

Virus Name: Sawgrass		Abbreviation: SAWV
Status Possible Arbovirus	Select Agent No	SALS Level 2
SALS Basis Results of SALS surveys and information from the Catalogue.		
Other Information		
Antigenic Group Sawgrass		

SECTION I - Full Virus Name and Prototype Number

Prototype Strain Number / Designation 64A-1247	Accession Number	Original Date Submitted 12/20/1984
Family Rhabdoviridae	Genus Not listed	
Information From Florida St. Bd. of Health	Address 4001 Tampa Bay Blvd., Tampa, Florida 33614, USA	
Information Footnote Reviewed by editor		

Section II - Original Source

Isolated By (name) A.L. Lewis	Isolated at Institute Encephalitis Research Center	
Host Genus Dermacentor variabilis (pool of 19 male and 11 female)	Species	Host Age/Stage Adult
Sex Not Answered		
<u>Isolated From</u>	<u>Isolation Details</u>	
Signs and Symptoms of Illness	Arthropod	
Time Held Alive before Inoculation		
Collection Method Combed from trapped raccoon	Collection Date 5/1/1964	
Place Collected (Minimum of City, State, Country) Sawgrass Lake, Tampa Bay area, Florida, USA		
Latitude 27° 50' N	Longitude 82° 40' W	
Macrohabitat Maple swamp bordering shallow marshy lake	Microhabitat	Method of Storage until Inoculated -20dC
Footnotes		

Section III - Method of Isolation

Inoculation Date
5/13/1964

Animal (Details will be in Section 6)
nb mice

Route Inoculated
ic and ip

Reisolation
Yes

Other Reasons

Homologous Antibody Formation by Source Animal

Test(s) Used

Footnotes

Section IV - Virus Properties

Physicochemical

Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

Stability of Infectivity (effects)

pH (infective range)

Lipid Solvent (ether - % used to test)	After Treatment Titer	Control Titer
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate) 1:1000	After Treatment Titer 2.25 dex	Control Titer 4.33 dex
Other (formalin, radiation)		

Virion Morphology

Shape Rhabdovirus morphology (4)	Dimensions 220 x 65 nm	
Mean nm	Range nm	
Measurement Method By electron microscopy (4)	Surface Projections/Envelope	Nucleocapsid Dimensions, Symmetry

Morphogenesis

Site of Constituent Formation in Cell Site of Virion Assembly Site of Virion Accumulation

Inclusion Bodies Other

Hemagglutination

Hemagglutination Antigen Source Erythrocytes (species used)
No **SMB ext. by sucrose-acetone; crude pH 9.0 antigen** **Goose**

pH Range pH Optimum
5.75-7.4

Temperature Range Temperature Optimum

Remarks

Serologic Methods Recommended

Footnotes

Section V - Antigenic Relationship and Lack of Relationship to Other Viruses

Tick Viruses

N. America:	Egypt:	Africa	Europe
Hughes	Chenuda	Dugbe	Uukuniemi
Colorado tick fever	Nyamanini	Thogoto	Tribec
CalAr 846 (Farallon)	Quaranfil	Wad Medani	Johnston Atoll
Silverwater	India:	USSR:	JA
Powassan	Ganjam	Kemerovo	
Trinidad:	Bhanja	Pakistan:	
Soldado	Wanowrie	Manawa	
	Kaisodi		

Other Arboviruses	Rwamba	VSV-Pinv	Marco
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	Saint Andrew viruses	Dwamba	VOV 17,	marco
Group A -	Polyvalent	California	Chandipura	Pacui
Group B -	Polyvalent	Changuinola	Bunyamwera Supergroup	Lebombo
	Modoc	Guama	Jurona	Cotia
Group C -	Polyvalent	Koongol	Mirim	Witwatersrand
Group -	Anopheles A	Mossuril	Tacaribe	Tembe
	Tacaiuma	Nyando	Others	Tribec
	Anopheles B	Simbu (4 viruses)	Navarro	Triniti
	Bakau (Ketapang)		Hart Park	Kowanyama
	Bunyamwera (6 viruses)	Turlock	Kern Canyon	Klamath

Serological tests were performed by Miss Gladys Sather in YARU laboratories.

Sawgrass antigen was non-reactive with immune sera to the above viruses by CF. Related by CF to registered New Minto virus. Both viruses were shown to be related to the subsequently registered Connecticut virus. These viruses now constitute the Sawgrass serogroup. See New Minto and Connecticut virus registrations.

Section VI - Biologic Characteristics

Virus Source (all VERTEBRATE isolates)

Lab Methods of Virus Recovery (ALL ISOLATIONS)
Newborn mice

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
Vero (CL)	SM 7				6	3 mm	4.6*	
LLC-MK2 (CL)					11	2 mm	5.2	
Vero (CL)		3-5	CPE (5)					
CER (CL)		2	CPE (2)					
BHK-21 (CL)		2	CPE (5)					
Duck embryo (PC)			No CPE (5)					

* Expressed in dex

Section VII - Natural Host Range (Additional text can be added below table)

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Dermacentor variabilis	16		Tampa Bay area, Florida, USA (3)
Haemaphysalis leporispalustris	12		

Section VIII - Susceptibility to Experimental Infection (include viremia)

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log ₁₀ /ml
Mice (nb)	SM 4	ic 0.01	Illness and death	3-4	5.2
Mice (nb)		ip			
Mice (nb)		sc			
Mice (wn)		ic 0.03	None		
Mice (wn)		ip			

Section IX - Experimental Arthropod Infection and Transmission

Arthropod species & virus source(a)	Method of Infection log ₁₀ /ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log ₁₀ /ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System

Section X - Histopathology

Character of lesions (specify host)

Inclusion Bodies

Intranuclear

Organs/Tissues Affected

Category of tropism

Section XI - Human Disease

In Nature	Residual	Death
Subclinical	Overt Disease	
Clinical Manifestations		
Number of Cases	Category (i.e. febrile illness, etc.)	

Section XII - Geographic Distribution

Known (Virus detected) Florida, USA
Suspected (Antibody only detected)

Section XIII - References

1. Sather, G.E., et al. 1970. Am. J. Trop. Med. Hyg. 19:319-326. 2. Stim, T.B. 1969. J. Gen. Virol. 5:329-338. 3. Wellings, F.M., et al. 1972. Am. J. Trop. Med. Hyg. 21:201-213. 4. Murphy, F.A. Personal communication. 1977. 5. Calisher, C.H. Personal communication. 1981.

Remarks

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