

<b>Virus Name: Sagiyama</b>		<b>Abbreviation: SAGV</b>
Status <b>Probable Arbovirus</b>	Select Agent <b>No</b>	SALS Level <b>3</b>
SALS Basis <b>Disease is sheep, cattle, or horses.</b>		
Other Information		
Antigenic Group <b>A</b>		

**SECTION I - Full Virus Name and Prototype Number**

Prototype Strain Number / Designation <b>M6/Mag 132</b>	Accession Number	Original Date Submitted <b>8/2/1984</b>
Family <b>Togaviridae</b>	Genus <b>Alphavirus</b>	
Information From <b>William F. Scherer</b>	Address <b>Cornell University Medical College, York Avenue, New York, N.Y. 10021</b>	
Information Footnote <b>Revised</b>		

**Section II - Original Source**

Isolated By (name) <b>Personnel 406 MGL (1,2)</b>	Isolated at Institute <b>Zama, Japan</b>	
Host Genus <b>Culex tritaeniorhynchus</b>	Species	Host Age/Stage <b>Adults</b>
Sex <b>Female</b>		
<u>Isolated From</u>	<u>Isolation Details</u>	
Signs and Symptoms of Illness	Arthropod <b>Engorged, Depleted</b>	
Time Held Alive before Inoculation <b>Less than one day</b>		
Collection Method <b>Pig-baited modified Magoon trap</b>	Collection Date <b>7/23/1956</b>	
Place Collected (Minimum of City, State, Country) <b>Sagiyama heronry, 20 miles north of Tokyo, Japan</b>		
Latitude	Longitude	
Macrohabitat <b>Heronry</b>	Microhabitat	Method of Storage until Inoculated <b>Mosquitoes frozen on dry ice on day of collection (1)</b>
Footnotes		

**Section III - Method of Isolation**

Inoculation Date  
**10/4/1956**

Animal (Details will be in Section 6)  
**nb mice**

Route Inoculated <b>ic and sc</b>	Reisolation <b>Yes</b>
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Other Reasons

Homologous Antibody Formation by Source Animal

Test(s) Used

Footnotes

**Section IV - Virus Properties**

**Physicochemical**

Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

**Stability of Infectivity (effects)**

pH (infective range)

Lipid Solvent (ether - % used to test) <b>1:5</b>	After Treatment Titer <b>&lt;3.4 dex</b>	Control Titer <b>6.4 dex</b>
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate) <b>1:1000</b>	After Treatment Titer <b>&lt;3.6 dex</b>	Control Titer <b>7.0 dex</b>
Other (formalin, radiation)		

**Virion Morphology**

Shape <b>Spherical</b>	Dimensions <b>40-50 nm</b>	
Mean nm	Range nm	
Measurement Method <b>Electron microscopy</b>	Surface Projections/Envelope	Nucleocapsid Dimensions, Symmetry

### Morphogenesis

Site of Constituent Formation in Cell	Site of Virion Assembly	Site of Virion Accumulation
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Inclusion Bodies	Other
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### Hemagglutination

Hemagglutination <b>Yes</b>	Antigen Source <b>SMB ext. by sucrose-acetone</b>	Erythrocytes (species used) <b>Goose</b>
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pH Range <b>6.1-6.6</b>	pH Optimum <b>6.2-6.4</b>
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Temperature Range <b>25dC and 37dC</b>	Temperature Optimum <b>25dC</b>
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Remarks

Serologic Methods Recommended  
**HI, CF, NT,**

Footnotes

### **Section V - Antigenic Relationship and Lack of Relationship to Other Viruses**

See References [1] , [10] , [11] for details.

Related to Group A viruses by NT and CF tests. Antiserum to Malayan viruses Getah (MM2021) and Bebaru (MM2354) neutralized each of three strains of Sagiya virus although there was only equivocal neutralization between the Malayan viruses themselves. It appears that Sagiya virus lies immunologically in between these two Malayan viruses and possesses a common antigen with one virus and another with the second virus. SAG virus should be studied further to determine whether it is a variety or a subtype of Getah virus.

The SIRACA now considers Sagiya and Bebaru viruses to be subtypes of Getah virus [11] .

**Section VI - Biologic Characteristics**

Virus Source (all VERTEBRATE isolates)

Lab Methods of Virus Recovery (ALL ISOLATIONS)  
Newborn mice

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
Procine kidney (PC)		3-5	CPE					
Hamster kidney (PC)		3-5	CPE					
BHK-21 (CL)		1	extensive	>8.0*	3	large	>8.0* (8)	
Aedes albopictus (CL)		1-2	moderate to extensive	>8.0	4	large	>8.0 (8)	

\* Expressed in dex

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Man	0/0	18/192 NT	Near Tokyo, Japan; 1956-58
Man		23/1,851 HI *	Japan; 1966
Swine	0/0	75/111 NT	Near Tokyo, Japan; 1956-57
Hérons and egrets	0/0	9/290 NT	
Chickens		18/27 HI *	Near Tokyo, Japan; 1965-66
Cow		2/13 HI *	
Goats		5/5 HI *	
Dogs		5/9 HI *	
Rabbits		3/32 HI *	
Horses	0/0	2/2 NT	Near Tokyo, Japan; 1956, 1958
<i>Culex tritaeniorhynchus</i>	8		Near Tokyo, Japan; 1956
<i>Aedes vexans</i>	2		
<i>Aedes vexans</i>	2		Near Tokyo, Japan; 1959

\* Using Getah antigen

**Section VIII - Susceptibility to Experimental Infection (include viremia)**

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log <sub>10</sub> /ml
Mice (nb)	SM 15	ic 0.01	CNS signs or paralysis	2-7	9.5-10.5
Mice (nb)		ip 0.01	CNS signs or paralysis	4-10	
Mice (nb)		sc			
Mice (wn)	SM 8	ic 0.03	None		
Mice (wn)		ip			

Titer of virus from suckling mouse muscle approximately = that from brain.

Host range: No overt disease in adult rabbits (inoculated ip) or in guinea pigs (weanlings inoculated ic and ip, and adults inoculated ip). Chicks develop viremia 2-4 days after sc and two days after iv inoculation of low mouse passage virus. Liver and spleen (pooled) and brain of baby chicks, dying 2 days after iv inoculation yielded virus. No regular deaths or signs of illness in chicks occurred after sc inoculation. In embryonated chicken eggs, certain strains produce death and hemorrhage in embryos upon inoculation by CAM or yolk sac routes. Virus has been passed by yolk sac inoculation and harvest of embryos through 15 passages. However, titers for embryonated chicken eggs remain low.

**Section IX - Experimental Arthropod Infection and Transmission**

Arthropod species & virus source(a)	Method of Infection log <sub>10</sub> /ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log <sub>10</sub> /ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System

### Section X - Histopathology

Character of lesions (specify host)

**Inflammation of central nervous system, spinal cord, brown fat, and skeletal muscle of suckling mice inoculated intracranially or intraperitoneally.**

Inclusion Bodies

Intranuclear

Organs/Tissues Affected

**Brain (LV), spinal cord (LV)**

Category of tropism

### Section XI - Human Disease

In Nature

Residual

Death

Subclinical

Overt Disease

Clinical Manifestations

Number of Cases

Category (i.e. febrile illness, etc.)

### Section XII - Geographic Distribution

Known (Virus detected)

**Japan and Okinawa (4)**

Suspected (Antibody only detected)

### Section XIII - References

1. Scherer, W.F., et al. 1962. Am. J. Trop. Med. Hyg. 11:255-268.
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6. Matsuyama, T., et al. 1968. Virus 18:11 (Haruna virus).
7. Nakamura, T., et al. 1967. Nihon Koshue. Zasshi 14:569.
8. Igarashi, A. Personal communication. 1981.
9. Igarashi, A. 1981. Am. J. Trop. Med. Hyg. 30:449-460.
10. Karabatsos, N. 1975. Am. J. Trop. Med. Hyg. 24:527-532.
11. Calisher, C.H., et al. 1980. Intervirology 14:229-232.

### Remarks

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