

<b>Virus Name: Vesicular Stomatitis, New Jersey serotype</b>		<b>Abbreviation: VSNJV</b>
Status <b>Possible Arbovirus</b>	Select Agent <b>No</b>	SALS Level <b>2</b>
SALS Basis <b>Extensive laboratory experience and mild nature of aerosol laboratory infections justifies Level 2.</b>		
Other Information		
Antigenic Group <b>Vesicular Stomatitis</b>		

**SECTION I - Full Virus Name and Prototype Number**

Prototype Strain Number / Designation <b>Hazelhurst</b>	Accession Number	Original Date Submitted <b>7/12/1984</b>
Family <b>Vesiculovirus</b>	Genus	
Information From <b>Robert B. Tesh</b>	Address <b>Yale Arbovirus Research Unit</b>	
Information Footnote <b>Revised</b>		

**Section II - Original Source**

Isolated By (name) <b>L. Mott</b>	Isolated at Institute <b>National Animal Disease Laboratory</b>	
Host Genus <b>suis (domestic pig)</b>	Species	Host Age/Stage <b>adult</b>
Sex <b>Not Answered</b>		
<u>Isolated From</u>	<u>Isolation Details</u>	
<b>Organs/Tissues</b>	<b>snout epithelium</b>	
Signs and Symptoms of Illness <b>vesicle formation on snout, salivation, elevated temperature (1)</b>	Arthropod	
Time Held Alive before Inoculation		
Collection Method	Collection Date <b>5/16/1952</b>	
Place Collected (Minimum of City, State, Country) <b>Hazelhurst, Georgia</b>		
Latitude <b>31° 50' N</b>	Longitude <b>82° 35' W</b>	
Macrohabitat <b>farm, pigs, running into woodland</b>	Microhabitat	Method of Storage until Inoculated <b>ice</b>
Footnotes		

Inoculation Date

Animal (Details will be in Section 6)

**(Embryonated Egg)**

Route Inoculated

Reisolation

Other Reasons

Homologous Antibody Formation by Source Animal**Yes**

Test(s) Used

**NT**

Footnotes

**Section IV - Virus Properties****Physicochemical  
RNA, Single Strand**Pieces (number of genome  
segments)**1**

Infectivity

**no**

Sedimentation Coefficients(s)

**38-45 S(S)**

Percentage wt, of Virion Protein

**60-70%**

Lipid

**20-25%**

Carbohydrate

**3-13% (2, 6); RNA 0.**

Virion Polypeptides: Number

**5**

Details

**L (MW: 150-200 x 10<sup>3</sup>), G (MW: 64 x 10<sup>3</sup>), N (MW: 52 x 10<sup>3</sup>), M (MW: 24 x 10<sup>3</sup>), NS (MW: 29-45 x 10<sup>3</sup>) (3, 6)**

Non-virion Polypeptides: Number

**0**

Details

Virion Density

**1.18-1.20 in sucrose**

Sedimentation Coefficients(s)

**625 S (6)(S)**

Nucleocapsid Density

**1.32 in CsCl**

Sedimentation Coefficients(s)

**140 S (6)(S)****Stability of Infectivity (effects)**

pH (infective range)

**unstable at pH 3; stable in range pH 5-10**Lipid Solvent (ether - % used to  
test)**sensitive**

After Treatment Titer

Control Titer

Lipid Solvent (chloroform)

**sensitive**

After Treatment Titer

Control Titer

Lipid Solvent (deoxycholate)

**sensitive**

After Treatment Titer

Control Titer

Other (formalin, radiation)

**rapidly inactivated by ultraviolet and x-radiation (3)****Virion Morphology**

Shape

**bullet-shaped**

Dimensions

**180 x 75 nm (4, 6)**

Mean

nm

Range

nm

Measurement Method <b>electron microscopy</b>	Surface Projections/Envelope <b>surface projections 6-10 nm; bilayer lipid membrane (3)</b>	Nucleocapsid Dimensions, Symmetry <b>extended; 3.5 nm; helical: 30-35 turns, 49nm outer, 29 nm inner diameter (6)</b>
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**Morphogenesis**

Site of Constituent Formation in Cell <b>cytoplasm</b>	Site of Virion Assembly <b>buds from plasma membrane (3, 4)</b>	Site of Virion Accumulation <b>extracellular and in cytoplasmic vesicles</b>
Inclusion Bodies <b>not usually</b>	Other	

**Hemagglutination**

Hemagglutination <b>Yes</b>	Antigen Source <b>infected BHK-21 cell cultures (6)</b>	Erythrocytes (species used) <b>goose</b>
pH Range	pH Optimum <b>6.4 (5)</b>	
Temperature Range	Temperature Optimum <b>low temperature</b>	

Remarks  
**hemagglutinin prepared from cells maintained in medium containing 0.4% bovine albumin and no serum (5)**

Serologic Methods Recommended  
**CF and NT**

Footnotes  
**hemagglutinin prepared from cells maintained in medium containing 0.4% bovine albumin and no serum (5)**

**Section V - Antigenic Relationship and Lack of Relationship to Other Viruses**

CF tests (6):

Antigens	Hyperimmune Mouse Sera				
	VS-New Jersey	VS-Indiana	Cocal	Piry	Chandipura
VS-New Jersey	256/512 <sup>a</sup>	0	0	0	0
VS-Indiana	0	256/512	32/128	0	0
Cocal	0	32/512	256/512	0	0
Piry	0	0	0	128/32	8/4
Chandipura	0	0	0	0	128/64

<sup>a</sup> Titer of serum/titer of antigen; 0 = <4/4

Log Neutralization Tests (6)

Virus	Hyperimmune Mouse Sera					
	Titer dex LD50	VS-New Jersey	VS-Indiana	Cocal	Piry	Chandipura
VS-New Jersey	5.5	$\geq 5.0^b$	2.2	<1.5	<1.5	<1.5
VS-Indiana	7.1	1.9	$\geq 6.2$	4.2	<1.5	2.6
Cocal	7.7	2.7	4.2	$\geq 7.0$	1.7	1.8
Piry	7.9	2.5	2.3	2.6	5.7	3.4
Chandipura	7.3	<1.5	2.5	2.7	4.6	$\geq 5.7$

<sup>b</sup> Log neutralization index in dex

Plaque Reduction Neutralization Tests (7)

Immune serum	Virus					
	VS-New Jersey	VS-Indiana	Cocal	Piry	Chandipura	Isfahan
VS-New Jersey	10240 <sup>c</sup>	<10	<10	<10	<10	<10
VS-Indiana	<10	327680	320	<10	<10	<10
Cocal	<10	160	5120	<10	<10	<10
Alagoas	<10	20	20	<10	<10	<10
Piry	<10	<10	<10	163840	80	<10
Chandipura	<10	<10	<10	<10	10240	<10
Isfahan	<10	<10	40	<10	<10	163840

<sup>c</sup> Reciprocal of highest serum dilution producing  $\geq 95\%$  plaque inhibition

For additional serologic test results, see Vesicular Stomatitis Alagoas registration card and Reference [6].

## Section VI - Biologic Characteristics

Virus Source (all VERTEBRATE isolates)

Lab Methods of Virus Recovery (ALL ISOLATIONS)  
Weanling mice

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
Vero (CL)		1-2	Total		3	3-4 mm	6.5 °	

VE-New Jersey produces CPE and plaques in a wide variety of mammalian and avian cell lines (80). It also grows in a number of insect cell lines without producing CPE (9, 10, 11).

° Expressed in dex

## Section VII - Natural Host Range (Additional text can be added below table)

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
horse	many	384/611 NT	Panama (12, 13)
cow	many	319/885 NT	
pig	many	94/187 NT	
man (lab workers or animal handlers)	few		USA (14, 15)
man (adult rural inhabitants)		980/2042 NT	Panama (16)
man (adult rural inhabitants)		209/1160 NT	Guatemala (16)
man (adult rural inhabitants)		144/670 NT	Honduras (16)
man (adult rural inhabitants)		146/675 NT	El Salvador (16)
man (adult rural inhabitants)		119/723 NT	Nicaragua (16)
man (adult rural inhabitants)		141/975 NT	Costa Rica (16)

Culex nigripalpus (unfed)	1/95	Guatamala (19)
Mansonia indubitans	1	Ecuador (20)
Musca domestica	22	Larimer City, CO (31)
Musca autumnalis	2	
Chloropidae	1	
Anthomyidae	4	
Simuliidae	2	
Culicoides variipennis	2	Western Colorado (32)
Culicoides stellifer	1	
Culicoides selfia sp.	1	

A single isolation of VSNJ made from Simulium flies collected while feeding on clinically infected cows in Colombia (21).

VSNJ neutralizing antibodies also found in a wide variety of wild mammals (17, 18)

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log <sub>10</sub> /ml
mice (nb)	any except ts	ic 3 dex	death	2-3	6.0-8.0
mice (nb)	mutants	ip	death	2-3	6.0-8.0
mice (nb)		sc	death	2-3	6.0-8.0
mice (wn)		ic	death	2-3	6.0-8.0
mice (wn)		ip	antibody and survival		
hamster (nb)		sc	death (22)	1-2	5.0-9.0
hamster (wn)		sc	antibody		
hamster (ad)		in	death (23)	5-6	
guinea pig (ad)		foot pad	vesicle on foot pad	2-4	4.0-5.0 (fluid)

Many wild mammals susceptible and develop inapparent infection and antibodies following subcutaneous inoculation (24)

**Section IX - Experimental Arthropod Infection and Transmission**

Arthropod species & virus source(a)	Method of Infection log10/ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log10/ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System
Aedes aegypti		1.4	7		mouse	10/10	2.9		plaques, BHK-21 (25)
Culex quinquefasciatus		2.0	9	28			5.4		plaques, Vero (26)
<p>VSNJ multiplies and produces CO2 sensitivity in Aedes albopictus, Culex quinquefasciatus, Toxorhynchites amboinensis and Drosophila melangaster following injection (27, 28). Transovarial transmission of VS NJ could not be demonstrated in Aedes albopictus or Culex quinquefasciatus (26).</p>									

**Section X - Histopathology**

Character of lesions (specify host)  
**spongiosis of the epithelium and multilocular intercellular edema; encephalitis in animals inoculated intracerebrally.**

Inclusion Bodies Intranuclear

Organs/Tissues Affected  
**see reference 30 for discussion of experimental pathology**

Category of tropism  
**epitheliotropic, neurotropic, viscerotropic**

**Section XI - Human Disease**

In Nature Reported	Residual	Death
Subclinical Significant	Overt Disease Significant	
Clinical Manifestations <b>fever, headache, myalgia, arthralgia, prostration</b>		
Number of Cases 30-40	Category (i.e. febrile illness, etc.) <b>febrile illness</b>	

**Section XII - Geographic Distribution**

Known (Virus detected)

Suspected (Antibody only detected)

### Section XIII - References

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### Remarks

**The original strain of VSNJ, isolated by Cotton in 1925 (29), has been lost. The basic epizootic unit considered to be the pasture rather than arthropods (33).**