

<b>Virus Name: Candiru</b>		<b>Abbreviation: CRUV</b>
Status <b>Possible Arbovirus</b>	Select Agent <b>No</b>	SALS Level <b>2</b>
SALS Basis <b>Results of SALS surveys and information from the Catalogue.</b>		
Other Information		
Antigenic Group <b>Phlebotomus Fever</b>		

**SECTION I - Full Virus Name and Prototype Number**

Prototype Strain Number / Designation <b>BeH 22511</b>	Accession Number	Original Date Submitted <b>1/27/1985</b>
Family <b>Bunyaviridae</b>	Genus <b>Phlebovirus</b>	
Information From <b>Belem Virus Lab</b>	Address <b>Belem Virus Laboratory, Instituto Evandro Chagas, Belem, Para, Brazil</b>	
Information Footnote <b>Reviewed by editor</b>		

**Section II - Original Source**

Isolated By (name) <b>Belem Virus Laboratory</b>	Isolated at Institute <b>Belem, Para, Brazil</b>	
Host Genus <b>Man</b>	Species	Host Age/Stage <b>Adult</b>
Sex <b>Male</b>		
<u>Isolated From</u> <b>Serum/Plasma</b>	<u>Isolation Details</u>	
Signs and Symptoms of Illness <b>Fever, headache, dizziness, pain in back, muscles, and joints</b>	Arthropod	
Time Held Alive before Inoculation		
Collection Method <b>Venepuncture</b>	Collection Date <b>8/27/1960</b>	
Place Collected (Minimum of City, State, Country) <b>Km 94, Belem-Brasilia Highway, Brazil</b>		
Latitude <b>2° S</b>	Longitude <b>47° W</b>	
Macrohabitat <b>Primary forest</b>	Microhabitat	Method of Storage until Inoculated <b>Not stored</b>
Footnotes		

**Section III - Method of Isolation**

Inoculation Date  
**8/27/1960**

Animal (Details will be in Section 6)  
**nb mice**

Route Inoculated  
**Intracerebral**

Reisolation  
**Not tried**

Other Reasons

Homologous Antibody Formation by Source Animal  
**Yes**

Test(s) Used  
**HI, NT, HI = 320; NT, LNI = >1.5 dex**

Footnotes

**Section IV - Virus Properties**

**Physicochemical**

Pieces (number of genome segments)	Infectivity	Sedimentation Coefficients (s) (S)
Percentage wt, of Virion Protein	Lipid	Carbohydrate
Virion Polypeptides: Number	Details	
Non-virion Polypeptides: Number	Details	
Virion Density	Sedimentation Coefficients(s) (S)	
Nucleocapsid Density	Sedimentation Coefficients(s) (S)	

**Stability of Infectivity (effects)**

pH (infective range)

Lipid Solvent (ether - % used to test)	After Treatment Titer	Control Titer
Lipid Solvent (chloroform)	After Treatment Titer	Control Titer
Lipid Solvent (deoxycholate) <b>1:1000</b>	After Treatment Titer <b>&lt;2.5 dex</b>	Control Titer <b>5.3 dex</b>
Other (formalin, radiation)		

**Virion Morphology**

Shape	Dimensions	
Mean nm	Range nm	
Measurement Method	Surface Projections/Envelope	Nucleocapsid Dimensions, Symmetry



Candiru	4.4		
Icoaraci	0.6	1.9	2.8
Itaporanga	0.5	<0.2	2.4
Anhanga	0.0	<0.2	1.3
Bujaru		<0.3	
Chagres		1.8	
Sicilian		1.0	

Hyperimmune serum or Antigen	CF titer of Candiru with	CF titer of serum with Candiru antigen	CF titer of serum with homologous antigen
Candiru	64		
Icoaraci	0	0	256
Naples	0	0	256
Sicilian	0	0	256

Mouse ascitic fluids were used instead of sera in some cases; all sera made in mice.

For further information on antigenic relationships, see References [3] - [5].

**Section VI - Biologic Characteristics**

Virus Source (all VERTEBRATE isolates)  
Serum (M)

Lab Methods of Virus Recovery (ALL ISOLATIONS)  
Newborn mice

Cell system (a)	Virus passage history (b)	Evidence of Infection						
		CPE			PLAQUES			Growth Without CPE +/- (g)
		Day (c)	Extent (d)	Titer TCD50/ml (e)	Day (c)	Size (f)	Titer PFU/ml (e)	
Vero (CL)	SMB 10				3-4	1 mm	6.8* (2)	
LLC-MK2 (CL)					3	1 mm	4.9 (2)	

\* Expressed in dex

Vertebrate (species and organ) and arthropod	No. isolations/No. tested	No. with antibody/No. tested Test used	Country and region
Man	1/2,095		Para, Brazil (1)
Man		1/253 HI	Amazon Valley, Brazil
Rodents	0/11,043		Para, Brazil (1)
Marsupials	0/1,950		
Monkeys	0/87		
Bats	0/878		
Edentates	0/127		
Carnivores	0/32		
Horses and cattle	0/185		
Birds	0/6,000	1/805 HI	
Reptiles	0/5,926		
Amphibians	0/42		
Arthropods pools	0/20,758		Amazon region, Brazil

NOTE: Above totals tested were through May, 1964. No further isolation has been made from any source at Belem through December, 1970.

**Section VIII - Susceptibility to Experimental Infection (include viremia)**

Experimental host and age	Passage history and strain	Inoculation Route-Dose	Evidence of infection	AST (days)	Titer log <sub>10</sub> /ml
Mice (nb)	SMB 3	ic 0.02	Death	3.7	7.0
Mice (nb)		ip 0.02	Death	4.8	
Mice (nb)		sc			
Mice (wn)		ic 0.03	Antibody		
Mice (wn)		ip 0.03	Antibody		
hamsters(25 days)	SMB 21		Viremia, HI and CF antibodies		

**Section IX - Experimental Arthropod Infection and Transmission**

Arthropod species & virus source(a)	Method of Infection log <sub>10</sub> /ml (b)		Incubation period (c)		Transmission by bite (d)		Assay of arthropod, log <sub>10</sub> /ml (e)		
	Feeding	Injected	Days	°C	Host	Ratio	Whole	Organ	System
Does not multiply in mosquitoes by inoculation of salivary glands. (Whitman, L. Personal communication).									

**Section X - Histopathology**

Character of lesions (specify host)

**nb mice: hyaline necrosis of isolated hepatocytes or of small groups of same (L.B. Dias)**

Inclusion Bodies

Intranuclear

Organs/Tissues Affected

Category of tropism

**Section XI - Human Disease**

In Nature  
**Reported**

Residual

Death

Subclinical

Overt Disease

Clinical Manifestations

**Fever (R), headache (R), prostration (R), myalgia (R), arthralgia (R), coryza**

Number of Cases  
**One**

Category (i.e. febrile illness, etc.)  
**Febrile illness**

**Section XII - Geographic Distribution**

Known (Virus detected)  
**Brazil**

Suspected (Antibody only detected)

**Section XIII - References**

1. Woodall, J.P. 1967. Atas Simpos. Biota Amazon. 6:31-63.
2. Stim, T.B. 1969. J. Gen. Virol. 5:329-338.
3. Tesh, R.B., et al. 1975. Am. J. Trop. Med. Hyg. 24:135-144.
4. Tesh, R.B., et al. 1982. Ibid. 31:149-155.
5. Tesh, R.B., et al. 1983. Ibid. 32:1164-1171.

**Remarks**