

Virus Name: Puumala		Abbreviation: PUUV
Status Probably not Arbovirus	Select Agent No	SALS Level
SALS Basis		
Other Information USDA Permit Required, DOC Permit Required		
Antigenic Group Hantaan		

SECTION I - Full Virus Name and Prototype Number

Prototype Strain Number / Designation Sotkamo strain	Accession Number	Original Date Submitted 4/1/1985
Family Bunyaviridae	Genus Hantavirus	
Information From M. Brummer-Korvenkontio, A. Vaheri, C.H. vonBonsdorff	Address Department of Virology, University of Helsinki, Helsinki, Finland and J. Dalrymple, USAMRID, Frederick, MD, USA	
Information Footnote		

Section II - Original Source

Isolated By (name) M. Brummer-Korvenkontio, et al.(1)	Isolated at Institute Helsinki, Finland	
Host Genus Clethrionomys glareolus (bank vole)	Species	Host Age/Stage
Sex Not Answered		
<u>Isolated From</u>	<u>Isolation Details</u>	
Organs/Tissues	Lungs	
Signs and Symptoms of Illness None (fluorescent antigen in lungs by IFA when passed in voles)	Arthropod	
Time Held Alive before Inoculation		
Collection Method Trapped	Collection Date 11/26/1981	
Place Collected (Minimum of City, State, Country) Sotkamo, Finland		
Latitude 64° N	Longitude 29° E	
Macrohabitat Forest	Microhabitat Meadow	Method of Storage until Inoculated Kept alive until processed
Footnotes		

Section III - Method of Isolation

Inoculation Date
12/23/1981

Animal (Details will be in Section 6)
animal

Route Inoculated
ip, intrapulmonary

Reisolation
Yes

Other Reasons

Homologous Antibody Formation by Source Animal
Yes

Test(s) Used
Indirect IFA

Footnotes

Section IV - Virus Properties

Physicochemical
RNA, Single Strand

Pieces (number of genome segments) 3	Infectivity	Sedimentation Coefficients(s) L=2.7x106 MW; M=1.2x106 MW; S=0.6x106 MW(S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number 4	Details Nucleocapsid protein=52,000 MW; envelope glycoproteins: G1=64,000 MW, G2=58,000 MW; L protein=approx. 200,000 MW	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

Stability of Infectivity (effects)

pH (infective range)

Lipid Solvent (ether - % used to test)	After Treatment Titer	Control Titer
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate)	After Treatment Titer	Control Titer

Other (formalin, radiation)
Cobalt irradiation (1.3 x 10⁷ rads) inactivates infectivity

Virion Morphology

Shape	Dimensions	
Mean nm	Range nm	
Measurement Method	Surface Projections/Envelope	Nucleocapsid Dimensions, Symmetry

MorphogenesisSite of Constituent Formation in Cell
Cytoplasm

Site of Virion Assembly

Site of Virion Accumulation

Inclusion Bodies

Other

HemagglutinationHemagglutination
Yes

Antigen Source

Erythrocytes (species used)

Purified cell culture propagated virions**Gander**pH Range
5.8-6.8pH Optimum
5.8-6.2

Temperature Range

Temperature Optimum
37dC

Remarks

Serologic Methods Recommended
PRNT, IFA, HI, ELISA

Footnotes

Antigenic relations of HFRS-related viruses: PRNT were performed on Vero E6 cell monolayers with rat antisera and cell culture-adapted viruses. Approximately 100 PFU of virus were incubated with twofold dilutions of sera for 1 hour at room temperature before inoculation of 25-cm² flasks. Inoculated cells were further incubated for 1 hour at 37°C before addition of overlay containing 0.6% agarose (Seakem ME) and 10% heated fetal bovine serum (FBS) in the nutrient mixture. Following incubation for 7 to 14 days after infection, a second overlay identical to the first except for a reduced FBS concentration (2%) and inclusion of neutral red (final concentration 0.167 mg/ml) was applied. Plaques were counted as they became visible 1 to 5 days after addition of the second overlay. Titers were expressed as the reciprocal of the highest dilution of antibody, resulting in greater than 80% reduction of approximately 100 plaques.

Serum pools	Virus							
	HTN	LEE	UR	TCH	GP	SR-11	PH	PUU
HTN	4000	4000	0	160	80	40	0	0
LEE	4000	2000	20	160	160	200	0	0
UR	2000	80	4000	16000	16000	8000	160	0
TCH	2000	200	4000	32000	8000	8000	200	0
GP	200	80	2000	8000	8000	8000	200	0
SR-11	2000	20	2000	2000	2000	2000	200	0
PH	0	0	0	0	20	20	4000	0
PUU	0	0	0	0	0	0	0	1280

Tchoupitoulas, Girard Point, and Sapporo rat viruses are unregistered.

HTN = Hantaan virus (76-118)

LEE = Human isolate from Korea

UR = Seoul virus

TCH = Tchoupitoulas

GP = Girard Point

SR-11 = Sapporo rat

PH = Prospect Hill

PUU = Puumala

Section VI - Biologic Characteristics

Virus Source (all VERTEBRATE isolates)
Lungs (LV)

Lab Methods of Virus Recovery (ALL ISOLATIONS)
Passage in Vero E6 cell cultures and examination by IFA

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
E6 cells (CL)	Cleth. glar. P 0-3		No CPE	2-3* after passes				+
E6 cells (CL)	E6, P5		No CPE		12-18	1-2 mm	6-7*	
A549 (CL)	Cleth. glar. P 0-3		No CPE	1-2 after passes				+

* Expressed in dex

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Clethrionomys glareolus	3/48 *	155/549 IFA	"Lake Finland", Finland; 1977, 1979, 1981 (1-3)
Clethrionomys glareolus	1/25 **		West of Umea, Sweden;1981 (5, 6)
Clethrionomys rufocanus	1/19	1/19 IFA	Sweden (7)
Clethrionomys glareolus	Several		European USSR (9, 10)
Clethrionomys glareolus(lungs)	1		Turnkout, Antwerp Prov., Belgium (11)

* Strains Puumala (2,3), Hankasalmi (1), Sotkamo (1)

** Strain Hallnas

Section VIII - Susceptibility to Experimental Infection (include viremia)

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log10/ml
Mice (nb)		ic			
Mice (nb)		ip			
Mice (nb)		sc			
Mice (wn)		ic			
Mice (wn)		ip			
<i>Cl. glareolus</i>	Sotkamo P2	ip,sc intrapulmonary	Develop subclinical infection (FA antigen in lungs and liver tissue). Animals sacrificed day 30.		5
<i>Meriones unguiculatus</i> (nb) (Mongolian gerbils)	Hallnas;P-2	ic,ip, in,sc	Develop subclinical persistent infection. (Virus in brain, lungs, spleen, liver, pancreas, parotid, distal small intestine(8)		
Wistar female <i>Rattus norvegicus</i>			12-16 wks. old, infected by inhalation (12).		

Section IX - Experimental Arthropod Infection and Transmission

Arthropod species & virus source(a)	Method of Infection log10/ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log10/ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System

Section X - Histopathology

Character of lesions (specify host)

Inclusion Bodies

Intranuclear

Organs/Tissues Affected

Category of tropism

Section XI - Human Disease

In Nature
Significant

Residual

Death
Reported

Subclinical
Reported

Overt Disease
Significant

Clinical Manifestations

Fever, backache, headache, proteinuria, microscopical hematuria, oliguria to polyuria, hepatitis, sometimes encephalitis

Number of Cases

Category (i.e. febrile illness, etc.)

About 300 clinical and 4,000 subclinical or mild per year in Finland

Section XII - Geographic Distribution

Known (Virus detected)

Finland, Sweden, European USSR, Belgium (11)

Suspected (Antibody only detected)

Section XIII - References

1. Brummer-Korvenkontio, M., et al. 1982. Scand. J. Inf. Dis. (Suppl.) 36:88-91.
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3. World Health Organization. 1983. Bull. WHO 61:269-275.
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6. Yanagihara, R., et al. 1984. Lancet i:1013.
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8. Yanagihara, R., et al. 1985. J. Virol. 53:973-975.
9. Tkachenko, E.A., et al. 1984. Abstracts, Eleventh International Congress for Tropical Medicine and Malaria. Calgary, Canada. September 16-22.
10. van der Groen, G., et al. 1984. Abstracts, Eleventh International Congress for Tropical Medicine and Malaria. Calgary, Canada. September 16-22.
11. van der Groen, G., et al. 1987. Acta Virologica. 31:180-184.
12. Nuzum, E.O., et al. 1988. Am. J. Trop. Med. Hyg. 38:636-640.

Remarks