

Virus Name: Simian hemorrhagic fever		Abbreviation: SHFV
Status Not Arbovirus	Select Agent No	SALS Level 2
SALS Basis Results of SALS surveys and information from the Catalogue.		
Other Information		
Antigenic Group Ungrouped		

SECTION I - Full Virus Name and Prototype Number

Prototype Strain Number / Designation VR42-0/M6941 (1,3)	Accession Number	Original Date Submitted 11/17/1984
Family Flaviviridae	Genus Not listed	
Information From Dr. Nicola M. Tauraso	Address Biologics Standards, National Institutes of Health, Bethesda, MD 20014 USA	
Information Footnote Reviewed by editor		

Section II - Original Source

Isolated By (name) Dr. Nicola M. Tauraso	Isolated at Institute Division of Biologics Standards, NIH	
Host Genus Rhesus (<i>Macaca mulatta</i>) monkey, no sentinel	Species	Host Age/Stage Juvenile
Sex Not Answered		
<u>Isolated From</u>	<u>Isolation Details</u>	
Serum/Plasma		
Signs and Symptoms of Illness Rapid onset, early fever, mild facial edema, anorexia, adipsia, dehydration, [proteinuria, cyanosis, skin [petechiae, melena, epistaxis [and shock.	Arthropod	
Time Held Alive before Inoculation		
Collection Method Terminal bleeding	Collection Date 1/29/1966	
Place Collected (Minimum of City, State, Country) Bethesda, Maryland		
Latitude	Longitude	
Macrohabitat	Microhabitat Indoors	Method of Storage until Inoculated Serum frozen at -70dC
Footnotes		

Section III - Method of Isolation

Inoculation Date
5/20/1966

Animal (Details will be in Section 6)
(Tissue Culture)

Route Inoculated Reisolation
Yes

Other Reasons
Other surviving animals developed CF, IFA and neut antibodies

Homologous Antibody Formation by Source Animal
Not tested

Test(s) Used
original animal died

Footnotes

Section IV - Virus Properties

Physicochemical
RNA

Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

Stability of Infectivity (effects)

pH (infective range)
3.0 labile; rel. heat (50C) stable; divalent cations enhanced inact. at 50C

Lipid Solvent (ether - % used to test)	After Treatment Titer	Control Titer
Lipid Solvent (chloroform) 1:20	After Treatment Titer <1.0 dex	Control Titer 4.8 dex
Lipid Solvent (deoxycholate)	After Treatment Titer	Control Titer
Other (formalin, radiation)		

Virion Morphology

Shape Spherical	Dimensions 40-45 nm	
Mean nm	Range nm	
Measurement Method Electron microscopy (4)	Surface Projections/Envelope Envelope observed (4);	Nucleocapsid Dimensions, Symmetry

Morphogenesis

Site of Constituent Formation in Cell	Site of Virion Assembly	Site of Virion Accumulation
Inclusion Bodies	Other	

Hemagglutination

Hemagglutination No	Antigen Source Infected cell cultures; extracted infected spleen and brain (5)	Erythrocytes (species used) Goose
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pH Range *6.8-7.4	pH Optimum
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Temperature Range 4dC, 24dC, and 37dC	Temperature Optimum
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Remarks

* Other erythrocytes tested included rhesus and African green monkeys, hamster, mouse, rat, guinea pig, human (type O) and day-old chick.

Serologic Methods Recommended
CF, IFA and NT

Footnotes

* Other erythrocytes tested included rhesus and African green monkeys, hamster, mouse, rat, guinea pig, human (type O) and day-old chick.

No relationship found to other viruses by CF and IFA tests (see Ref. [6] , [7]).

In one-way tests using 20X concentrated cell culture SHF antigen for CF tests or SHF virus-infected coverslips for fluorescent-antibody (FA) tests, with antisera against the following viruses, negative results were obtained [6] .

Adenoviruses:	SV-11, SV-15, SV-17, SV-20, SV-25, SV-27, SV-36, ICH.	
Arboviruses:	Group A:	Bebaru, EEE, WEE, Una, Uruma (Mayaro)
	Group B:	Bussuquara, dengue 1, 2, 3, 4, Ilheus, MVE, Powassan, SLE, WN
	Bunyamwera group:	Batai, bunyamwera, Cache Valley, Germiston, Guaroa, Ilesha, Kairi, Sororoca, Tensaw, Calovo, Wyeomyia
	California group:	CE, Melao, Tahyna
	Miscellaneous:	Anopheles A, Changuinola, Chagres, EHD, Icoaraci, Kemerovo, Naples SF, Tacaribe, Junin, Tribec, VS-Indiana, VS-NJ, Cocal
Herpesviruses:	H. simplex, canine herpes, simian cytomegalic, B virus	
Myxoviruses:	Parainfluenza 1, 2, 3, SV-5; avian leukosis (RSV/RIF and RSV-Harris); SV-41, measles, mumps, respiratory syncytial, rubella, NDV, rabies	
Papovavirus:	SV-40, K virus	
Picornavirus:	ECHO 1, 2, 3, 5, 6, 7, 8, 9, 11, 12, 14; SV-2, SV-16, SV-19, Theiler's GD VII; poliovirus 1, 2, 3	
Reovirus:	Reo 1, 2, 3, SV-12, SV-59	
Miscellaneous:	SV-4, SV-28, Sv-31, SA-1; PVM, LCM, Crimmean HF, Korean HF, Himalaya HF; hamster HF, human gamma globulin (1:10)	

Positive results were obtained only with sera from simian hemorrhagic fever outbreaks: Sukhumi/64, Bethesda/64, Davis/67, and Sussex/68.

SHF not related to Marburg virus [7] .

Section VI - Biologic Characteristics

Virus Source (all VERTEBRATE isolates)
Blood (LV)

Lab Methods of Virus Recovery (ALL ISOLATIONS)
USU-104, embryonic rhesus kidney cell culture.

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
USU-104 (CL)	M/2, MA-104/8	6-7	CPE	5.0* (3)				

* Expressed in dex

Section VII - Natural Host Range (Additional text can be added below table)

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Erythrocebus patas (monocytes) (Chronically infected, continuously viremic)	1 *		NIH, Bethesda, MD, USA (13)

Rhesus (*Macaca mulatta*) monkeys Virus isolated only from clinically affected animals recently imported from New Delhi, India (1). Other species listed below have developed disease after contact with recently imported sick rhesus monkeys. Many virus isolations made of the Bethesda/64 agent (8).

Other monkey species affected: *M. nemestrina* (9,10); *M. speciosa* (1-3); *M. assamensis* (9,10); *M. irus* (1-3,11) with a disease similar to that seen in *M. mulatta*.

* Isolate differs antigenically and biologically from prototype SHF virus (13)

Section VIII - Susceptibility to Experimental Infection (include viremia)

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log ₁₀ /ml
Mice (nb)		ic	None		
Mice (nb)		ip	None		
Mice (nb)		sc			
Mice (wn)		ic	None		
Mice (wn)		ip	None		
rhesus (<i>M. mulatta</i>) (juvenile)	M/2, MA-104/8	sc 1	Anorexia, hemorrhagic diatheses, eventually shock and death	5-14	7-8

Unable to infect the following hosts: Suckling and young adult Syrian hamster, Hartley guinea pig, white mouse, Sprague-Dawley rat, and cat, and embryonated chicken egg.

Section IX - Experimental Arthropod Infection and Transmission

Arthropod species & virus source(a)	Method of Infection log ₁₀ /ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log ₁₀ /ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System

Section X - Histopathology

Character of lesions (specify host)

Lesions of SHF in rhesus monkey include capillary and venous hemorrhages in the intestine, lung, nasal mucosa, dermis, spleen, perirenal and lumbar subperitoneum, adrenal and liver. Two striking lesions occur in the duodenum and spleen (2).

Inclusion Bodies

Intranuclear

Organs/Tissues Affected

Lungs (LV), liver (LV), spleen (LV), blood vessels (LV)

Category of tropism

Section XI - Human Disease

In Nature

Residual

Death

Subclinical

Overt Disease

Clinical Manifestations

Human disease not observed

Number of Cases

Category (i.e. febrile illness, etc.)

Section XII - Geographic Distribution

Known (Virus detected)

From recently imported rhesus (*Macaca mulatta*) monkeys

Suspected (Antibody only detected)

Section XIII - References

1. Palmer, A.E., et al. 1968. Am. J. Trop. Med. and Hyg. 17:404-412.
2. Allen, A.M., et al. 1968. Am. J. Trop. Med. and Hyg. 17:413-421.
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4. Wood, O.L., et al. 1970. J. Gen. Virol. 7:129-136.
5. Clarke, D.H. and Casals, J. 1958. Am. J. Trop. Med. and Hyg. 7:561-573.
6. Tauraso, N.M., et al. In "Marburg Virus Disease", Springer-Verlag, Berlin-West. 1971. pp. 208-215.
7. Casals, J. 1971. In "Marburg Virus Disease". Springer-Verlag, Berlin-West. 1971. pp. 98-104.
8. Tauraso, N.M., et al. In "Infections and Immunosuppression in Sub-human Primates". H. Balner and W.I.B. Beveridge (Ed.) Munksgaard, Copenhagen, Denmark. 1970. pp. 101-109.
9. Shevtsova, Z.V. 1967. Vop. Virusol, 12:47-51.
10. Lapin, B.A., et al. 1967. Vop Virusol. 12:168-173.
11. Shelokov, A., et al. In "Marburg Virus Disease". Springer-Verlag, Berlin-West. 1971. pp. 203-207.
12. Tauraso, N.M., et al. 1968. Nature 218:876-877.
13. Gravell, M., et al. 1980. J. Gen. Virol. 51:99-106.

Remarks

At present there is no evidence to suggest that arthropods play a role in the transmission of SHF (6).