

<b>Virus Name: Sin Nombre</b>		<b>Abbreviation: SNV</b>
Status <b>Probably not Arbovirus</b>	Select Agent <b>No</b>	SALS Level
SALS Basis		
Other Information <b>DOC Permit Required</b>		
Antigenic Group <b>hantaan</b>		

**SECTION I - Full Virus Name and Prototype Number**

Prototype Strain Number / Designation	Accession Number	Original Date Submitted 11/19/1993
Family <b>hantavirus</b>	Genus	
Information From <b>Peters, Ksiazek, Nichol, Childs, Rollin</b>	Address <b>Special Pathogens Branch, DVRD, NCID, Centers for Disease Control, Atlanta, GA 30333</b>	
Information Footnote		

**Section II - Original Source**

Isolated By (name) <b>Elliot et al, (in preparation)</b>	Isolated at Institute <b>Atlanta, Georgia</b>	
Host Genus <b>Peromyscus maniculatus</b>	Species	Host Age/Stage <b>adult</b>
Sex <b>Not Answered</b>		
<u>Isolated From</u>	<u>Isolation Details</u>	
<b>Organs/Tissues</b>	<b>lungs</b>	
Signs and Symptoms of Illness <b>none (PCR positive in tissues and antibody negative when sacrificed)</b>	Arthropod	
Time Held Alive before Inoculation		
Collection Method <b>trapped</b>	Collection Date <b>6/1/1993</b>	
Place Collected (Minimum of City, State, Country) <b>Ramah, New Mexico</b>		
Latitude <b>36° 10' N</b>	Longitude <b>108° 28' W</b>	
Macrohabitat	Microhabitat	Method of Storage until Inoculated <b>-70dC</b>
Footnotes		

**Section III - Method of Isolation**

Inoculation Date <b>1/1/1993</b>	
Animal (Details will be in Section 6) <b>yes</b>	
Route Inoculated <b>intraperitoneal</b>	Reisolation <b>Not tried</b>
Other Reasons	
Homologous Antibody Formation by <u>Source Animal</u> <b>Yes</b>	
Test(s) Used <b>IFA, ELISA</b>	
Footnotes	

**Section IV - Virus Properties**

Physicochemical <b>RNA, Single Strand</b>		
Pieces (number of genome segments) <b>3</b>	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number <b>4</b>	Details <b>nucleoprotein (N) 50,000MW; glycoproteins (G1, G2); 55,000MW, 72,000MW; (L); 200,000MW</b>	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	
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<b><u>Stability of Infectivity (effects)</u></b>		
pH (infective range)		
Lipid Solvent (ether - % used to test)	After Treatment Titer	Control Titer
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate)	After Treatment Titer	Control Titer
Other (formalin, radiation)		
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<b><u>Virion Morphology</u></b>		
Shape <b>bunyavirus-like</b>	Dimensions <b>80-115 nm</b>	
Mean nm	Range nm	
Measurement Method <b>by electron microscopy</b>	Surface Projections/Envelope <b>surface projections, lipid envelope</b>	Nucleocapsid Dimensions, Symmetry

**Morphogenesis**

Site of Constituent Formation in Cell	Site of Virion Assembly	Site of Virion Accumulation
Inclusion Bodies	Other	

**Hemagglutination**

Hemagglutination <b>Not tried</b>	Antigen Source	Erythrocytes (species used)
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pH Range	pH Optimum
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Temperature Range	Temperature Optimum
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Remarks

Serologic Methods Recommended  
**IFA, ELISA \*2: (a) Handling serum from potentia**

Footnotes

**Section V - Antigenic Relationship and Lack of Relationship to Other Viruses**

Antigenically related to, but distinct from the other registered hantavirus, Prospect Hill, Hantaan, Puumala, Seoul. Data resulting from the serological comparison of Muerto Canyon virus to other registered hantaviruses, Prospect Hill, Hantaan, Puumala and Seoul is not available. However, nucleotide sequence characterization of the three RNA segments composing the viral genome showed that each segment was unique and that there was approximately 30% nucleotide sequence divergence from the respective segments of the closest relative, Prospect Hill virus [14]. Up to 10% nucleotide sequence variation was seen between hantavirus pulmonary syndrome (HPS) viruses from New Mexico, Arizona and Colorado, both from rodents and from human autopsy tissues. Even greater variation (up to 14%) was observed when the analysis included viruses from other regions [14].

**Section VI - Biologic Characteristics**

Virus Source (all VERTEBRATE isolates)  
tissues of vertebrates (mainly lungs, spleen, kidney)

Lab Methods of Virus Recovery (ALL ISOLATIONS)  
Peromyscus maniculatus, Vero E-6 cells

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Peromyscus maniculatus		30%/813 * ELISA	Four Corners area, USA June-August, 1993 (15)
Peromyscus truei		19.6%/275	
Peromyscus boylii		5.9%/51	
Tamias dorsalis		3.6%/166	
Tamias minutus		5.0%/19	
Tamias quadrivittatus		13.0%/30	
Mus musculus		3.9%/51	
Neotoma albigula		2.9%/69	
Reithrodontomys megalotis		28.6%	
Spermophilus variegatus		1.0%/90	
Sylvilagus auduboni (nonrodent species)		12.5%/8	

\* Percent of rodents positive for ELISA antibody using PH antigen/total number of specimens tested. Almost all specimens also tested with SEO and PUU antigens. With two exceptions, PH antigen gave a higher percent positive reactions than the other two antigens. The following rodent species (number tested vs PH antigen) were negative for ELISA antibody for either PH and SEO or, PH, SEO and PUU antigens. Peromyscus nasutus (10), Peromyscus crinitus (1), Ammospermophilus leucurus (14), Cynomys gunnisoni (1), Dipodomys merriami (4), Dipodomys ordii (6), Dipodomys species (1), Microtus longicaudus (2), Microtus mexicanus (1), Microtus montanus (2), Microtus pennsylvanicus (4), Neotoma mexicana (20), Neotoma micropus (4), Neotoma stephansi (6), Neotoma species (2), Onychomys leucogaster (10), Onychomys species (2), Perognathus flavus (5), Perognathus species (2), Sigmodon hispidus (6), Spermophilus spilosoma (4), Spermophilus tridecemlineatus (7), Spermophilus lateralis (2).



Section XI - Human Disease

In Nature <b>Significant</b>	Residual	Death <b>Significant</b>
Subclinical	Overt Disease	
Clinical Manifestations <b>acute respiratory distress after a few days of non-specific prodromal syndrome (fever, myalgia, malaise)</b>		
Number of Cases	Category (i.e. febrile illness, etc.) <b>respiratory distress</b>	

Section XII - Geographic Distribution

Known (Virus detected) <b>states west of the Mississippi River, Arizona, California, Colorado, North Dakota, South Dakota, Louisiana, Montana, Nevada, New Mexico, Oregon, Texas</b>
Suspected (Antibody only detected)

Section XIII - References

<ol style="list-style-type: none"><li>Centers for Disease Control. 1993. M.M.W.R. 42:421-424.</li><li>Centers for Disease Control. 1993. M.M.W.R. 42:441-443.</li><li>Centers for Disease Control. 1993. M.M.W.R. 42:477-479.</li><li>Centers for Disease Control. 1993. M.M.W.R. 42:495-496.</li><li>Centers for Disease Control. 1993. M.M.W.R. 42:517-519.</li><li>Centers for Disease Control. 1993. M.M.W.R. 42:570-572.</li><li>Centers for Disease Control. 1993. M.M.W.R. 42:707.</li><li>Centers for Disease Control. 1993. M.M.W.R. 42:770.</li><li>Centers for Disease Control. 1993. M.M.W.R. 42:816-820.</li><li>Feldmann, H., Sanchez, A., Morzunov, S., et al. 1993. Virus. Res. In press.</li><li>Hughes, J.M., Peters, C., Cohen, M., et al. 1993. Science. 262:832-836.</li><li>Nichol, S., Spiropoulou, C., Morzunov, S., et al. 1993. Science. 262:914-917.</li><li>Marshall, E., Stone, R. 1993. Science. 262:832-836.</li><li>Spiropoulou, C., Morzunov, S., Feldmann, H., et al. 1994. Virology. 200:715-723.</li><li>Childs, J.E., Ksiazek, T.G., Spiropoulou, C., et al. 1994. J. Inf. Dis. 169:1271-1280.</li></ol>
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Remarks

<b>Sin Nombre virus is the apparent etiologic agent of Hantavirus Pulmonary Syndrome (11).</b>
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