

<b>Virus Name: Toscana</b>		<b>Abbreviation: TOSV</b>
Status <b>Probable Arbovirus</b>	Select Agent <b>No</b>	SALS Level <b>2</b>
SALS Basis <b>Results of SALS surveys and information from the Catalogue.</b>		
Other Information		
Antigenic Group <b>Phlebotomus Fever</b>		

**SECTION I - Full Virus Name and Prototype Number**

Prototype Strain Number / Designation <b>ISS.Phl.3</b>	Accession Number	Original Date Submitted <b>8/31/1984</b>
Family <b>Bunyaviridae</b>	Genus <b>Phlebovirus</b>	
Information From <b>P. Verani et al.</b>	Address <b>Instituto Superiore di Sanita, Viale Regina Elena 299, I-00161 Rome, Italy</b>	
Information Footnote <b>Reviewed by editor</b>		

**Section II - Original Source**

Isolated By (name) <b>P. Verani et al. (1, 5)</b>	Isolated at Institute <b>Instituto Superiore di Sanita, Rome</b>	
Host Genus <b>Phlebotomus perniciosus</b>	Species	Host Age/Stage <b>80 adults</b>
Sex <b>Not Answered</b>		
<u>Isolated From</u>	<u>Isolation Details</u>	
Signs and Symptoms of Illness	Arthropod <b>Depleted</b>	
Time Held Alive before Inoculation <b>A few hours</b>		
Collection Method <b>Caught by hand</b>	Collection Date <b>7/17/1971</b>	
Place Collected (Minimum of City, State, Country) <b>Monte Argentario, Toscana, Central Italy</b>		
Latitude <b>42° 25' N</b>	Longitude <b>11° 10' E</b>	
Macrohabitat <b>Hills surrounding seashore</b>	Microhabitat <b>Animal stables</b>	Method of Storage until Inoculated <b>-70dC</b>
Footnotes		

**Section III - Method of Isolation**

Inoculation Date  
**2/16/1972**

Animal (Details will be in Section 6)  
**nb mice (Tissue Culture)**

Route Inoculated <b>Intracerebral</b>	Reisolation <b>Yes</b>
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Other Reasons

Homologous Antibody Formation by Source Animal

Test(s) Used

Footnotes

**Section IV - Virus Properties**

Physicochemical

Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients(s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

**Stability of Infectivity (effects)**

pH (infective range)

**6 dex of infectivity inactivated at pH 3.0**

Lipid Solvent (ether - % used to test) <b>20%</b>	After Treatment Titer <b>3.2 dex</b>	Control Titer <b>6.6 dex</b>
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate) <b>1:1000</b>	After Treatment Titer <b>&lt;2.5 dex</b>	Control Titer <b>6.6 dex</b>
Other (formalin, radiation)		

**Virion Morphology**

Shape <b>Spherical</b>	Dimensions <b>80 nm</b>	
Mean <b>84nmnm</b>	Range <b>77-97nm</b>	
Measurement Method <b>Electron microscopy of thin sections</b>	Surface Projections/Envelope <b>Enveloped</b>	Nucleocapsid Dimensions, Symmetry

**Morphogenesis**

Site of Constituent Formation in Cell	Site of Virion Assembly	Site of Virion Accumulation
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Inclusion Bodies	Other
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**Hemagglutination**

Hemagglutination <b>Yes</b>	Antigen Source <b>SMB ext. by sucrose-acetone</b>	Erythrocytes (species used) <b>Goose</b>
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pH Range <b>5.75-6.2</b>	pH Optimum <b>6.0</b>
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Temperature Range <b>22dC, 37dC</b>	Temperature Optimum <b>37dC</b>
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Remarks  
**HA production is difficult, usually of poor titer**

Serologic Methods Recommended  
**CF, NT, HI**

Footnotes  
**HA production is difficult, usually of poor titer**

The virus antigen did not react in CF testing with the following ascitic fluids: Icoaraci, Itaporanga, Karimabad, Punta Toro, Sicilian SF, Gabek Forest, but reacted with the ascitic fluid Naples SF. Antigenic relationships between Toscana and Naples SF by CF, HI, and NT (plaque method) tests were noted as follows:

Ascitic fluid	Antigen or Virus					
	CF	Toscana			SFN	
		HI	NT	CF	HI	NT
Toscana	256/128 *	160	640	128/128	80	160
SFN	64/64	40	0	256/128	640	640

\* Antibody titer/antigen titer

At YARU [2], it was shown by CF testing that immune serum to Toscana virus did not react with the antigens of Anhangá, Arumowot, Bujaru, Candiru, Chagres, Icoaraci, Itaporanga, Karimabad, Punta Toro, Sicilian SF, Gabek Forest. It was related to Naples SF by CF.

Cross NT (plaque method) were performed at Pacific Research Unit Hawaii [3] using Toscana virus and hyperimmune ascitic fluids (homologous titer = >5120) against each of the following viruses and specific antisera with negative results: Aguacate, Alenquer, Anhangá, Arumowot, Buenaventura, Bujaru, Cacao, Caimito, Candiru, Chagres, Charleville, Chilibre, Gordil, Tehran, Icoaraci, Itaporanga, Karimabad, Nique, Pacui, Punta Toro, Rift Valley fever (antiserum), Rio Grande, Salehabad, Sicilian SF, Gabek Forest, Urucuri. Antigenic relationships between Toscana, Naples SF, Frijoles, and Saint Floris were noted as follows:

Antiserum or Ascitic Fluid	Virus			
	Toscana	SFN	Frijoles	Saint Floris
Toscana	> 5120	10	40	80
SFN	20	320	40	40
Frijoles	0	0	10240	0
Saint Floris	0	0	0	5120

Virus Source (all VERTEBRATE isolates)

Lab Methods of Virus Recovery (ALL ISOLATIONS)  
Newborn mice

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
Vero (CL)	SMB 4	2	4+	7.0**	5	pinpoint	9.0**	
Vero (CL)	SMB 4 V2	2	4+	7.5				
BHK-21 (CL)	SMB 4	2	4+					
CV-1 (CL)		2	4+					

\*\* Expressed in dex

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Man		76/357 PRNT	Toscana, Italy (4)
Man		2/135 PRNT	Piemonte, Italy
Man		1/67 PRNT	Sicily, Italy
Man		0/120 HI	Lazio, Italy
Sheep		3/75 PRNT	Toscana, Italy
<i>Apodemus sylvaticus</i>		10/119 PRNT	
<i>Phlebotomus perniciosus</i>	3/329 (4 pools)		Monte Argentario, Toscana, Italy
<i>Phlebotomus perniciosus</i> (males)	several		Italy (6)
Man (CSF)	1		Albufeira, Portugal * (7)
<i>Pipistrellus kuhli</i>	1		Tuscany Region, Italy (8)

\* Virus isolated and identified in Sweden.

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log <sub>10</sub> /ml
Mice (nb)	SMB 4	ic 0.01	Death	3*	9.0
Mice (nb)		ip			
Mice (nb)		sc			
Mice (wn)	SMB 3	ic 0.03	Death	3.7	
Mice (wn)		ip 0.05	Death	6.5	
guinea pig		ic 0.10	Paralysis, irregular deaths		
guinea pig		ip 1.0	Antibodies		
rabbit		ic 0.20	Paralysis, few deaths		
rabbit		ip 2.50	Antibodies		
Monkey (Macaca fascicularis)	SMB 4	ic 1.0	Death	9	
Macaca fascicularis		ip 2.0	Antibodies		

\*Incubation rather than AST

**Section IX - Experimental Arthropod Infection and Transmission**

Arthropod species & virus source (a)	Method of Infection log10/ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log10/ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System

Phlebotomus perniciosus fed on blood through chick membranes; 3-14 days post-feeding, 21-51% of flies infected. In one exp., 100 flies exposed to membrane blood meals, then eggs were laid. Virus recovered from 2 3rd instar larvae F1 progeny (6).

**Section X - Histopathology**

Character of lesions (specify host)

Inclusion Bodies

Intranuclear

Organs/Tissues Affected

Category of tropism

**Section XI - Human Disease**

In Nature  
**Reported**

Residual

Death

Subclinical

Overt Disease

Clinical Manifestations

**Severe headache, Slight fever, photophobia; spontaneous recovery.**

Number of Cases  
**> 1**

Category (i.e. febrile illness, etc.)  
**Aseptic meningitis**

**Section XII - Geographic Distribution**

Known (Virus detected)  
**Toscana, Italy**

Suspected (Antibody only detected)

### Section XIII - References

1. Verani, P., et al. 1980. Zbl. Bak. Hyg. Suppl. 9, pp. 147-153.
2. Casals, J. Personal communication.
3. Tesh, R.B. Personal communication.
4. Nicoletti, L., et al. 1980. Zbl. Bak. Hyg. Suppl. 9, pp. 154-159.
5. Verani, P., et al. 1984. Acta Virol. 28:39-47.
6. Ciufolini, M.G., et al. 1985. Am. J. Trop. Med. Hyg. 34:174-179.
7. Ehrnst, A., et al. 1985. Lancet i:1212-1213.
8. Verani, P., et al. 1988. Am. J. Trop. Med. Hyg. 38:433-439.

### Remarks