Laboratory Procedure Manual

Analyte:	High Sensitivity C-Reactive Protein (hs-CRP)
Matrix:	Serum
Method:	Roche Cobas 6000 (c501 module)
Method No.:	
Revised:	
As portares d bu	
As performed by:	University of Minnesota Advanced Research and Diagnostic Laboratory (ARDL) 1200 Washington Ave S, Suite 175 Minneapolis, MN 55415
Contact:	Anthony Killeen, MD, PhD, Laboratory Director

Jennifer Peters, MT, ASCP, Laboratory Manager

Important Information for Users

The Advanced Research and Diagnostic Laboratory (ARDL) periodically refine these laboratory methods. It is the responsibility of the user to contact the person listed on the title page of each write-up before using the analytical method to find out whether any changes have been made and what revisions, if any, have been incorporated.

Public Release Data Set Information

This document details the Lab Protocol for testing the items listed in the following table:

Data File Name	Variable Name	SAS Label
HSCRP_J	LBXHSCRP	High Sensitivity C-reactive protein (hs-CRP) (mg/L)

1. SUMMARY OF TEST PRINCIPLE AND CLINICAL RELEVANCE

C-reactive protein (CRP) is an acute phase protein, synthesized in the liver, involved in the activation of complement, enhancement of phagocytosis, and detoxification of substances released from damaged tissue. It is one of the most sensitive (albeit nonspecific) indicators of inflammation. CRP levels may rise within six hours of an inflammatory stimulus. Measurement of CRP concentrations by this highly sensitive method is performed primarily to ascertain the level of cardiovascular disease risk in individuals who have no existing inflammatory conditions. Increases in CRP concentration are non-specific and should not be interpreted without a complete medical history.

This is a two-reagent, immunoturbidimetric system. The specimen is first combined with a Tris buffer, then incubated. The second reagent (latex particles coated with mouse anti-human CRP antibodies) is then added. In the presence of circulating CRP the latex particles aggregate, forming immune complexes. These complexes cause an increase in light scattering that is proportional to the CRP concentration. The light absorbance resulting from this light scatter is read against a stored CRP standard curve. The concentration of CRP is determined from this line. Turbidity is measured at a primary wavelength of 546 nm (secondary wavelength 800 nm).

Cobas 6000 Application Code: 217

2. SAFETY PRECAUTIONS

Caution: This product is of human and animal origin. Handle as though capable of transmitting infectious disease. Wear appropriate PPE when handling equipment, reagents, and samples.

3. COMPUTERIZATION; DATA SYSTEM MANAGEMENT

ARDL utilizes a highly specialized Laboratory Information System (LIS) (STARLIMS, Abbott Informatics Corporation; Hollywood, FL, 33021-6755) for all lab functions. Major instrument platforms are interfaced directly to the LIS, allowing data to be electronically transferred directly to the main database. The system provides an extensive quality assurance package and data management tools. Numerous networked computer workstations are used in the laboratory for data management and transmission, and also include software for word and spreadsheet creation and manipulation, statistical analysis, report presentation, and electronic communication. All workstations are user password protected with job specific security access levels and have idle time out functionality. All systems are redundantly backed up on a real time basis.

4. SPECIMEN COLLECTION, STORAGE, AND HANDLING PROCEDURES; CRITERIA FOR SPECIMEN REJECTION

a. **Specimen Type and Requirements:** Serum, lithium heparin plasma, and K2-EDTA plasma are acceptable specimens. The NHANES Biochem study uses frozen serum. This test is analyzed from NHANES Vial 013.

b. **Specimen Volume:** Optimum/Minimum volume: 100 µL in a sample cup or 2 mL microtube (6 µL serum or plasma; remainder for dead volume).

c. **Acceptable Specimens/Unacceptable Specimens:** Serum, lithium heparin plasma, and K2-EDTA plasma are acceptable specimens. Other anticoagulants are not acceptable.

d. **Specimen Stability and Storage:** Separated serum or plasma should be removed from the cells within one hour of collection. Serum or plasma is stable for 11 days at 15-25°C, 2 months at 2-8°C, three years at -15 to -25°C, and longer at –70°C. Specimens must be at room temperature prior to assay.

e. Interferences or limitations: Icteric index < 60: no interference. Hemolytic index <1000: no interference. Lipemic index <600: no interference. Rheumatoid factors up to 1200 IU/mL do not interfere. High dose hook-effect: No false result occurs up to a CRP concentration of 1000 mg/L. Drugs: No interference was found at therapeutic concentrations using common drug panels. Therapeutic drugs: Significantly decreased CRP values may be obtained from samples taken from patients who have been treated with carboxypenicillins. In very rare cases, gammopathy, in particular type IgM (Waldenström's macroglobulinemia), may cause unreliable results. Although measures were taken to minimize interference caused by human anti-mouse antibodies, erroneous findings may be obtained from samples taken from patients who have been

treated with monoclonal mouse antibodies or have received them for diagnostic purposes.

f. **Specimen Handling and Transport:** Mix specimens well, allow clot to fully form (if serum), and centrifuge 10 minutes at 2000 x g before use. Aliquot a minimum of 0.1 mL. Freeze sample until shipment. Ship frozen on dry ice.

5. PROCEDURES FOR MICROSCOPIC EXAMINATIONS; CRITERIA FOR REJECTION OF INADEQUATELY PREPARED SLIDES

Not applicable for this procedure.

6. EQUIPMENT AND INSTRUMENTATION, MATERIALS, REAGENT PREPARATION, CALIBRATORS (STANDARDS), AND CONTROLS

a. Reagents and Supplies

Roche Cat. No. 04628918190, CRPHS reagent kit (300 tests):

b. Reagent Preparation (*Reagents are ready to use; no preparation required)

•R1 reagent. TRIS buffer with bovine serum albumin and immunoglobulins (mouse); preservatives, stabilizers.

•R2 reagent. Latex particles coated with anti-CRP (mouse) in glycine buffer; preservative, stabilizers.

Storage and stability: Keep reagents refrigerated (2-8°C) until use. The reagents are stable for 12 weeks refrigerated on the analyzer.

c. Equipment/Instrumentation-

•Roche Cobas 6000 Chemistry Analyzer (Roche Diagnostics Corporation, Indianapolis, IN 46250)

•The Millipore Elix Gulfstream Clinical 35 System is designed to meet CLSI Clinical Laboratory Reagent Water (CLRW) standards. Water purification is achieved by reverse osmosis, electrodeionization, bactericidal 254 nm UV lamp and 0.22 µm filtration.

d. Specimens are run in singleton

e. Quality Control

- Normal pooled serum control (CQ). Stable at -80°C for up to 4 years, at refrigerated temperature for up to 1 day and at room temperature for up to 4 hours.
- Roche Precipath U Plus Control (catalog #12149443160). Roche Diagnostics 9115 Hague Road Indianapolis, IN 46250-0457). Stable until expiration date on package when unopened and stored at 2-8°C. To prepare, open bottle 1 and pipette in exactly 3.0 mL of diluent (bottle 2). Dissolve by gentle swirling for 30 minutes. Prepared control is stable for 12 hours at room temperature, 5 days at 2-8°C, and one month at -20°C (when frozen once).

7. CALIBRATION AND CALIBRATION VERIFICATION PROCEDURES

Roche C.F.A.S. Proteins Calibrator, catalog # 11355279160: Liquid ready-for-use calibrator based on human serum. Calibrator is stable when unopened and stored at 2-8°C until the expiration date on the bottle. Once opened, the calibrator is stable for 4 weeks when stored at 2-8°C. Calibration frequency: Perform a six-point calibration (H2O + C.F.A.S. Proteins set) when there is a reagent lot number change. The Cobas 6000 will not allow testing to proceed until a successful calibration has been completed. Monitor control values to determine stability of the current calibration. Traceability: This method has been standardized against the reference preparation of the IRMM (Institute for Reference Materials and Measurements) BCR470/CRM470 (RPPHS-Reference Preparation for Proteins in Human Serum).

Manual calibration should be performed if:

- A reagent lot change has not occurred in the past 6 months
- After major service or repairs
- As needed for troubleshooting

If calibration fails perform the following corrective action steps in sequence:

• Check reagent and calibrator for appropriate lot numbers, expiration dates, preparation and storage conditions.

- Repeat calibration with new calibrator.
- Repeat calibration with new reagent and new calibrator
- If successful calibration is not achieved, discontinue testing and notify the supervisor.

8. OPERATING PROCEDURE INSTRUCTIONS; CALCULATIONS; INTERPRETATION OF RESULTS

- a. Instrument Operation: The Roche/Hitachi Cobas 6000 analyzer series is a fully automated, random-access, software controlled system for immunoassay and photometric analyses intended for qualitative and quantitative in vitro determinations using a wide variety of tests. The Cobas 6000 analyzer series is optimized for workloads using a combination of photometric and ion-selective electrode (ISE) determinations (c501 module), and electrochemiluminescence (ECL) signal in the immunoassay analysis module (e601 module). The ISE system is used in the quantitation of sodium, potassium and chloride. The photometric system can measure colorimetric or immunoturbidimetric reactions utilizing end point or kinetic (rate) absorbance measurements. Test ordering end execution on the Cobas 6000 and data entry in the STARLIMS host computer system may be done manually or these tasks may be executed via a barcode-based bi-directional interface. The Cobas 6000 can utilize both of these two systems simultaneously.
- b. **Professional Judgement:** Check results for error flags and take appropriate corrective action. Investigate alert values and delta checks.

c. Result Entry

STARLIMS test code: CRP

Manual entry

- Results are reported to two decimal places, as in x.xx mg/L.
- Report low results as <0.15 mg/L.
- Check results for error flags and take appropriate corrective action.
- Investigate alert values and delta checks.

9. REPORTABLE RANGE OF RESULTS

Out of Range results: Certain tests have pre-programmed limits that trigger an automatic re-analysis by the COBAS. These limits may be low-end values or high-end values (but within technical range). If the duplicate value is in agreement with the initial value, then the initial value is reported. CRP values <0.15 mg/L are automatically repeated by the instrument.

Results are reported to two decimal places (as x.xx) in mg/L. Report low results as <0.15 mg/L.

a. Reportable Range of Test Results: Reportable Range 0.15-300 mg/L Intra-assay %CV (10 within-day replicates at a concentration of 3.44 mg/L) 4.0% Intra-assay %CV (10 within-day replicates at a concentration of 9.14 mg/L) 0.9% Inter-assay %CV (between day replicates at a concentration of 3.06 mg/L) 6.8% Inter-assay %CV (between day replicates at a concentration of 1.00 mg/L) 3.8% Dilutions: The confirmed analytical measurement range of the assay is 0.15-20.0 mg/L (serum). Specimens exceeding the high limit are automatically diluted (1:15) by the instrument. Results from samples diluted using the rerun function are automatically multiplied by a factor of 15. If a manual dilution is required, dilute the specimen in normal saline, and multiply the result by the dilution factor. For example, to perform a 1:5 dilution, pipette 50 μ L of the patient sample into 200 μ L of normal saline. Mix thoroughly, perform the assay, and multiply the result by a factor of 5. The maximum allowable dilution is 1:15.

Reference Range: Serum, adult: <5 mg/L

Critical Results: None

Analytical Measurement Range: 0.15-20 mg/L

Reportable Range: 0.15-300 mg/L

Limit of Detection (standard 1 + 3 SD): 0.15 mg/L

10. QUALITY CONTROL (QC) PROCEDURE

•Normal pooled serum control (CQ). Stable at -80°C for up to 4 years, at refrigerated temperature for up to 1 day and at room temperature for up to 4 hours.

•Roche Precipath U Plus Control (catalog #12149443160). Roche Diagnostics 9115 Hague Road Indianapolis, IN 46250-0457). Stable until expiration date on package when unopened and stored at 2-8°C. To prepare, open bottle 1 and pipette in exactly 3.0 mL of diluent (bottle 2). Dissolve by gentle swirling for 30 minutes. Prepared control is stable for 12 hours at room temperature, 5 days at 2-8°C, and one month at -20°C (when frozen once).

•Both levels of quality control are analyzed at the start of the day and results are verified for acceptability prior to testing specimens. Quality control is also analyzed at the end of the shift, with change in reagent, after major maintenance, or as needed for troubleshooting.

•The analytical measurement range (AMR) must be validated every 6 months or after major maintenance or service procedures. The laboratory enrolls in the College of American Pathologist (CAP) linearity program. C-reactive protein, high sensitivity is included in the LN21 kit that is shipped twice per year. Follow kit instructions for preparation. Analyze samples in duplicate. Results are due within two to four weeks of receipt of kit. Results are submitted online to the CAP website by the lead or supervisor. The linearity report is available online at the CAP website shortly after the due date. Confirm reported values are within acceptability limits. Place instrument printouts, worksheets and CAP results in the CAP three ring binder.

•New Lot Verification: Each new reagent lot must be verified for acceptability before being placed into use. Calibration, quality control, and comparison of at least 5 patient samples on the old and new lots must be performed and found to be within acceptable limits before a new lot can be placed into use.

11. REMEDIAL ACTION IF CALIBRATION OR QC SYSTEMS FAIL TO MEET ACCEPTABLE CRITERIA

• If QC values are outside of specified ranges, do the following, in order, until QC is acceptable:

- 1. Repeat the analysis using fresh QC material.
- 2. Perform a calibration.
- 3. Check for system problems.

4. Contact Roche Technical Support for assistance and possible service dispatch. Phone: 1-800-428-2336; account number: 55042919

12. LIMITATIONS OF METHOD; INTERFERING SUBSTANCES AND CONDITIONS

- a. Limit of Detection (standard 1 + 3 SD): 0.15 mg/L
- b. Analytical Measurement Range: 0.15-20.0 mg/L
- c. Icteric index < 60: no interference. Hemolytic index <1000: no interference. Lipemic index <600: no interference. Rheumatoid factors up to 1200 IU/mL do not interfere. High dose hook-effect: No false result occurs up to a CRP concentration of 1000 mg/L. Drugs: No interference was found at therapeutic concentrations using common drug panels. Therapeutic drugs: Significantly decreased CRP values may be obtained from samples taken from patients who have been treated with carboxypenicillins. In very rare cases, gammopathy, in particular type IgM (Waldenström's macroglobulinemia), may cause unreliable results. Although measures were taken to minimize interference caused by human anti-mouse antibodies, erroneous findings may be obtained from samples taken from patients who have been treated with monoclonal mouse antibodies or have received them for diagnostic purposes.</p>

13. REFERENCE RANGES (NORMAL VALUES)

Serum, adult: <5 mg/L

14. CRITICAL CALL RESULTS ("PANIC VALUES")

Not applicable.

15. SPECIMEN STORAGE AND HANDLING DURING TESTING

Specimens are frozen at -70°C between sample receipt and analysis on the instrument. Specimens must be at room temperature prior to assay. Specimens are returned to refrigerated or frozen temperature post analysis depending on the study specific requirements.

16. ALTERNATE METHODS FOR PERFORMING TEST OR STORING SPECIMENS IF TEST SYSTEM FAILS

Should the testing system become inoperable, discontinue testing and notify the supervisor. While instrument trouble-shooting or repair occurs; keep specimens at refrigerated or frozen temperature depending on study specific requirements.

17. TEST RESULT REPORTING SYSTEM; PROTOCOL FOR REPORTING CRITICAL CALLS (IF APPLICABLE)

All data is reported electronically via an eFile that is uploaded to the WESTAT secure website within 21 days of receipt of specimens.

18. TRANSFER OR REFERRAL OF SPECIMENS; PROCEDURES FOR SPECIMEN ACCOUNTABILITY AND TRACKING

Specimen Receipt:

Shipments for NHANES generally will arrive on Tuesdays and/or Wednesdays. These shipments are recorded on the Log of Quality Assurance located in a binder labeled NHANES Shipping Log in the receiving area. The specimen barcode numbers in the boxes are checked against the manifests. The receipt date is written on top of the boxes. The frozen samples (vial 11-Iron/UIBC & vial 13-CRP) are placed in the designated -70°C freezer and the refrigerated samples (vial 18-Biochem panel) are placed in the designated 2-8°C refrigerator until analysis. The manifests are filed in a binder labeled NHANES Shipping Manifests located in the receiving area. All labels are removed from the shipping box and the provided airbill is attached for return shipment.

Quality Assurance Log:

A Quality Assurance Specimen Receipt and Specimen Return Log is maintained by laboratory staff. The following parameters are tracked: NHANES shipper I.D., NHANES Container I.D., Vial #, Date Received, Specimen Receipt Conditions, Number of Specimens Received, 2.5% QC Repeats, Total Number of Specimens, 21 Day Due Date, Analysis Date, Date Results Sent, Number of Days For Result Return, Thaw Date (if applicable), Return To Freezer Date, Number of Days at Refrigerated Temperature, 1 Year Discard or Return Date, NHANES Quarterly Report Date

QUALITY ASSUR SPECIMEN RECEIPT					# received, a		
NHANES NHANES				receive	and transmitted		
shipper	container		date	spec.		2.5%	
I.D.	I.D. #	vial #	rec'd	cond.	spec.	qc rpts	total
778177102370	368009	018	1/17/2017	ok	47	1	48
778204066788	368058	018	1/24/2017	ok	44	1	45
778299222501	368100	018	1/31/2017	ok	41	1	42
778354403016	368143	018	2/7/2017	ok	25	1	26
778355410426	369009	018	2/7/2017	ok	42	1	43
778408602420	369092	018	2/14/2017	ok	54	1	55
778407805716	368181	018	2/14/2017	ok	22	1	23
778454414575	368221	018	2/21/2017	ok	21	1	22
778463664926	369135	018	2/21/2017	ok	44	1	45
778511320887	369176	018	2/28/2017	ok	29	1	30
778517067017	370009	018	2/28/2017	ok	44	1	45
778564705940	370058	018	3/7/2017	ok	64	1	65

		date	# days		return to	# days	discard	NHANES
21-day	analyzed	result	results	thaw	freezer	at frig	or return	Quarter
due date	date	sent	return	date	date	temp	date	Report
2/7/2017	1/18/2017	1/30/2017	13	NA	1/20/2017	2		
2/14/2017	1/25/2017	2/6/2017	13	NA	1/27/2017	2		
2/21/2017	2/1/2017	2/6/2017	6	NA	2/2/2017	1		
2/28/2017	2/8/2017	2/10/2017	3	NA	2/10/2017	2		
2/28/2017	2/8/2017	2/10/2017	3	NA	2/10/2017	2		
3/7/2017	2/15/2017	2/17/2017	3	NA	2/16/2017	1		
3/7/2017	2/15/2017	2/17/2017	3	NA	2/16/2017	1		
3/14/2017	2/22/2017	2/27/2017	6	NA	2/24/2017	2		
3/14/2017	2/22/2017	2/27/2017	6	NA	2/24/2017	2		
3/21/2017	3/1/2017	3/3/2017	3	NA	3/3/2017	2		
3/21/2017	3/1/2017	3/3/2017	3	NA	3/3/2017	2		
3/28/2017	3/8/2017	3/10/2017	3	NA	3/10/2017	2		

Specimen Ordering/Labeling:

Electronic files for all NHANES specimens are sent via email from Westat, Inc to the NHANES contact person shortly before they are to be received. These files include the Sample ID, Analyte Type, Slot No, Sample Collection Date, Sample Comment, Age Grouping, Astro ID, Receipt Date, Analysis Date, Run Number, Tech ID, Analyte Result, Result Comment, Adjusted Result, QC Repeat, LOD, Change Reason, and Change Reason Other. The first seven columns are protected and cannot be altered. The files are saved on the laboratory's common S drive in the NHANES Biochem folder. After analysis, the contact person returns the completed files via their website to Westat, Inc.

The NHANES spreadsheets are used to set up pending batches for batch accession upload in the Laboratory Information system (STARLIMs). New labels are generated out of the Laboratory Information System (STARLIMs). The new bar-coded labels are attached to a carrier tube. The Cobas analyzer reads the bar-coded label for the sample ID and test information.

Specimen Storage:

The temperatures for all freezers and refrigerators are monitored 24 hours a day/ 7 days a week. If the temperature for any unit falls outside the allowable range, action is taken to resolve the problem. If the temperature cannot be corrected, the contents are moved to a different unit.

Specimen Handling/Specimen Return:

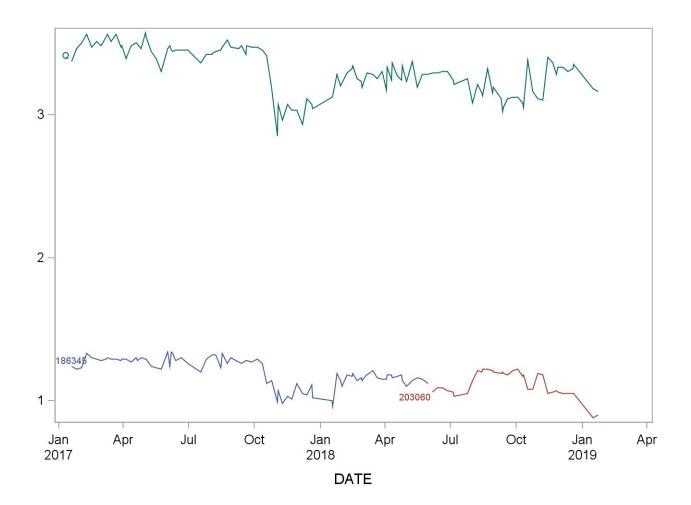
Prior to analysis, the specimens are stored in the designated -70°C freezer. On the day of analysis, the specimens are selected and thawed by the technician operating the COBAS. After analysis and the QC repeats have been run, the specimens are refrozen. After 1 year, the specimen vials that have at least 0.2ml of sample remaining will be shipped to SriSai Biopharmaceuticals in Frederick, MD. These specimens will be shipped on dry ice via Federal Express.

19. SUMMARY STATISTICS AND QC GRAPHS

See following page.

Lot	N	Start Date	End Date	Mean	Standard Deviation	Coefficient of Variation
Q	122	19JAN17	23JAN19	3.303	0.161	4.9
186345	86	19JAN17	31MAY18	1.205	0.096	8.0
203060	36	06JUN18	23JAN19	1.115	0.088	7.9

Summary Statistics and QC Chart for HS C-Reactive Protein (mg/L)



REFERENCES

- Cobas 6000 Analyzer Operator's Manual. Roche Diagnostics Inc., 9115 Hague Road Indianapolis, IN 46250-0457. Version 5.0.
- Roche Calibrator for Automated Systems, Proteins (C.F.A.S. Proteins) package insert. Version 2017-09, V6.0. Roche Diagnostics Inc., 9115 Hague Road Indianapolis, IN 46250-0457.
- 3. Roche CRPHS reagent package insert. Version 2017-05, V10.0. Roche Diagnostics Inc., 9115 Hague Road Indianapolis, IN 46250-0457.
- Roche Precipath U Plus package insert, Version 2011-04, V4. Roche Diagnostics Inc., 9115 Hague Road Indianapolis, IN 46250-0457.