

Laboratory Procedure Manual

Analyte: **Iron**

Matrix: **Refrigerated Serum**

Method: **Beckman UniCel® DxC 800 Synchron &
Beckman UniCel® DxC 660i Synchron Access
Clinical Systems (Identical Method)**

Method No.:

Revised:

As performed by: *Collaborative Laboratory Services, L.L.C*

Contact: Collaborative Laboratory Services
1005 Pennsylvania Suite 102
Ottumwa, IA 52501

Important Information for Users

Collaborative Laboratory Services periodically refines these laboratory methods. It is the responsibility of the user to contact the person listed on the title page of each write-up before using the analytical method to find out whether any changes have been made and what revisions, if any, have been incorporated.

Public Release Data Set Information

This document details the Lab Protocol for testing the items listed in the following table:

Data File Name	Variable Name	SAS Label
BIOPRO_I	LBXSIR	Iron, refrigerated (µg/dL)
	LBDSIRSI	Irons, refrigerated (µmol/L)

1. SUMMARY OF TEST PRINCIPLE AND CLINICAL RELEVANCE

The method used to measure the iron concentration is a timed-endpoint method. In the reaction, iron is released from transferrin by acetic acid and is reduced to the ferrous state by hydroxylamine and thioglycolate. The ferrous ion is immediately complexed with the FerroZine Iron Reagent. The system monitors the change in absorbance at 560 nm at a fixed-time interval. This change in absorbance is directly proportional to the concentration of iron in the sample.

Serum iron measurements in conjunction with total iron binding capacity are useful in the diagnosis and treatment of disorders relating to iron intake, absorption, storage, and release mechanisms. Such changes are indicative of a wide range of dysfunctions including anemias, nephrosis, cirrhosis and hepatitis.

2. SAFETY PRECAUTIONS

Consider all plasma or serum specimens potentially positive for infectious agents including HIV and the hepatitis B virus. We recommend the hepatitis B vaccination series for all analysts working with whole blood and/or plasma. Observe universal precautions; wear protective gloves, laboratory coats. Place disposable plastic, glass, and paper (pipette tips, gloves, etc.) that contact plasma and any residual sample material in a biohazard bag and keep these bags in appropriate containers until disposal by maceration chlorination. Wipe down all work surfaces with Germicidal Disposable Wipe when work is finished.

Handle acids and bases with extreme care; they are caustic and toxic. Handle organic solvents only in a well-ventilated area or, as required, under a chemical fume hood.

Reagents and solvents used in this study include those listed in Section 6. Material safety data sheets (MSDSs) for these chemicals are readily accessible as hard copies in the lab.

3. COMPUTERIZATION; DATA SYSTEM MANAGEMENT

- A. Microsoft Excel software on a PC and our Laboratory Information Systems (L.I.S.) are used to manage the data. The test is analyzed on a Beckman Coulter UniCel[®] Dx^C800 Synchron Clinical System. The Dx^C800 is interfaced to the Laboratory Information Systems (L.I.S.) with a bi-directional interface. After tests are completed, the results will go to the L.I.S. Host Computer Interface to be verified by qualified analyst.
- B. Reflex testing is set up in the L.I.S. to order a repeat of any critical result, to verify abnormal values.
- C. Statistical evaluation of the runs is accomplished with Microsoft Excel software on a PC.
- D. A result file is generated in the L.I.S. database. The file is opened and copied to an Excel spreadsheet for evaluation. The run numbers, and date specimens were received are entered into the Excel file. The Excel spreadsheet results file data are copied to the shipment Excel file and sent using Internet FTP transfer of files or e-mailed to Westat within 21 days of sample receipt.
- E. The Excel files containing all raw data and results are backed up once a week using a CD writer or External drive for storage. Files stored on the L.I.S. network are automatically backed up nightly to tape.
- F. Documentation for data system maintenance is contained in printed copies of data records, as well as in "system log" files on the local hard drives used for the archival of data.

4. SPECIMEN COLLECTION, STORAGE, AND HANDLING PROCEDURES; CRITERIA FOR SPECIMEN REJECTION

A. *Interferences:*

- 1) No interference from lipemia or bilirubin ≤ 30 mg/dL.
- 2) Do not use hemolyzed specimens, if possible.
- 3) Samples with >3+ lipemia should be treated with Lipoclear Clarifying agent prior to analysis (see Lipoclear procedure).
- 4) Serum/plasma should be separated from the red cells within two hours after collection.

- B. Separated serum or plasma should not remain at +15°C to +30°C longer than 8 hours. If assays are not completed within 8 hours, serum or plasma should be stored at +2°C to +8°C. If assays are not completed within 48 hours, or the separated sample is to be stored beyond 48 hours, samples should be frozen at -15°C to -20°C. Frozen samples should be thawed only once. Analyte deterioration may occur in samples that are repeatedly frozen and thawed.
- C. Samples should be drawn in the morning due to diurnal variation.
- D. Iron-dextran administration can cause elevations of iron with this method.
- E. Ingestion of oral contraceptives will elevate iron or TIBC values.
- F. A minimum of 0.6 mL serum is needed for the Multi-Analyte Panel.
- G. Sample volume for individual test is 25 µl added to 210 µl reagent.
- H. Sample is run singly as part of Multi-analyte Biochemistry Panel.
- I. Only plastic or borosilicate glass containers should be used to store samples.

5. PROCEDURES FOR MICROSCOPIC EXAMINATIONS; CRITERIA FOR REJECTION OF INADEQUATELY PREPARED SLIDES

Not applicable for this procedure

6. EQUIPMENT AND INSTRUMENTATION, MATERIALS, REAGENT PREPARATION, CALIBRATORS (STANDARDS), AND CONTROLS

- A. Instrumentation: Beckman Coulter UniCel® DxC 800 & DxC 660i Synchron Clinical Systems
- B. Materials
 - 1) Beckman Micro Tube (*Part #448774*)
 - 2) S/P Plastic Transfer Pipet (*Cat. #P5214-10*)
 - 3) S/P Brand Accutube Flange Caps (*Cat. #T1226-37*)
- C. Reagent Preparation: Beckman Synchron Systems Iron Reagent (*Part #467910*).
 - 1) No preparation required.
 - 2) When stored unopened at 2-8°C, the reagent is stable until the expiration printed on the label.
 - 3) When first opened or installed on the instrument, the reagent is stable for 60 days unless the expiration date is exceeded.
 - 4) Do not freeze.
- D. Standards Preparation: No preparation required.
- E. Synchron Iron/TIBC Calibrator Kit (*Part #442772*).
- F. Control Material
 - 1) Bio-Rad Liquid Unassayed Multiqual level 1 and 3 (Part# 697 and #699).
 - Thaw bottle of control and mix very well.
 - Thawed control is stable 7 days. Mix well prior to each use.
 - Use disposable labware whenever possible. Rinse glassware with 0.1 N HCl before use.

7. CALIBRATION AND CALIBRATION VERIFICATION PROCEDURES

- A. Calibrators: Synchron Iron/TIBC Calibrator Kit (*Part #442772*).
 - 1) Store at ambient temperature.
- B. Calibrators are stable until expiration date on bottle when stored at ambient temperature.
 - 1) The bottles of calibrator solutions should be kept tightly closed, except when drops are being dispensed. Evaporation and/or contamination will result in erroneous assay values.
 - 2) Caution. The calibrator solutions contain HCl, pH<2, which is corrosive. Do not pipette by mouth. Protective clothing, gloves, and safety glasses should be worn when handling this product. Avoid contact with eyes and skin. If spilled, flush with water.
 - 3) Refer to UniCel DxC 600/800 System *Instructions For Use (IFU)* manual for storage and stability information
- C. Calibration:
 - 1) Calibration frequency:
 - 2) Fourteen days for each individual reagent cartridge.
Note: Within-lot calibration is not available for Iron.
 - 3) Refer to UniCel DxC 600/800 System *Instructions For Use (IFU)* manual or *DxC800 and DxC600 Operating Procedure* for programming a calibration.

8. PROCEDURE OPERATING INSTRUCTIONS; CALCULATIONS; INTERPRETATION OF RESULTS

- A. Preliminaries
 - 1) Enter test in L.I.S. as a part of a panel according to procedure listed in this document (See Attachment A).
- B. Sample Preparation
 - 1) Procedure for labeling Micro tube (CX tube) and transferring serum (See Attachment B).
- C. Operation
 - 1) Refer to Operation Procedures for programming controls/patients and loading sectors/racks in the Beckman Coulter Synchron UniCel DxC 600/800 System Instructions for Use (IFU) manual or DxC800 and DxC600 Operating Procedure. (See Attachment C for specific procedure for NHANES samples).
- D. Recording of Data
 - 1) Operator will review and verify results in the L.I.S.
 - 2) The L.I.S. reorders tests to verify any critical results. These results are stored in the L.I.S. along with the original results. Original values are used when repeat results match the original within 3 CV.
 - 3) Project supervisor will export data from the L.I.S. into an Excel file. The data is copied in into another Excel file for further evaluation.
 - 4) An Excel spreadsheet printout of the results for each container ID is made and comments noted.
 - 5) Project supervisor reviews the results. If problems noted with results or QC, Project Supervisor investigates and discusses issues if necessary with Laboratory Director. Repeat samples if necessary.
 - 6) Daily log sheets are completed and any problems or issues noted.
- E. Replacement and Periodic Maintenance of Key Components
(See Attachment D for DxC800 Maintenance Schedule).
- F. Calculations

Synchron Systems perform all calculations internally to produce the final reported result. The system will calculate the final result for sample dilutions made by the operator when the dilution factor is entered into the system during sample programming.

9. REPORTABLE RANGE OF RESULTS

A. Analytical Range:

- 1) 5-500 µg/dL.
- 2) Specimens which are out of analytical range high must be reanalyzed after making a manual dilution of the sample with saline. The dilution factor must be entered at the sample information screen. If the dilution factor is not entered into the system, the instrument print-out must be multiplied by the dilution factor to obtain the final result.
- 3) Limits of detection (LOD) are established by Beckman Coulter and linearity data verifies the reportable range. Detection of results below the reportable range is not relevant and formal limit of detection study is unnecessary.
- 4) Sensitivity is defined as the lowest measurable concentration which can be distinguished from zero with 95% confidence. Sensitivity for the iron determination is 5 µg/dL.
- 5) Samples which are out of instrument range low should be reported as "<5 µg/dL".
- 6) 0 is not a reportable value.
- 7) Repeat all irons <5 µg/dL before reporting.

10. QUALITY CONTROL (QC) PROCEDURES

- A. Blind QC Specimens are included in the samples received from NHANES.
- B. Controls are assayed in early A.M. and if a new reagent pack is loaded, controls are assayed again. One level is assayed in middle of the day and both control levels are assayed after running NHANES sample.
- C. BioRad Liquid Unassayed Multiqual Controls Levels 1 and 3 are assayed for CDC-NHANES runs to allow long term control use. Multiqual controls are analyzed at beginning and end of runs with CDC-NHANES samples.
- D. Acceptable Answer:
 - 1) Controls must be within ± 2 S.D.
 - 2) Refer to Quality Control Flow Chart for action decisions guidelines (*See Attachment F*).

11. REMEDIAL ACTION IF CALIBRATION OR QC SYSTEMS FAIL TO MEET ACCEPTABLE CRITERIA

Remedial action for out of control conditions includes examination of the pipetting and detection equipment and examination of reagent materials. The QC parameters are compared to the patient means to look for confirmatory or disconfirmatory evidence. When the 2 2s and/or 1 3s rules are violated, samples are repeated following corrective maintenance or reagent changes.

12. LIMITATIONS OF METHOD; INTERFERING SUBSTANCES AND CONDITIONS

- A. Do not use hemolyzed sample. Hemoglobin levels of 60 mg/dL (1+) will elevate results by 12 µg/dL.
- B. Bilirubin ≤ 30 mg/dL has no significant interference.
- C. Lipemia has no significant interference.
- D. Copper and magnesium have no significant interference.
- E. Iron-dextran administration can cause elevations of iron with this method.
- F. Ingestion of oral contraceptives will elevate iron or TIBC values.

G. Refer to References for other interferences caused by drugs, disease and preanalytical variables.

13. REFERENCE RANGES (NORMAL VALUES)

Iron

Serum or Plasma Gender Group	Reference Range (µg/dL)
Male	50 to 160
Female	40 to 150

Reference Range values were established from wellness participants with an age mix similar to our patients. These data were analyzed using non-parametric techniques described by Reed (Clin Chem 1971;17:275) and Herrera (J Lab Clin Med 1958;52:34-42) which are summarized in recent editions of Tietz' textbook. Descriptions appear in Clinical Chemistry 1988; 34:1447 and Clinics in Laboratory Medicine June 1993; 13:481.

Pediatric Reference Range Guidelines for Synchron Systems- Multicenter study using data from Montreal, Quebec, Miami, FL and Denver, CO. Beckman 1995

14. CRITICAL CALL RESULTS ("PANIC VALUES")

There are no critical call back values for iron

15. SPECIMEN STORAGE AND HANDLING DURING TESTING

Specimens arrive refrigerated. Specimens are kept refrigerated until ready to transfer to Micro tubes. Capped Micro tubes are kept refrigerated until ready to put on instrument.

Specimen vials are returned to container and refrigerated after transfer of aliquot and double checking of pour off tubes. Specimen vial container is placed in -70°C Freezer after testing is complete. Micro tubes are refrigerated, and then frozen after analysis.

16. ALTERNATE METHODS FOR PERFORMING TEST OR STORING SPECIMENS IF TEST SYSTEM FAILS

Samples will remain in refrigerator until instrument is back in operation.

17. TEST RESULT REPORTING SYSTEM; PROTOCOL FOR REPORTING CRITICAL CALLS (IF APPLICABLE)

The collaborating agency with access to patient identifiers or the responsible medical officer receives an Excel file with all results for a specimen with any critical values. These files with critical values are sent in advance of results that are not abnormal, unless all results are ready to send at the same time. The earliest reporting of results would be the day after arrival of specimens. More frequently two to three days after receiving specimens.

Test results that are not abnormal are reported to the collaborating agency at a frequency and by a method determined by the study coordinator. Generally, data from this analysis are compiled with results from other analyses and sent to the responsible person at the collaborating agency as an Excel file, either through Internet FTP transfer of files or electronic mail or other electronic means.

All data are reported electronically to Westat within 21 days of receipt of specimens.

Internet FTP transfer of files is available and is preferred for data transfer.

18. TRANSFER OR REFERRAL OF SPECIMENS; PROCEDURES FOR SPECIMEN ACCOUNTABILITY AND TRACKING

In general, when specimens are received, the specimen ID number, and a name identifying the container ID and slot number is entered into the Laboratory Information System (L.I.S.) database. New barcodes are printed and the specimens stored in a refrigerator. Samples are aliquoted to a Micro tube with the new barcodes. The specimen ID is read off of the tube by a barcode reader. Tracked in the database are the date and time of entry into the L.I.S., date and time analysis completed, and who certified the results.

Microsoft Excel spreadsheets are used to keep records and track specimens with the data taken from the Laboratory Information System. Logs are kept including information of when samples arrive, are processed and tested, when frozen after testing, and when returned to NHANES for long term storage.

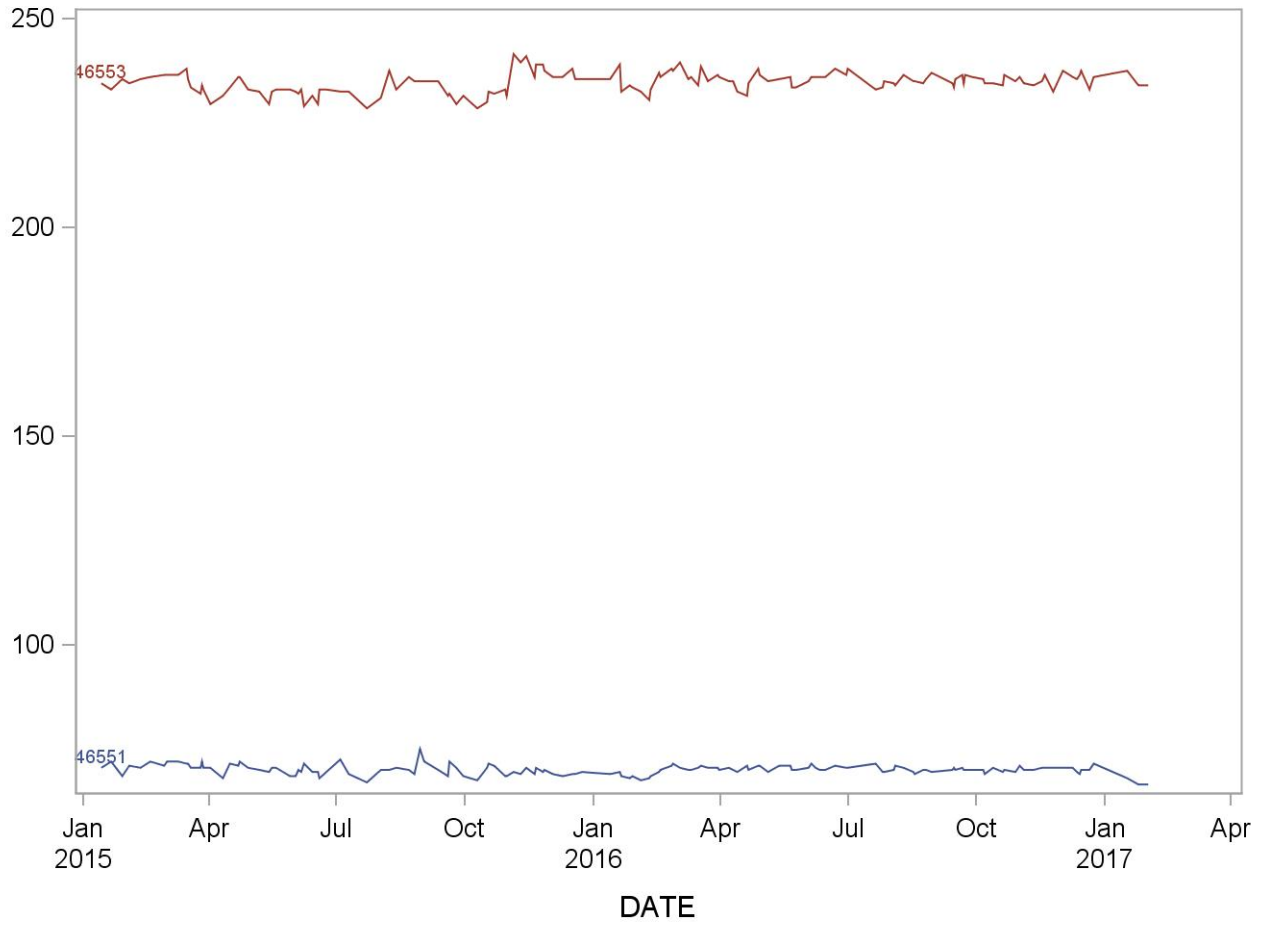
The Project supervisor is responsible for keeping a logbook containing the ID numbers of specimens prepared incorrectly, those with labeling problems, and those with abnormal results, together with information about these discrepancies. It is recommended that records, including related QA/QC data, be maintained for 10 years after completion of the NHANES study.

19. SUMMARY STATISTICS AND QC GRAPHS

See following pages.

2015-2016 Summary Statistics and QC Chart for Iron, refrigerated serum (ug/dL)

Lot	N	Start Date	End Date	Mean	Standard Deviation	Coefficient of Variation
46553	147	14JAN15	01FEB17	234.7	2.4	1.0
46551	147	14JAN15	01FEB17	70.0	1.2	1.7



REFERENCES

- Beckman Coulter Synchron Clinical Systems Chemistry Information Manual, 2007.
- Friedman, R.B. and D.S. Young, Effects of Disease on Clinical Laboratory Tests, 3rd Edition, AACC Press, Washington, D.C. (1997).
- Henry, J.B., ed., Clinical Diagnosis and Management by Laboratory Methods, 18th Edition, W.B. Saunders, Philadelphia, PA (1991).
- National Committee for Clinical Laboratory Standards, How to Define, Determine, and Utilize Reference Intervals in the Clinical Laboratory, Approved Guideline, NCCLS publication C28-A, Villanova, PA (1995).
- National Committee for Clinical Laboratory Standards, Method Comparison and Bias Estimation Using Patient Samples; Approved Guideline, NCCLS publication EP9-A, Villanova, PA (1995).
- National Committee for Clinical Laboratory Standards, Precision Performance of Clinical Chemistry Devices, Tentative Guideline, 2nd Edition, NCCLS publication EP5-T2, Villanova, PA (1992).
- National Committee for Clinical Laboratory Standards, Procedures for the Handling and Processing of Blood Specimens, Approved Guideline, NCCLS publication H18-A, Villanova, PA (1990).
- Tietz, N.W., ed., Clinical Guide to Laboratory Tests, 3rd Edition, W.B. Saunders, Philadelphia, PA (1995).
- Tietz, N.W., ed., Fundamentals of Clinical Chemistry, 3rd Edition, W.B. Saunders, Philadelphia, PA (1987).
- Tietz, N.W., "Specimen Collection and Processing; Sources of Biological Variation," Textbook of Clinical Chemistry, 2nd Edition, W.B. Saunders, Philadelphia, PA (1994).
- Tietz, N.W. Textbook of Clinical Chemistry, W.B. Saunders, Philadelphia, PA (1986).
- Young, D.S., Effects of Drugs on Clinical Laboratory Tests, 4th Edition, AACC Press, Washington, D.C. (1995).
- Young, D.S., Effects of Preanalytical Variables on Clinical Laboratory Tests, 2nd Edition, AACC Press, Washington, D.C. (1997).