

Virus Name: Uukuniemi		Abbreviation: UUKV
Status Probable Arbovirus	Select Agent No	SALS Level 2
SALS Basis Results of SALS surveys and information from the Catalogue.		
Other Information		
Antigenic Group Uukuniemi		

SECTION I - Full Virus Name and Prototype Number

Prototype Strain Number / Designation S 23	Accession Number	Original Date Submitted 10/17/1984
Family Bunyaviridae	Genus Uukuvirus	
Information From N. Oker-Blomm and P. Saikku	Address Department of Virology, University of Helsinki, Finland SF 00290	
Information Footnote Reviewed by editor		

Section II - Original Source

Isolated By (name) N. Oker-Blom, et al. (1)	Isolated at Institute Helsinki, Finland	
Host Genus Ixodes ricinus (L.) (pool of 182 engorged specimens)	Species	Host Age/Stage Adult
Sex Female		
<u>Isolated From</u>	<u>Isolation Details</u>	
Signs and Symptoms of Illness	Arthropod Engorged	
Time Held Alive before Inoculation 22 days		
Collection Method Picked off from cows	Collection Date 6/26/1950	
Place Collected (Minimum of City, State, Country) Uukuniemi, SE Finland		
Latitude 61° 48' N	Longitude 30° 3' E	
Macrohabitat Deciduous forest-pasture	Microhabitat On cows	Method of Storage until Inoculated Alive at 22dC
Footnotes		

Section III - Method of Isolation

Inoculation Date
7/18/1960

Animal (Details will be in Section 6)
nb mice (Tissue Culture)

Route Inoculated
Intracerebral

Reisolation
Yes

Other Reasons
Repeated isolations from the same area.

Homologous Antibody Formation by Source Animal

Test(s) Used

Footnotes

Section IV - Virus Properties

Physicochemical
RNA, Single Strand

Pieces (number of genome segments)
3

Infectivity

Sedimentation Coefficients(s)
29; 23; 17 s (7)(S)

Percentage wt, of Virion Protein

Lipid

Carbohydrate

Virion Polypeptides: Number
3

Details

Nucleocapsid (N): 25,000 MW; glycoproteins (G1, G2): 75,000, 65,000 MW (4, 33, 34).

Non-virion Polypeptides: Number

Details

Virion Density
1.20 gm/cc in CsCl

Sedimentation Coefficients(s)
450 s(S)

Nucleocapsid Density
1.31 gm/cc in CsCl (4)

Sedimentation Coefficients(s)
(S)

Stability of Infectivity (effects)

pH (infective range)

Rel. stable at pH 4.6-11.4 (mouse brain suspension)(8)

Lipid Solvent (ether - % used to test)
1:5

After Treatment Titer
1.7 dex

Control Titer
5.6 dex

Lipid Solvent (chloroform)

After Treatment Titer

Control Titer

Lipid Solvent (deoxycholate)
1:1000

After Treatment Titer
1.0 dex

Control Titer
5.5 dex

Other (formalin, radiation)

Not stabilized by MgCl₂; sens. to chloroform, nonionic and ionic detergents (4,8,9)

Virion Morphology

Shape

Dimensions
95 nm

Mean
nm

Range
nm

Measurement Method
Electron microscopy (2-5)

Surface Projections/Envelope
10 nm spikes, hexagonal array (2); lipid envelope (4,6)

Nucleocapsid Dimensions, Symmetry

Morphogenesis

Site of Constituent Formation in Cell

Site of Virion Assembly
Buds through smooth surface cytoplasmic membranes (3)

Site of Virion Accumulation
Cisternae and vesicles of Golgi complex and endoplasmic reticulum

Inclusion Bodies

Other

Hemagglutination

Hemagglutination
Yes

Antigen Source
SMB ext. by sucrose-acetone; BHK-21 culture fluid-serum free

Erythrocytes (species used)
Goose

pH Range
5.6-6.6

pH Optimum
5.6-5.8

Temperature Range
4dC, RT, 3dC

Temperature Optimum

Remarks

HA not readily produced from all strains

Serologic Methods Recommended
CF, HI, NT

Footnotes

HA not readily produced from all strains

Section V - Antigenic Relationship and Lack of Relationship to Other Viruses

Neutralizing indices in suckling mice [1]:

Immune serum	Virus strain		
	S 23	AK 9	Tribec
Uukuniemi (S 23)	2.0 *	0.1	0.0
Tick-borne encephalitis (TBE)	<0.6	1.9	ND
AK 9 (local strain of TBE)	0.4	>2.7	ND
Tribec	<0.9	ND	>1.8
Lymphocytic choriomeningitis	0.0	ND	ND

* LNI in dex.

In Yale Arbovirus Research Unit the Uukuniemi virus was found to be related to some recent isolates (mainly from ticks) which together form the Uukuniemi group.

Results of complement-fixation tests (J. Casals):

Antigen	Immune serum			
	S 23	Potepli	GA	MWA
Uukuniemi S 23	256/128	512/128	8/8	0
Uukuniemi, Potepli	128/256	512/256	8/8	0
Grand Arbaud	16/16	16/16+	256/256	Traces
Manawa	0	0	8/8	512/512

Serum titer/antigen titer; 0 = no fixation at dilutions 1:8.

Results of hemagglutination-inhibition test (J. Casals):

Serum	Antigen, 8 Units					
	UUK	GA	MWA	LJN	BHA	QRF
Uukuniemi	640	40	40	0	0	0
Grand Arbaud	160	160	40	0	0	0
Manawa	80	20	160	0	0	0
Lanjan	0	0	0	80	0	0
Bhanja	0	0	0	0	80	0
Silverwater	0	0	0	0	0	0
Quaranfil	0	0	0	0	0	80
Johnston Atoll	0	0	0	0	0	320

0 = <10

(Ponteves and Zaliv Terpeniya are Uukuniemi group viruses listed in this catalogue not appearing in these tables.)

Section VI - Biologic Characteristics

Virus Source (all VERTEBRATE isolates)

Lab Methods of Virus Recovery (ALL ISOLATIONS)
Newborn mice

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
BHK-21 (CL)	TC, SMB	7	CPE	7.0**				
BHK-21 (CL)					4	Plaques	8.5**	
BS-C-1 (CL)	SMB	14	CPE	7.0				
Chick embryo (PC)	TC, SMB	5	CPE	2.0				
Chick embryo(PC)					4	Plaques	7.5	
Chick embryo(PC)		5		7.0 (a)				
PK (CL)		5	CPE	6.6				
LLC-MK2(CL)						Plaques		
Vero(CL), marten(CL), dormouse kidney(CL)			Partial to no CPE					+

** Expressed in dex

(a) Interference assay

See references 3, 9, 26, 27

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Man		0/191 NT	Kumlinge, SW-Finland (11)
Man		6/100 HI	Czechoslovakia (12)
Man		19/127 HI	Hungary (13)
Cow		0/727 NT	N. Finland (11)
Cow		371/1,809 NT	S., Middle-Finland (11)
Cow		95/100 NT	Uukuniemi, Finland(11)
Cow		84/92 NT	Kumlinge, Finland (11)
Apodemus flavicollis	1	11/288 CF	Czechoslovakia (14, 15)
Apodemus flavicollis	1		Poland (16)
Passerine birds in summer (10 species)	12/736	5/23 HI	SW-, SE-Finland(9, 17) Czechoslovakia (18)
Blackbird, Turdus merula	1		USSR (10)
Ixodes ricinus	12/12,001		SE-, SW-Finland (9)
Ixodes ricinus	39/30,872		Czechoslovakia (19, 20)
Ixodes ricinus	2/3,811		Poland (16)
Ixodes ricinus	3/4,204		USSR (21)
I. ricinus	2		Norway (32)
I. ricinus and I. persulcatus	17		Lithuania (30)
I. Uriae	1 *		Norway (32)

* Uukuniemi-like

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log ₁₀ /ml	
Mice (nb)	SMB 3	ic 0.01	Paralysis, death (9,22-24)	6	7.8	
Mice (nb)		ip 0.03	Paralysis, death (9,22-24)	12	2.6	
Mice (nb)		sc				
Mice (wn)		ic 0.03	No symptoms (9,22-24)			
Mice (wn)		ip 0.1	Antibody production (9,22-24)			
Apodemus flavicollis		ic, sc	No viremia; isolations from organs (14)			
Microtus arvalis (nb)		ic	Death (14)	5-8	7.0	
Chlethrionomys glareolus (ad)		ic	No viremia; isolations from organs (14)			
guinea pig (2 mo)			Antibody formation			
chicken (6 day)		sc	Viremia, antibody formation (9)		3.6	
embryonated egg (6-11 day)		CAM, ys	Death, CAM inflamm.(22)	3-6	7.5 (yolk)	
rhesus monkey		ic	Viremia, meningeal symptoms (25)			
rhesus monkey		ip	Viremia		7.0	

Section IX - Experimental Arthropod Infection and Transmission

Arthropod species & virus source(a)	Method of Infection log10/ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log10/ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System
Ixodes ricinus	Passes transtadially (28); experimental transovarial transmission demonstrated (35).								
Aedes aegypti	Multiplication, transmission (5)								

Section X - Histopathology

Character of lesions (specify host)

Meningoencephalitis, myelitis (14,23) and myositis (24) in suckling rodents; lymphocytary meningitis in rhesus monkeys when inoculated ic (25).

Inclusion Bodies

Intranuclear

Organs/Tissues Affected

Category of tropism

Neurotropic

Section XI - Human Disease

In Nature

Residual

Death

Subclinical

Overt Disease

Clinical Manifestations

Number of Cases

Category (i.e. febrile illness, etc.)

Section XII - Geographic Distribution

Known (Virus detected)

Finland (1), Czechoslovakia (19), Poland (16), USSR (10,21,29), Lithuania (30), Norway (32).

Suspected (Antibody only detected)

Section XIII - References

1. OKER-BLOM, N. et al. 1964. *Ann. Med. Exp. Fenn.* 42:109.
2. SAIKKU, P. et al. 1970. *Acta Virol.* 14:103.
3. BONSDORFF, C.-H. von et al. 1970. *Acta Virol.* 14:109.
4. PETTERSSON, R. et al. 1971. *Virology* 46:721.
5. RAIKOVA, A. et al. 1971. *Vopr. Virusol.* 16:731.
6. RENKONEN, O. et al. 1972. *Virology* 50:899.
7. PETTERSSON, R. and KAARIANEN, L. 1973. *Virology* 56:608-619.
8. KOLMAN, J. 1970. *Acta Virol.* 14:159.
9. SAIKKU, P. and BRUMMER-KORVENKONTIO, M. 1973. *Am. J. Trop. Med. Hyg.* 22:390.
10. GAIDAMOVICH, S. et al. 1971. *Acta Virol.* 15:155. p.
11. SAIKKU, P. 1973. *Am. J. Trop. Med. Hyg.* 22:400.
12. SEKEYOVA, M. et al. 1970. *Folia Parasitol.* 17:341.
13. MOLNAR, E. et al. 1973. *J. Hyg. Epid. Microbiol. Immunol.* 17:1.
14. KOZUCH, O. et al. 1970. *Acta Virol.* 14:163.
15. KOLMAN, J. and HUSOVA, M. 1972. *Folia Parasitol* 19:61.
16. WROBLEWSKA-MULARCZYKOWA, Z. et al. 1970. *Folia Parasitol.* 17:375.
17. SAIKKU, P. 1974. *Med. Biol.* 52:98-103.
18. ERNEK, E. et al. 1971. *Acta Virol.* 15:335. p.
19. KOLMAN, J. et al. 1966. *Acta Virol.* 10:171.
20. KOLMAN, J. and HUSOVA, M. 1971. *Folia Parasitol.* 18:329.
21. GAIDAMOVICH, S. et al. 1971. *Acta Virol.* 15:333.
22. KOLMAN, J. 1970. *Acta Virol.* 14:151.
23. RAJCANI, J. and GRESIKOVA, M. 1970. *Acta Virol.* 14:59.
24. GUSOVSKY, Ya. et al. 1973. *Vopr. Virusol.* 18:167.
25. GRESIKOVA, M. et al. 1970. *Acta Virol.* 14:408.
26. STIM, T. 1969. *J. Gen. Virol.* 5:329.
27. MARHOUL, Z. 1970. *Acta Virol.* 14:249.
28. KOLMAN, J. and MEERGANSOVA, J. 1972. *Folia Parasitol.* 19:4.
29. SAMOILOVA, T. et al. 1973. *Vopr. Virusol.* 18:111.
30. CHUMAKOV, M.P. et al. 1973. *Zh. Mikrobiol. Epidem. Immunobiol.* 5:83-87.
31. THOMAS, L.A. et al. 1973. *J. Med. Entomol.* 10:165.
32. TRAAVIK, T. and MEHL, R. 1977. *Arch. Virol.* 54:317-332.
33. BONSDORFF, C.-H. von and PETTERSSON, R. 1975. *J. Virol.* 16:1296-1307.
34. PETTERSSON, R. et al. 1977. *Cell* 11:51-63.
35. SAMOILOVA, T.I. and DANILOVA, G.M. In: *Ecology of Viruses Associated with Birds.* V.I. Votyakov and D.K. Lvov, editors. Minsk Mater. Resp. Simp. Kamenyuki "Belovezh Puschuchu". 1974. pp. 18-19.

Remarks

First strains were isolated in Finland from ticks collected in June 1959 (Uukuniemi I 1 and Jomala A 21) (1,9). In many experiments Czech. strains (prototype Potepli PO 63) (19) and the Russian geographical variant "Shumakh" (10) have been used.