

<b>Virus Name: Slovakia</b>		<b>Abbreviation: SLOV</b>
Status <b>Not Arbovirus</b>	Select Agent <b>No</b>	SALS Level <b>3</b>
SALS Basis <b>Insufficient experience with virus; i.e., experience factor from SALS surveys was less than 500 in laboratory facilities with low biocontainment.</b>		
Other Information		
Antigenic Group <b>Ungrouped</b>		

**SECTION I - Full Virus Name and Prototype Number**

Prototype Strain Number / Designation <b>Argas persicus 265</b>	Accession Number	Original Date Submitted <b>5/31/1984</b>
Family <b>Not listed</b>	Genus <b>Not listed</b>	
Information From <b>M. Gresikova, et al.</b>	Address <b>Institute of Virology, Slovak Academy of Sciences, Bratislava, Czechoslovakia</b>	
Information Footnote		

**Section II - Original Source**

Isolated By (name) <b>M. Gresikova, et al. (1)</b>	Isolated at Institute <b>Institute of Virology, Bratislava</b>	
Host Genus <b>Argas persicus ticks (not fully engorged)</b>	Species	Host Age/Stage <b>Imago</b>
Sex <b>Female</b>		
<u>Isolated From</u>	<u>Isolation Details</u>	
Signs and Symptoms of Illness	Arthropod <b>Engorged</b>	
Time Held Alive before Inoculation <b>55 days</b>		
Collection Method <b>Manual collection from chickens</b>	Collection Date <b>7/7/1976</b>	
Place Collected (Minimum of City, State, Country) <b>S. Slovakia, Bratislava Co., Vajnory area, CZ</b>		
Latitude <b>48° 8' N</b>	Longitude <b>17° 8' E</b>	
Macrohabitat <b>Chicken house</b>	Microhabitat <b>Cracks in wood</b>	Method of Storage until Inoculated <b>In glass tube at +4dC</b>
Footnotes		

**Section III - Method of Isolation**

Inoculation Date  
**9/1/1976**

Animal (Details will be in Section 6)  
**nb mice**

Route Inoculated <b>Intracerebral</b>	Reisolation <b>Yes</b>
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Other Reasons  
**Agent different from any virus in laboratory**

Homologous Antibody Formation by Source Animal

Test(s) Used

Footnotes

**Section IV - Virus Properties**

**Physicochemical**

Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

**Stability of Infectivity (effects)**

pH (infective range)  
**Persists at pH 3.0**

Lipid Solvent (ether - % used to test) <b>1:1</b>	After Treatment Titer <b>3.0 dex</b>	Control Titer <b>4.5 dex</b>
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate) <b>1:1000</b>	After Treatment Titer <b>4.5 dex</b>	Control Titer <b>4.5 dex</b>

Other (formalin, radiation)  
**Virus protected against thermal inactivation by 1M MgCl2**

**Virion Morphology**

Shape	Dimensions <b>&lt;50 nm</b>	
Mean nm	Range nm	
Measurement Method <b>Millipore filtration</b>	Surface Projections/Envelope	Nucleocapsid Dimensions, Symmetry

### Morphogenesis

Site of Constituent Formation in Cell	Site of Virion Assembly	Site of Virion Accumulation
Inclusion Bodies	Other	

### Hemagglutination

Hemagglutination <b>No</b>	Antigen Source <b>SMB ext. by sucrose-acetone</b>	Erythrocytes (species used) <b>Goose*</b>
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pH Range <b>5.7-7.0</b>	pH Optimum
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Temperature Range	Temperature Optimum
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#### Remarks

\* **Human 0 erythrocytes also tested with negative results.**

#### Serologic Methods Recommended

CF

#### Footnotes

\* **Human 0 erythrocytes also tested with negative results.**

### **Section V - Antigenic Relationship and Lack of Relationship to Other Viruses**

#### Complement-fixation:

The Slovakia antigen (homologous titer of 64/32) gave negative results by CF when tested against the following immune grouping fluids obtained from NIH: Groups A, B, C, Guama, Simbu, VSV, Bunyamwera, California, Tacaribe, Phlebotomus fever, Polyvalent Quarantil, Polyvalent Anopheles A, Anopheles B and Turlock, Polyvalent Patois, Group Capim, Polyvalent Palyam, Group Kemerovo, Polyvalent Congo, Polyvalent rabies, and Polyvalents 1 through 10 and Polyvalent 12.

In other CF tests, Slovakia antigen failed to fix complement in the presence of hyperimmune sera prepared against tick-borne encephalitis, Kemerovo, Uukuniemi, Tettang, and Eyach viruses.

Slovakia virus antibody cross-reacts with an antigen of mouse hepatitis virus by complement-fixation. Slovakia virus may be contaminated with mouse hepatitis virus or it may be closely related to mouse hepatitis virus [3].

**Section VI - Biologic Characteristics**

Virus Source (all VERTEBRATE isolates)  
**Blood (LV)**

Lab Methods of Virus Recovery (ALL ISOLATIONS)  
**Newborn mice**

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
Chick embryo(PC)	MB 5		No CPE			No plaques visible	-	
PS (CL)			No CPE			No plaques visible	-	
Vero (CL)			No CPE				-	

**Section VII - Natural Host Range (Additional text can be added below table)**

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Argas persicus	1/406 pools * (6,386 ticks)		Vajnory, Bratislava, Slovakia, CZ

\* A suspension prepared from a single female tick yielded the virus.

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log <sub>10</sub> /ml
Mice (nb)	MB 4	4.5 dex ic 0.01	Death	7.0**	4.5
Mice (nb)		ip			
Mice (nb)		sc 0.05	Nonpathogenic		
Mice (wn)		ic 0.03	No deaths, CF antibody prod.		
Mice (wn)		sc 0.1	No deaths, CF antibody prod.		
rats (nb)		ic 0.1	Nonpathogenic		
rats (nb)		sc 0.2	Nonpathogenic		
rats (nb)		intracut 0.2	Nonpathogenic		
rats (nb)		ip 0.2	Nonpathogenic		
guinea pigs (nb)		ic 0.05	Nonpathogenic		
guinea pigs (nb)		sc 0.01	Nonpathogenic, no CF antibody		
guinea pigs (nb)		ip 0.01	Nonpathogenic, no CF antibody		
guinea pigs (ad)		ip 0.05	Nonpathogenic, no CF antibody		
rabbit (ad)		MB 5	iv 0.1	Nonpathogenic, no CF antibody	
Anas platyrhynchos			4.5 dex sc	Viremia	

\*\* Incubation rather than AST.

**Section IX - Experimental Arthropod Infection and Transmission**

Arthropod species & virus source (a)	Method of Infection log10/ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log10/ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System
Argas persicus, third state nymphs, infected by feeding on viremic Anas platyrhynchos (2)	15 min		2						Suckling mice

**Section X - Histopathology**

Character of lesions (specify host)  
**Meningoencephalitis, focal necrosis of neurons in cortex, basal ganglia, brain stem, extensive edema. Slight cuffing with polynuclear cells, mononuclear leukocytes. Positive immuno- fluorescence in neurons by indirect method**

Inclusion Bodies Intranuclear

Organs/Tissues Affected  
**CNS after ic inoculation**

Category of tropism

**Section XI - Human Disease**

In Nature	Residual	Death
Subclinical	Overt Disease	
Clinical Manifestations		
Number of Cases	Category (i.e. febrile illness, etc.)	

**Section XII - Geographic Distribution**

Known (Virus detected)  
**Czechoslovakia**

Suspected (Antibody only detected)

### Section XIII - References

1. Gresikova, M. et al. 1978. Acta Virol. 23:82-85.
2. Nosek, J. et al. 1978. Z. Parasitenk. 242:141-147.
3. Gresikova, M. Personal communication. 1981.

### Remarks

**Argas persicus 265 was resistant to deoxycholate treatment, and it was protected by 1 M MgCl<sub>2</sub> against thermal inactivation. It persisted at pH 3.0.**