests, Piry hyperimmune serum failed to inhibit HA of 33 arboviruses.

Serological relationship to other members of VSV group found as follows:

Immune Sera	Piry Antigen/Virus					Antigens/ Viruses	Piry Antiserum				
	HI		CF		NT		HI		CF		NT
	Ht/Ho	Ratio	Ht/Ho	Ratio	Ht/Ho		Ht/Ho	Ratio	Ht/Ho	Ratio	Ht/Ho
VSNJ			0/256	0	2.5/5.0+	VSNJ			0/128	0	0/5.7
VSI			0/256	0	2.3/6.2+	VSI			0/128	0	0/5.7
Cocal			0/256	0	2.6/7.0+	Cocal			0/128	0	1.7/5.7
Chandipura			8/128	1/16	3.4/5.7+	Chandipura			0/128	0	4.6/5.7
NT: LNI in dex											

## Section VI - Biologic Characteristics

Virus Source (all VERTEBRATE isolates)  
Serum (M), pool of spleen and liver (LV)

Lab Methods of Virus Recovery (ALL ISOLATIONS)  
Newborn mice

Cell system (a)	Virus passage history (b)	Evidence of Infection								
		CPE			PLAQUES			Growth Without CPE +/- (g)		
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)			
BHK-21(CL)	P-3				<4.0	Plaques	8.3*(4)			
Chick embryos(PC)					2-3	2 sizes	7.5(4)			
Turkey embryo(PC)					2-3	2 sizes	7.5(4)			
Mouse embryo(PC)							7.95(4)			
GMK (CL)						Plaques (4)				
Vero (CL)			CPE (2)							

\* Expressed in dex

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Man (blood; lab infection)	1	15/83 NT	Xingu, Brazil
Man		3%/31 NT	Rondonia, Brazil
Man		5/374 NT	Elsewhere, Brazil
Philander opossum	1		Para, Brazil (3)
Philander opossum		0/31 NT	Para and Amapa, Brazil
Other marsupials		16/337 NT	
Monkeys		8/104 NT	
Edentates		8/91 NT	
Rodents		13/508 NT	Para, Brazil
Bats		1/39 NT	
Birds		0/178 NT	
Pigs		4/178 NT	
Water buffalo		2/291 NT	
Horses, cattle		0/103 NT	
Man		105/261 *	Rio Grande do Sul, Brazil (7)
No isolation from arthropods			
* Rates increased with age			



Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log10/ml
Mice (nb)	P-3	ic 0.02	Death	1.3	10.7+
Mice (nb)		ip 0.02	Death	1.7	
Mice (nb)		sc			
Mice (wn)		ic 0.03	Death	3.8	
Mice (wn)		ip 0.03	Antibody		
white rats (ad)	Prototype	ip 0.2	Antibody		>20.0
guinea pigs (ad)		ip 0.2	Viremia, antibody		
hamsters (ad)		ip 0.2	Viremia, death	2.5	
Didelphis, Philander(ad)		im,sc 0.2	Viremia, death	4-12	

S-180 ascites tumor of mice destroyed by Piry virus, but virus titer potentiated (6).

Ponies, intralingual, elevated temp., ulcers at inoc. site; antibody production (8).

Steers, sheep, goats, intralingual, antibodies (8).

Pigs, inoc. in snout, heel, coronary band, antibodies (8).

## Section IX - Experimental Arthropod Infection and Transmission

Arthropod species & virus source(a)	Method of Infection log10/ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log10/ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System
Ae aegypti	By parenteral inoculation achieved 3 serial passages in salivary glands (5).								

## Section X - Histopathology

Character of lesions (specify host)

**Diffuse in connective tissue, liver and kidneys (mice). In a Didelphis, reticulohistiocytic hyperplasia, with long mononuclear cells, in focal areas of renal interstitial tissue, hepatic sinusoids, and red pulp of spleen (L.B. Dias).**

Inclusion Bodies

Intranuclear

Organs/Tissues Affected

Category of tropism

## Section XI - Human Disease

In Nature	Residual	Death
Subclinical	Overt Disease Significant	
Clinical Manifestations <b>Fever (S), headache (R), prostration (S), myalgia (S), arthralgia (S), and RUQ tenderness, loss of desire to smoke, anorexia</b>		
Number of Cases <b>6 (all laboratory infections)</b>	Category (i.e. febrile illness, etc.)	

## Section XII - Geographic Distribution

Known (Virus detected)

**Brazil**

Suspected (Antibody only detected)



### Section XIII - References

1. Bergold, G.H. and Munz, K. 1970. Arch. ges. Virusforsch. 31:152-167.
2. Director, YARU. Personal communication. 1968.
3. Woodall, J.P. 1967. Atas Simpos. Biota Amazon. 6:31-63.
4. Pinheiro, F.P. Personal communication.
5. Whitman, L. Personal communication.
6. Belem Virus Laboratory, Belem, Brazil. 1969. Unpublished data.
7. Pinheiro, F.P., et al. 1974. PAHO Bull. 8:111-122.
8. Wilks, C.R. and House, J.A. 1984. J. Hyg. 93:147-156".

### Remarks