

Virus Name: Sindbis		Abbreviation: SINV
Status Arbovirus	Select Agent No	SALS Level 2
SALS Basis Results of SALS surveys and information from the Catalogue.		
Other Information		
Antigenic Group A		

SECTION I - Full Virus Name and Prototype Number

Prototype Strain Number / Designation Ar 339	Accession Number	Original Date Submitted 12/9/1984
Family Togaviridae	Genus Alphavirus	
Information From R.M. Taylor	Address School of Public Health, University of California, Berkeley, California 94720	
Information Footnote Reviewed by editor		

Section II - Original Source

Isolated By (name) R.M. Taylor, et al. (1,2)	Isolated at Institute NAMRU-3, Cairo, Egypt	
Host Genus Culex univittatus	Species	Host Age/Stage Adult (imago)
Sex Female		
<u>Isolated From</u>	<u>Isolation Details</u>	
Signs and Symptoms of Illness	Arthropod	
Time Held Alive before Inoculation		
Collection Method Light trap	Collection Date 8/14/1952	
Place Collected (Minimum of City, State, Country) Sindbis Village, Nile Delta, Egypt		
Latitude 30° N	Longitude 32° E	
Macrohabitat Irrigated, highly cultivated, delta plain, subtropical climate	Microhabitat Light trap set in village street during night	Method of Storage until Inoculated None; inoculated same day as captured
Footnotes		

Section III - Method of Isolation

Inoculation Date
8/14/1952

Animal (Details will be in Section 6)
nb mice

Route Inoculated
ic and sc

Reisolation
Yes

Other Reasons

Original source; subsequently 12 other isolations and presence of antibodies in vertebrates of region

Homologous Antibody Formation by Source Animal

Test(s) Used

Footnotes

Section IV - Virus Properties

Physicochemical
RNA, Single Strand

Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

Stability of Infectivity (effects)

pH (infective range)

Lipid Solvent (ether - % used to test) 20%	After Treatment Titer <2 dex	Control Titer 4.5 to 6 dex (4)
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate) 1:1000	After Treatment Titer <2 dex	Control Titer 4.5 to 6 dex (4)
Other (formalin, radiation)		

Virion Morphology

Shape	Dimensions 32 nm	
Mean nm	Range nm	
Measurement Method Electron microscopy (3)	Surface Projections/Envelope Envelope present (27)	Nucleocapsid Dimensions, Symmetry Capsid (27)

Morphogenesis

Site of Constituent Formation in Cell	Site of Virion Assembly	Site of Virion Accumulation
Inclusion Bodies	Other	

Hemagglutination

Hemagglutination Yes	Antigen Source SMB ext. by acetone-ether	Erythrocytes (species used) Goose
pH Range	pH Optimum	
Temperature Range	Temperature Optimum	

Remarks

Originally reported as resistant to both ether and sodium deoxycholate, but subsequently found to be sensitive though not to the degree of some other arboviruses (5). Sindbis and SF viruses are both RNA viruses and have been used extensi

Serologic Methods Recommended

HI, CF, NT

Footnotes

Originally reported as resistant to both ether and sodium deoxycholate, but subsequently found to be sensitive though not to the degree of some other arboviruses (5). Sindbis and SF viruses are both RNA viruses and have been used extensi

Section V - Antigenic Relationship and Lack of Relationship to Other Viruses

In the initial studies by Casals [6], Sindbis was found by the HI method to fall into serogroup A and to be most closely related to WEE. Since then a number of strains of viruses have been isolated in widely separated parts of the world that are either indistinguishable or very closely related to Sindbis virus and are regarded as strains of this virus [7]. These strains are SAAr-86 isolated in South Africa [8], IA-1036 isolated in India [9], MM-2215 isolated in Malaya [10], P-886 isolated in the Philippines [11], MRM-39 isolated in Australia [12] and MP-684 isolated in Uganda [13].

As far as is known Sindbis is not related to any viruses not in Group A.

References on antigenic and serologic relation to other members of Group A are too numerous to include.

Section VI - Biologic Characteristics

Virus Source (all VERTEBRATE isolates)
 Blood (M)(LV), heart (LV), spleen (LV) (9), skin lesions (M)
 (14)

Lab Methods of Virus Recovery (ALL ISOLATIONS)
 Newborn mice; primary monkey kidney cell cultures (9)

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
Vero (CL)	SM 9				3	10 mm	7.7* (17)	
LLC-MK2 (CL)					2	1 mm	8.0 (17)	
BHK-21 (CL)	Amm 2215 SM 2	1	4+	>8.5* (40)				

Grows readily and forms plaques on chick embryo, primary monkey kidney, human amniotic cells, and other mammalian cell cultures (16). Both primary and continuous cell cultures have been used. Numerous other references are available.

* Expressed in dex

Section VII - Natural Host Range (Additional text can be added below table)

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Man	5	74/938 HI	Uganda (13); SE Anatolia, Turkey (37)
Man	1	122/445 NT	South Africa (14); Nile Delta, Egypt (2)
Man	1	16/132 NT	Cent. Af. Rep. (34); Sudan (2)
Mammals		52/168 NT	Nile Delta, Egypt (2)
Avian species		9/125 NT	
Corvus corone sardonius	1/159		
Birds, wild	1		South Africa (15)
Motacilla alba	1		India (9)

Gracula religiosa	1	
Sentinel mice	4	Ibadan, Nigeria (20)
Culex pseudovishnui	16	Sarawak, Malaysia (21)
Culex univittatus	9	Nile Delta, Egypt (2); South Africa (14)
Cx atennatus	4	Nile Delta, Egypt (2)
Cx tritaeniorhynchus	1	Malaya, Malaysia (10)
Cx bitaeniorhynchus	1	Philippines (11)
Cx annulirostris	5	Australia (12)
Culex mixed pools	12	S. Africa (8, 14); India (9)
Aedes normanensis	1	Australia (12)
Ae vigilax	1	Australia (12)
Anopheles pharoensis	1	Nile Delta, Egypt (2)
Mansonia semptempunctata	1	Australia (12)
Ma fuscopennata	1	Uganda (13)
Bdellonyssus bursa	1	India (9)
Culex quinquefasciatus	1	Queensland, Australia (39)
Hyalomma marginatum Hipposideridae sp. and Rhinolophidae sp.	1	Sicily (35)
(insectivorous bats; organs)	1	Zimbabwe (38)
Culex univittatus	13	Al Khobar, Saudi Arabia (41)
Cx. tritaeniorhynchus	1	
Cx. pipiens complex	1	
Culex spp.	1	
Aedes mosquitoes (89% Ae. communis)	1	Karelian area, USSR (42)

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log ₁₀ /ml
Mice (nb)	SMB 5	ic 0.02	Paralysis and death	2-4	7.5
Mice (nb)		ip 0.02	Paralysis and death	2-4	6.9
Mice (nb)		sc			
Mice (wn)		ic 0.03	No deaths*		
Mice (wn)		ip			
emb. eggs(8 day)	SMB 3	ys 0.1	Death	1-3	6.0

Cercopithecus aethiops and *Macaca mulatta*, young chickens and cattle egret (*Bubulcus ibis ibis*) can be infected by bite of mosquito, circulate virus and develop NT antibodies. Crows (hooded) and doves susceptible to sc inoculation. One trial to infect sheep by mosquitoes failed. Adult hamsters and rabbits survive ic inoculation.

Chickens and pigeons circulate virus and death may result. Rabbits, rats, and marsupials are resistant (22). 42 of 53 S. African wild birds of 14 species circulated virus after im inoculation and developed antibodies (23).

* Strain isolated in South Africa was adapted to adult mice (8).

Section IX - Experimental Arthropod Infection and Transmission

Arthropod species & virus source(a)	Method of Infection log10/ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log10/ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System

Culex univittatus readily become infected and transmit by bite. *Ornithodoros savignyi* ticks infected by parenteral inoculation will infect by bite but complete cyclic transmission by this tick and *Argas persicus* was not achieved. *Ae aegypti* and *Anopheles albimanus* will also transmit by bite (24), and virus is expelled from anus by infected *Ae aegypti* (25).

Sindbis appears to be essentially a bird virus transmitted mainly by *Culex* mosquitoes, but man and other mammals may become tangentially infected.

--	--	--	--	--	--	--	--	--	--

Section X - Histopathology

Character of lesions (specify host)

sm; ic, ip: CNS and skeletal muscles principally involved (2). Necrosis in periarticular tissue, marrow, thymus, heart muscle, connective tissue and brown fat (14). Eosinophilic intranuclear inclusions observed in tissue culture (14)

Inclusion Bodies

Intranuclear

Organs/Tissues Affected

Brain (LV), heart (LV), blood vessels (LV), marrow (LV), skeletal muscles (LV), secretory glands (LV)

Category of tropism

Section XI - Human Disease

In Nature
Reported

Residual
Reported

Death

Subclinical

Overt Disease

Clinical Manifestations

Fever(R), headache (S), prostration(R), stiff neck(R), myalgia(S), arthralgia (R), hemorrhagic signs (R)(36), rash (S), jaundice (R)

Number of Cases

>30; Uganda; South Africa (13,14,15,18); Australia (26)

Category (i.e. febrile illness, etc.)

Febrile illness with rash

Section XII - Geographic Distribution

Known (Virus detected)

Egypt (2), Uganda (13), South Africa (8,14,15), Cameroun (28), India (9), Philippines (11), Malaysia (10,21,29), Australia (12), Cent. Afr. Rep. (28), Israel (30), Mozambique (31), USSR (32), Czechoslovakia (33), Italy (35), Zimbabwe (38), Nigeria (

Suspected (Antibody only detected)

1. TAYLOR, R.M. 1953. *Atti del VI Congr. Internaz. di Microbiol.* 3:236.
2. TAYLOR, R.M., et al. 1955. *Am. J. Trop. Med. Hyg.* 4:844.
3. HORZINEK, M. and MUSSGAY, M. 1969. *J. Virol.* 4:514.
4. PFEFFERKORN, E.R., et al. 1967. *Viol.* 33:239.
5. SUNAGA, H., et al. 1960. *Am. J. Trop. Med. Hyg.* 9:419.
6. CASALS, J. 1957. *Trans. New York Acad. Sciences* 19:219.
7. CASALS, J. Personal communication. 1960.
8. WEINBREN, M.P., et al. 1956. *S. Afr. Med. J.* 30:631.
9. SHAH, K.V., et al. 1960. *Indian Med. Res.* 48:300.
10. BUESCHER, E.L. Personal communication.
11. RUDNICK, A., et al. 1962. *Am. J. Trop. Med. Hyg.* 11:546.
12. DOHERTY, R.L., et al. 1963. *Aust. J. Exp. Biol. Med. Sci.* 41:17.
13. *E. Afr. Virus Res. Inst. Rep.* 1962. No. 12.
14. MALHERBE, H., et al. 1963. *S. Afr. Med. J.* 37:547.
15. McINTOSH, B.M. Personal communication.
16. FROTHINGHAM, T.E. 1955. *Am. J. Trop. Med. Hyg.* 4:863.
17. STIM, T.B. 1969. *J. Gen. Virol.* 5:329.
18. McINTOSH, B.M., et al. 1964. *S. Afr. Med. J.* 38:291.
19. BEN-PORATH, E., et al. 1965. *Israel J. Med. Sci.* 1:88.
20. MOORE, D.L. Personal communication. 1971.
21. SIMPSON, D.H., et al. 1970. *Ann. Trop. Med. and Parasitol.* 64:137.
22. WHITEHEAD, R.H. 1969. *Aust. J. Exp. Biol. Med. Sci.* 47:11.
23. McINTOSH, et al. 1969. *S. Afr. J. Med. Sci.* 34:77, 83.
24. COLLINS, W.E., et al. 1966. *Mosq. News* 26:91.
25. MUUNGMAN, D., et al. 1969. *Am. J. Trop. Med. Hyg.* 18:401.
26. DOHERTY, R.L., et al. 1969. *Med. J. Australia* 2:1016.
27. BROWN, D.T., et al. 1972. *J. Virol.* 10:524.
28. *Rapport Annuel de l'Institut Pasteur de Dakar.* 1972.
29. LIM, T.W., et al. 1972. *Med. J. Malaysia* 27:147.
30. NIR, Y., et al. 1972. *Israel J. Med. Sci.* 8:1695.
31. Director, S. Afr. Inst. Med. Res. Personal communication. 1973.
32. GROMASHEVSKY, K.B., et al. 1973. *Acta Virol.* 17:155.
33. ERNEK, E., et al. 1973/1974. *Intervirology.* 2:186.
34. *Rapport Annuel de l'Institut Pasteur de Bangui.* 1974.
35. GRESIKOVA, M., et al. 1978. *Acta Virol.* 22:231.
36. GUARD, R.W., et al. 1982. *Pathology* 14:89.
37. MECO, O. 1981. *Mikrobiyoloji Bulteni* 15:1.
38. BLACKBURN, N.K., et al. 1982. *Cent. Afr. J. Med.* 28:201.
39. DOHERTY, R.L., et al. 1973. *Trans. R. Soc. Trop. Med. Hyg.* 67:536.
40. KARABATSOS, N. and BUCKLEY, S.M. 1967. *Am. J. Trop. Med. Hyg.* 16:99.
41. WILLS, W.M. et al. 1984. *Trns. Roy. Soc. Trop. Med. Hyg.* 79:63-66".

Remarks